

Biodiversity on Seamounts in Aotearoa

Katrina Goddard, October 2021

A report on the importance of the biodiversity found on seamounts and other similar features within Aotearoa and the Pacific.



IMAGE: CSIRO, Bottlebrush gold corals *Chrysogorgia* on *Solenosmilia* reef, Tasmania 2018

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Why seamount protection is critical

1. Seamounts are home to disproportionate amounts of biodiversity

Seamounts are underwater mountains that rise more than 100m off the seafloor¹ and are regarded globally as hotspots of marine biodiversity because they are home to many diverse, often unique, and endemic species or communities². Only a fraction of Aotearoa's seamounts and associated biodiversity has been sampled or surveyed, and what is known is that Aotearoa seamounts can host abundant and diverse vulnerable marine ecosystems (VME) indicator species³. VME indicator species include rare and unique species, species that are functionally significant to the health of marine habitats and species which are particularly complex, slow growing or long lived⁴. Seamounts are important to migratory pelagic species like whales⁵, sharks and even tuna⁶. There is international evidence that near sea-surface seamounts can also be important feeding grounds for different seabird species⁷.

2. Thriving biodiversity underpins overall ocean health

The deep-sea constitutes the largest source of species and ecosystem diversity on Earth⁸ as well as supporting diverse ecosystem processes and functions necessary for the Earth's natural systems to operate. Thriving healthy oceans including deep-sea ecosystems associated with seamounts and similar features build resilience against climate change⁹. The ocean is an important carbon store as it captures CO₂ from the atmosphere¹⁰, absorbing around 25% of all CO₂ emissions¹¹ and then storing it.

¹ Staudigel et al. 2010

² Ministry for the Environment and Statistics NZ, 2019

³ NIWA, 2016

⁴ Watling & Auster, 2021

⁵ Garrigue et al. 2015

⁶ Morato et al. 2010

⁷ Iyer et al. 2012

⁸ UNESCO, 2015

⁹ Sweetman et al. 2017

¹⁰ Bijma et al. 2013

¹¹ Watson et al. 2020

3. Healthy seamounts are critical for the long-term sustainability of fisheries

Seamounts are highly productive due to their physical hard structure providing substrate for species to grow on and live amongst¹², and level of nutrient rich upwellings which support high levels of plankton productivity which attracts a range of biodiversity^{13,14}. Upwelled nutrients from the deep-sea fuel photosynthesis supporting major fisheries such as anchovies¹⁵ and tuna¹⁶. Seamounts are also critical habitats, such as sites of spawning aggregations for some deep-sea fish species like orange roughy¹⁷.

4. Damage done to seamounts is essentially irreversible

Seamount biodiversity, like protected corals are slow-growing and highly fragile making them highly vulnerable to bottom trawling especially given the core depth distributions of most protected corals¹⁸ overlaps with the main 'fishable' depths. As few as ten individual bottom trawls can reduce deep-sea coral forests to rubble¹⁹. Research has shown that recovery of some deep-sea coral vulnerable marine ecosystems from the impacts of benthic impacting fishing methods is possible but will take several decades to centuries²⁰.

5. Existing protected areas do not adequately protect seamounts and associated biodiversity

Seventy two percent of seamounts (>100m) and similar features are unprotected and vulnerable to benthic fishing methods²¹. No type 1 MPAs (marine reserves) can be established beyond the territorial sea (12 nm) into the EEZ where the majority of seamounts occur. Existing benthic protected areas (BPAs) provide little protection as much of the seabed and features like seamounts are well beyond the current trawlable depths that extend to around 1,600 meters²², and also the core-depth distribution of VME taxa, such as protected corals²³.

6. Bottom trawling is expanding into new unfished areas both in Aotearoa and beyond

While the commercial trawl footprint comparatively to a decade ago is smaller, new areas are still being trawled²⁴. Within the SPRFMO area New Zealand is the only country currently bottom trawling on seamounts. New Zealand is still reporting high bycatch rates of VME indicator

¹² Schlacher et al. 2014

¹³ Annasawmy et al. 2020

¹⁴ Demarcq et al. 2020

¹⁵ Le & Sato, 2017

¹⁶ Morato, et al. 2010

¹⁷ Bull et al. 2001

¹⁸ Tracey & Hjørvarsdóttir, 2019

¹⁹ Clark et al. 2010

²⁰ Baco et al. 2019

²¹ Fisheries New Zealand, 2020

²² Maximum trawlable depth is about 1,600 m, from Tracey, 2018.

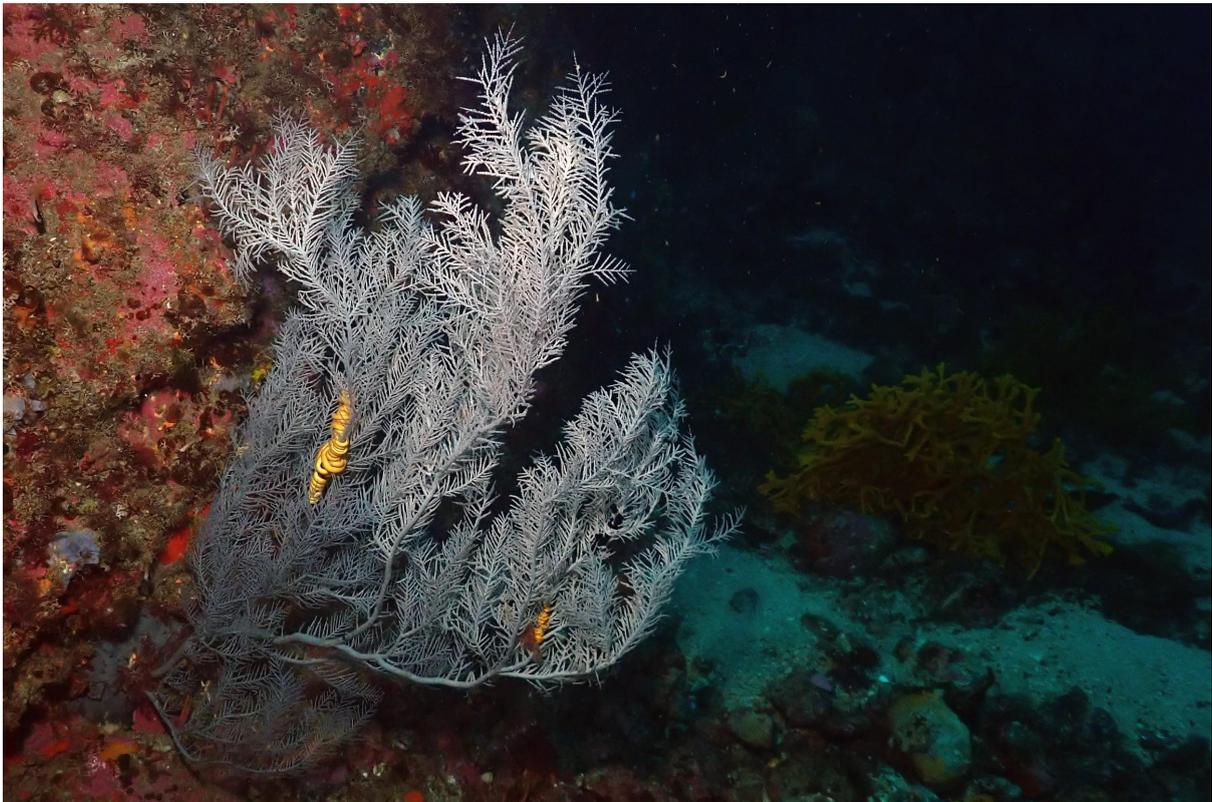
²³ Tracey & Hjørvarsdóttir, 2019

²⁴ WWF-NZ, 2021

species to SPRFMO. Given scientists are discovering new deep-sea invertebrates from areas fished and unfished, it means there is the potential for New Zealand vessels to destroy endemic or rare invertebrate species before they can be discovered and identified.

7. Banning bottom trawling on seamounts is essential

Protecting all seamounts and similar features from all benthic impacting fishing methods will ensure Aotearoa's vulnerable marine ecosystems such as protected endemic, rare and diverse deep-sea corals not only survive but thrive. Protecting VMEs would also go some way towards New Zealand meeting its international obligations, such as United Nations General Assembly resolutions and domestic policies and legislation.



© Andy Stewart. Black coral, Hen & Chickens Islands, Northland New Zealand, 2021

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Section 1: Seamounts and similar features

In 2015 the first World Ocean Assessment stated that the deep-sea constitutes the largest source of species and ecosystem diversity on Earth²⁵. The deep-sea also supports the diverse ecosystem processes and functions necessary for the Earth's natural systems to operate, yet despite this and the importance of the deep, only a tiny fraction of the deep-sea has ever been studied²⁶.

What are Seamounts and similar features?

Seamounts are undersea mountains often referred to as deep-sea biological 'hotspots' as they are highly productive areas²⁷. Similar features include hydrothermal vents, which are underwater hot springs found on the ocean seafloor, often referred to as 'black smokers' or 'white smokers' depending on the minerals they produce. Other similar underwater topographic features are hills, knolls and guyots.

Despite there being no universally agreed definition of the term 'seamounts', seamounts are widely defined as undersea mountains that rise more than 100m from the surrounding seafloor^{28, 29}. The Food and Agriculture Organisation of the United Nations (FAO) *generally*³⁰ defines seamounts as having an elevation of greater than 1,000m from the seafloor³¹. However, researchers have shown that the physical characteristics of seamounts and similar features less than 1,000m can affect biodiversity in similar ways to those greater than 1,000m above the seafloor^{32,33,34}.

In Aotearoa, the Department of Conservation (DOC) and Fisheries New Zealand (FNZ) agree that seamounts and similar features (including knolls and hills) are those that rise more than 100m above the seafloor³⁵. In addition to Government agencies, the 100m elevation definition is widely used by New Zealand scientists at NIWA³⁶ and in international scientific publications. If the 1,000m definition was used in Aotearoa over the 100m then many seamounts and similar features would be excluded³⁷, such as the well studied Northwest hills seamount complex on the Chatham Rise. NIWA scientists have shown in Aotearoa it's not the height of the seamount, but the depth from the surface that influences the abundance and diversity of endemic, rare and vulnerable invertebrates like corals and sponges, as many protected corals have core depth

²⁵ UNESCO, 2015

²⁶ UNESCO, 2015

²⁷ Ministry for the Environment and Statistics NZ, 2019

²⁸ Staudigel et al. 2010 defined seamounts as: "*any geographically isolated topographic feature on the seafloor taller than 100 m, including ones whose summit regions may temporarily emerge above sea level, but not including features that are located on continental shelves or that are part of other major landmasses*" See also Rogers, 1994 & Rogers, 2019.

²⁹ Kim, 2014

³⁰ emphasis added

³¹ FAO seamount definition: "*A large isolated elevation characteristically of conical form. Seamounts are under-sea mountains whose summits lie beneath the ocean surface. They are usually volcanic in origin and are **generally** defined as having an elevation of greater than 1 000 metres from the seabed*" from Cochrane, 2002 [emphasis added].

³² Rogers, 2018

³³ Lyer et al. 2012

³⁴ Tracey & Hjørvarsdóttir, 2019

³⁵ Department of Conservation & Fisheries New Zealand, 2021

³⁶ NIWA - National Institute of Water and Atmospheric Research is a New Zealand Crown Research Institute established in 1992

³⁷ Like the number of seamounts described by Rowden & Clark, 2004

distribution ranges of between 200 - 1,000m deep³⁸, meaning many protected corals are unable to live on the deeper seamounts, even those that have elevations over 1,000m as they are simply too deep.

Seamounts are usually volcanic in origin³⁹ and tend to occur in groups or chains but can also be isolated. Generally, most are found near convergent tectonic plates in areas of vertical movement⁴⁰. Seamounts can vary significantly in height, from hundreds of meters to several kilometres in elevation off the seafloor⁴¹. Typically, seamounts have a steep outer slope, a near flat circular summit and often collapsed features such as calderas or pit craters^{42,43}. Figure 1 shows a typical seamount from Aotearoa mapped using underwater seabed mapping technology by NIWA.

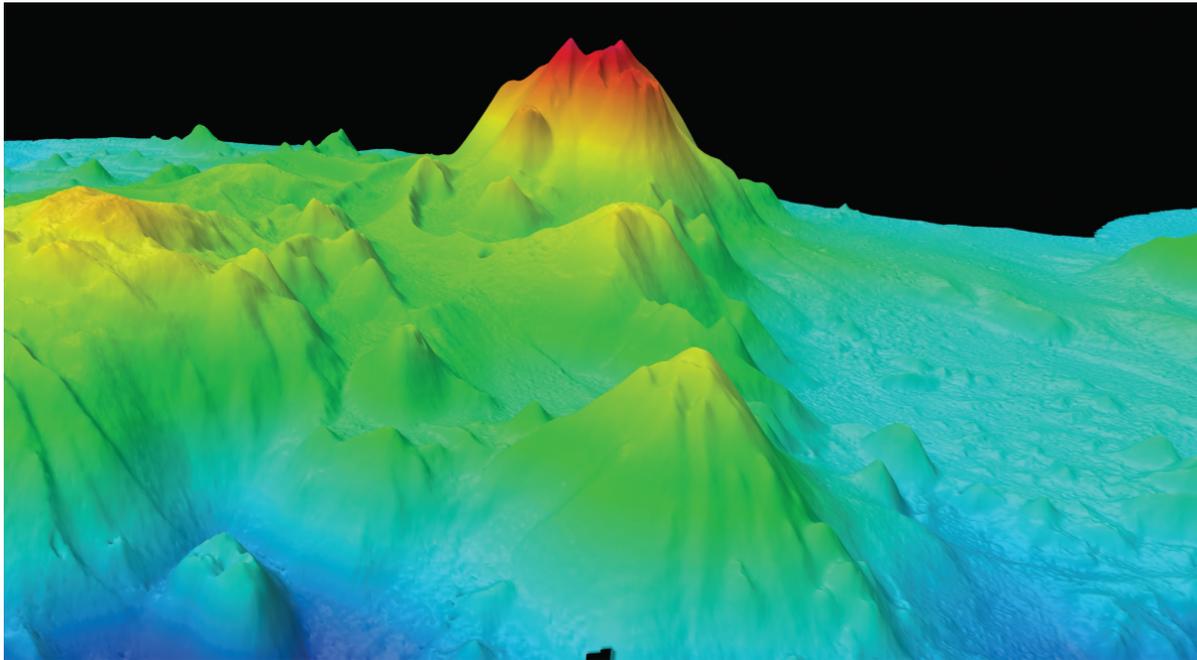


Figure 1: Hinepua Seamount on the Kermadec Arc. Credit: NIWA⁴⁴

Seamount Biodiversity & Productivity

Seamounts are regarded globally as hotspots of marine biodiversity because they are home to many diverse, often unique, and endemic species or communities⁴⁵. These include corals, sponges and other sessile epifauna organisms⁴⁶. It is estimated that only 0.002% of global seamounts have been sampled adequately, meaning our knowledge of seamount biological communities is very limited and it is highly likely many benthic communities and species are yet to be discovered⁴⁷.

³⁸ Tracey & Hjørvarsdóttir, 2019

³⁹ Rogers, 2018

⁴⁰ Rogers, 2018.

⁴¹ Lyer et al. 2012

⁴² Lyer et al. 2012

⁴³ Rogers, 2018

⁴⁴ NIWA, 2013 [Here be dragons | NIWA](#)

⁴⁵ Ministry for the Environment and Statistics NZ, 2019

⁴⁶ Schlacher et al. 2014

⁴⁷ Rogers, 2018

Seamounts are highly productive due to their physical hard structure providing substrate for species to grow on and live amongst⁴⁸, disruption of ocean currents, and level of nutrient rich upwellings which support high levels of plankton productivity which attracts a range of biodiversity^{49,50}. Seamounts are important to migratory pelagic species like whales, sharks and even tuna⁵¹. Seamounts are also critical habitats, such as sites of spawning aggregations for some deep-sea fish species like orange roughy⁵² and even deep-sea eels, like basketwork⁵³.

Satellite tracking of endangered South Pacific humpback whales highlights that seamounts are not only important during migrations, but also during the breeding season, as tagged whales used oceanic seamounts frequently during both⁵⁴. This study described the multiple ecologically important roles seamounts played – from breeding locations, resting areas, navigational landmarks and even likely supplemental feeding grounds⁵⁵.

There is international evidence that near sea-surface seamounts can be important feeding grounds for different seabird species, as areas surrounding these near surface seamounts have higher densities of seabirds than adjacent waters⁵⁶.

Upwelled nutrients from the deep-sea fuel photosynthesis supporting major fisheries such as anchovies⁵⁷ and tuna⁵⁸. But deep-sea ecosystems don't only support extractive services like fisheries, they also play an important role in non-extractive services such as climate regulation⁵⁹.

Thriving healthy oceans including deep-sea ecosystems associated with seamounts and similar features build resilience against climate change⁶⁰. The ocean is an important carbon store as it captures CO₂ from the atmosphere⁶¹, absorbing around 25% of all CO₂ emissions⁶² and then stores it. Carbon sequestration (the physical process of sinking biologically produced carbon from the upper ocean into the deep sea) is an important biological climate change mitigation pathway as it can store the CO₂ from the atmosphere⁶³ for thousands to millions of years⁶⁴.

Anthropogenic CO₂ emissions have resulted in ocean warming, deoxygenation, and acidification^{65,66}. Research is limited looking at how global climate change will affect carbon sequestration and deep-sea ecosystems and their associated biodiversity such as reproduction, distribution, and

⁴⁸ Schlacher et al. 2014

⁴⁹ Annasawmy et al. 2020

⁵⁰ Demarcq et al. 2020

⁵¹ Morato et al. 2010

⁵² Bull et al. 2001

⁵³ Williams et al. 2021

⁵⁴ Garrigue et al. 2015

⁵⁵ Garrigue et al. 2015

⁵⁶ Iyer et al. 2012

⁵⁷ Le & Sato, 2017

⁵⁸ Morato, et al. 2010

⁵⁹ Le & Sato, 2017

⁶⁰ Sweetman et al. 2017

⁶¹ Bijma et al. 2013

⁶² Watson et al. 2020

⁶³ UNESCO, 2015

⁶⁴ Sabine et al. 2004

⁶⁵ Thiagarajan et al. 2013

⁶⁶ Sweetman et al. 2017

survival⁶⁷. What is certain is that warming of deep-sea ecosystems will likely affect carbon sequestration (predicted to decline significantly⁶⁸), and the survival and distribution of many species. For example, most deep-sea corals are slow growing, extremely long-lived and have evolved in a specialised and stable environment⁶⁹. This means they are highly susceptible to ocean acidification and other anthropogenic climate change impacts^{70,71}, which could result in changes in their depth range distributions⁷² (potentially become more restricted⁷³) and could even cause some species to go extinct⁷⁴.

While deep-sea ecosystems are increasingly vulnerable from the impacts of climate change⁷⁵, they are also highly vulnerable to physical disturbance such as destructive bottom trawling and marine mining. These destructive methods break down these carbon stores as they can re-mineralise sedimentary carbon to CO₂ which could increase ocean acidification, reduce the buffering capacity of the ocean, and potentially add to the build-up of atmospheric CO₂⁷⁶.

Clark et al. (2010) found that as few as ten individual bottom trawls can reduce deep-sea coral forests to rubble⁷⁷. This is important as it not only illustrates the impact trawling has had, but that trawling on new areas, even just a few times, can have catastrophic cumulative impacts to deep-sea benthic communities and ecosystem services.

The latest government data indicates that 14 tonnes of coral⁷⁸ was dragged up in bottom trawl nets in Aotearoa waters in a single year (2018-2019)⁷⁹. Scientists estimate that only a small fraction of the coral that is smashed on the seabed by trawlers, actually makes it up to the surface in the net. For example, 250 kilograms of deep-sea corals brought up in the net indicates that 27-85 tonnes of corals were destroyed on the seabed⁸⁰.

International Obligations to Protect Vulnerable Marine Ecosystems, like Seamounts and similar features

The ecosystems associated with seamounts and other similar underwater features like hydrothermal vents are often referred to as vulnerable marine ecosystems (VMEs)⁸¹. This is because of the diverse and often unique species or communities that are associated with them,

⁶⁷ Sweetman et al. 2017

⁶⁸ Sweetman et al. 2017

⁶⁹ Thiagarajan et al. 2013

⁷⁰ Thiagarajan et al. 2013

⁷¹ Dijkstra et al. 2021

⁷² Tracey & Hjørvarsdóttir, 2019

⁷³ Thiagarajan et al. 2013

⁷⁴ Dijkstra et al. 2021

⁷⁵ Le & Sato, 2017

⁷⁶ Sala et al. 2021

⁷⁷ Clark et al. 2010 found that for New Zealand seamounts in depths of 700 - 1,000m as few as 10 trawls could eliminate corals from 15 to 20% cover to no visible cover.

⁷⁸ This estimate is based on the Department of Conservation's calculated figures for observed coral bycatch per 100 trawls and is based on just under 20% observer coverage. The squid fishery and deep-water bottom trawl fisheries (orange roughy, cardinal and oreo) accounted for 85% of the observed coral bycatch. Source: Weaver, 2020

⁷⁹ Weaver, 2020

⁸⁰ Pitcher et al. 2019. From this ratio, 1 mt in the net equates to 108 - 340 mt on the seabed

⁸¹ Food and Agriculture Organization, 2009 Note VMEs are described on FAO website [Vulnerable Marine Ecosystems | Food and Agriculture Organization of the United Nations \(fao.org\)](https://www.fao.org/vulnerable-marine-ecosystems/)

referred to as indicator species (or communities⁸²). VME indicator species have specific life characteristics, primarily that they are long-lived and are easily damaged by physical disturbance, such as fishing gear due to their physical and functional fragility, such as corals, sponges and other epifauna⁸³.



© Paul Caiger. Gorgonian fan (Alcyonacea), Poor Knights Islands, New Zealand 2020

The UN Convention on the Law of the Sea (UNCLOS) sets out the obligations of States⁸⁴ to protect, conserve and manage the ocean. Part XII of UNCLOS⁸⁵, including Article 192 and Article 194(5) imposes on states a general obligation to protect and preserve the marine environment as well as more specific obligations relating to the protection and preservation of rare or fragile ecosystems and the habitat of depleted, threatened, or endangered species. Such species and habitats include corals⁸⁶.

The United Nations Fish Stocks Agreement⁸⁷ provides for obligations to implement the precautionary approach (Article 6), protect ecosystems and protect biodiversity (Article 5(g)). The United Nations General Assembly (UNGA) resolutions have addressed the impacts of fishing on VMEs in international waters, with specific reference to seamounts, hydrothermal vents, and cold-water corals.

⁸² Watling & Auster, 2021

⁸³ Watling & Auster, 2021

⁸⁴ State / Country - referring to a political unit that has sovereignty over an area of territory and the people within it. UN Member States can be found here: [Member States | United Nations](#)

⁸⁵ UNGA resolution 59/25, paragraphs 66-71. Adopted 2004

⁸⁶ DSCC, 2021

⁸⁷ United Nations, 1995. The UN Fish Stocks Agreement

In 2004⁸⁸ and 2006 UNGA adopted resolutions calling for urgent action to protect seamounts and other VMEs in international waters. States and Regional Fisheries Management Organisations (RFMOs) were urged to “*implement measures, in accordance with the precautionary approach, ecosystem approaches and international law*” by the end of 2008 or else prohibit bottom fishing on the high seas VMEs. Resolution 61/105 (2006)⁸⁹ required specific measures including prior impact assessments, identifying VMEs and closing them or otherwise protecting them, and requiring vessels to stop fishing when they encounter a VME by bringing up deep-sea coral and/or other indicator species in their nets (the “move-on rule”)⁹⁰.

These UNGA resolutions and FAO guidelines for seamount protection in international waters were established because of the crucial importance of seamounts. In 2009 the UNGA reviewed the measures that had been taken to protect deep-sea biodiversity and concluded that more needed to be done by States to implement the 2006 resolution.

New Zealand has also committed in Aichi Target 6 that by 2020 “*fisheries have no significant adverse impacts on threatened species and vulnerable ecosystems*”⁹¹, and committed in Sustainable Development Goal 14 to Target 14.2: “*By 2020 sustainably manage and protect marine and coastal ecosystems to avoid significant adverse impacts, including by strengthening their resilience and take action for their restoration, to achieve healthy and productive oceans*”⁹².

More recently, the UN General Assembly has declared the period 2021-2030 as the Decade on Ecosystem Restoration for preventing, halting and reversing the degradation of ecosystems worldwide⁹³.

Aotearoa Obligation to Protect Vulnerable Marine Ecosystems, like Seamounts and similar features

New Zealand not only has these international obligations to implement within Aotearoa’s Exclusive Economic Zone (EEZ⁹⁴) but has domestic legislation and policies to protect biodiversity and avoid, remedy or mitigate against adverse fishing effects on seamounts and similar features.

Key legislation, policies and fish plans include the Fisheries Act 1996, Wildlife Act 1953, Exclusive Economic Zone and Continental Shelf (Environmental Effects) Act 2012, Te Mana o te Taiao, the Aotearoa New Zealand Biodiversity Strategy 2020 and the Deepwater Fish Plan.

Additional legislations and policies such as the Marine Protected Areas Policy and the Marine Reserves Act 1971 are relevant when it comes to protecting areas, which could include seamounts and similar features if they are located within the territorial waters (0-12nm). Beyond 12 nm, outside of the territorial seas to the EEZ (200 nm offshore) marine reserves cannot be established. Benthic protection can be put in place using provisions in the Fisheries Act or via special legislation.

⁸⁸ UNGA resolution 59/25, paragraphs 66-71. Adopted 2004.

⁸⁹ UNGA resolution 61/105, paragraphs 76-87. Adopted 2006.

⁹⁰ DSCC, 2021

⁹¹ [Convention on Biological Diversity \(cbd.int\)](https://www.cbd.int/)

⁹² [Goal 14: Life Below Water | The Global Goals](#)

⁹³ [UN Decade on Restoration](#)

⁹⁴ The EEZ extends 200 nautical miles from the coast of New Zealand (including the coast of all islands)

The Fisheries Act 1996

The Fisheries Act 1996 does not explicitly require or provide for an ecosystem-based approach to be applied to fisheries management. The purpose under Section 8 is “to provide for the utilisation of fisheries resources while ensuring sustainability”.

Ensuring sustainability is defined as meaning under s8(2)⁹⁵:

- (a) “Maintaining the potential of the fisheries resources to meet the reasonably foreseeable needs of future generations”, and
- (b) **“Avoiding, remedying, or mitigating any adverse effects of fishing on the aquatic environment”**⁹⁶

Utilisation is defined as meaning under s8(2)⁹⁷ “means conserving, using, enhancing, and developing fisheries resources to enable people to provide for their social, economic, and cultural well-being”

Section 9 of the Fisheries Act describes the environmental principles⁹⁸ that are to be taken into account when decisions are made by the Minister for Oceans and Fisheries:

- (a) “Associated or dependent species should be maintained above a level that ensures their long-term viability”
- (b) **“Biological diversity of the aquatic environment should be maintained”**⁹⁹
- (c) “Habitat of particular significance for fisheries management should be protected”

Section 10 of the Fisheries Act describes the four information principles¹⁰⁰ to be taken into account:

- (a) “Decisions should be based on the best available information”
- (b) “Decision-makers should consider any uncertainty in the information available in any case”
- (c) “Decision-makers should be cautious when information is uncertain, unreliable, or inadequate”
- (d) “The absence of, or any uncertainty in, any information should not be used as a reason for postponing or failing to take any measure to achieve the purpose of the Fisheries Act”

As stated in the High Court ruling¹⁰¹ “the purpose of the Act to provide for the utilisation of fisheries resources while ensuring sustainability (Fisheries Act Section 8)” “As the Supreme Court said in *New Zealand Recreational Fishing Council Inc v Sanford Ltd* (Supreme Court Kahawai case)¹⁰²”

“Section 8(1) appears in Part 2 of the Act headed “Purpose and principles”. It expresses a single statutory purpose by reference to the two competing social policies reflected in the Act. Those competing policies are “utilisation of fisheries” and “ensuring sustainability”. The meaning of each term in the Act is defined in s 8(2). The statutory purpose is that both policies are to be accommodated as far as is practicable in the administration of fisheries under the quota management system. But recognising the

⁹⁵ [Fisheries Act 1996 No 88 \(as at 01 April 2021\), Public Act Contents – New Zealand Legislation](#)

⁹⁶ [emphasis added]

⁹⁷ [Fisheries Act 1996 No 88 \(as at 01 April 2021\), Public Act Contents – New Zealand Legislation](#)

⁹⁸ [Fisheries Act 1996 No 88 \(as at 01 April 2021\), Public Act Contents – New Zealand Legislation](#)

⁹⁹ [emphasis added]

¹⁰⁰ [Fisheries Act 1996 No 88 \(as at 01 April 2021\), Public Act Contents – New Zealand Legislation](#)

¹⁰¹ *Royal Forest and Bird Protection Society of New Zealand Incorporated v Minister of Fisheries* [2021] NZHG 1427

¹⁰² *New Zealand Recreational Fishing Council Inc v Sanford Ltd* [2009] NZSC 54, [2009] 3 NZLR 438 [Supreme Court Kahawai case]

inherent unlikelihood of those making key regulatory decisions under the Act being able to accommodate both policies in full, s 8(1) requires that in the attribution of due weight to each policy that given to utilisation must not be such as to jeopardise sustainability. Fisheries are to be utilised, but sustainability is to be ensured^{103, 104}.

Court rulings confirm fisheries utilisation is not a balance or a trade off made against sustainability by the Minister but that “*sustainability is to be ensured*” and as such must be prioritised. To date s 8(2)(b) and s 9 have not been applied adequately to protect seamounts and similar features and the VMEs (biodiversity) associated with them as the Act has not ensured that “*biological diversity of the aquaculture environment should be maintained*” and that adverse effects of fishing on the aquatic environment are avoided, remedied or mitigated. In Aotearoa waters under the Fisheries Act a commercial fishing vessel can destroy an unlimited amount of VME taxa or indicator species, such as protected corals while legally bottom trawling¹⁰⁵. This is inconsistent with international law, with the Law of the Sea Convention in Article 192 requiring that “*States have the obligation to protect and preserve the marine environment.*” and in Article 194(5) requiring measures “*necessary to protect and preserve rare or fragile ecosystems as well as the habitat of depleted, threatened or endangered species and other forms of marine life*”.

New Zealand regulations are weaker than those in place in international waters regarding the amount of VME indicator species such as corals, that can be destroyed by New Zealand flagged commercial fishing vessels¹⁰⁶. Within the EEZ there is no “move-on rule”, which would require a vessel to move one nautical mile away from where it was trawling if a certain amount of VME indicator taxa is caught, which is the case for example in the South Pacific¹⁰⁷ where New Zealand flagged vessels fish.

Under the Fisheries Act, in 2007 32% of Aotearoa’s EEZ¹⁰⁸ across a number of areas were closed to bottom trawling and dredging, these are known as benthic protected areas (BPAs). These BPAs have been widely criticised for lacking in scientific basis related to their biodiversity value or species representativeness^{109,110}. The areas were largely chosen by the commercial fishing industry and 72% of these BPAs are waters deeper than two kilometres¹¹¹. This means most of these BPAs are well beyond the current trawlable depths that extend to around 1,600 meters¹¹², and also the core-depth distribution of VME taxa, such as protected corals¹¹³.

A small fraction of Aotearoa’s seamounts and associated biodiversity has been surveyed and benthic taxa described meaning there is no way to accurately determine how much biological diversity loss has occurred due to the effects of benthic fishing. There is no requirement to take a

¹⁰³ [emphasis added]

¹⁰⁴ New Zealand Recreational Fishing Council Inc v Sanford Ltd [2009] NZSC 54, [2009] 3 NZLR 438 [Supreme Court Kahawai case]

¹⁰⁵ As long as the Wildlife Act 1953 regulations are followed

¹⁰⁶ Discussed further in Section 2

¹⁰⁷ South Pacific Regional Fisheries Management Area

¹⁰⁸ Ministry for Primary Industries, 2021 Benthic Protected Areas [Benthic protection areas | Fishing and aquaculture | NZ Government \(mpi.govt.nz\)](https://www.mpi.govt.nz/benthic-protection-areas/)

¹⁰⁹ Rieser et al. 2013

¹¹⁰ Leathwick et al. 2008

¹¹¹ Leathwick et al. 2008

¹¹² Maximum trawlable depth is about 1,600 m, from Tracey, 2018.

¹¹³ Tracey & Hjørvarsdóttir, 2019

precautionary approach. If Section 9 included the precautionary approach and required management decisions to be guided by this, it could help give effect to these principles, as this would ensure that the lack of full scientific certainty is not used as a reason not to act to prevent further environmental and biodiversity degradation and loss.

Under Section 11¹¹⁴ or Section 297(1)(a)(vii)¹¹⁵ of the Fisheries Act 1996 the Minister could regulate bottom trawling and ban it on seamounts and similar underwater features within Aotearoa. If such closers were put in place to meet s 8(2)(b) and s9 principles this would not reduce the amount of the allocated quota (TAC or specifically the TACC) for any affected fish stocks. These can only be adjusted using s 13 – total allowable catch of the Fisheries Act.

Exclusive Economic Zone and Continental Shelf (Environmental Effects) Act 2012

The Exclusive Economic Zone and Continental Shelf (Environmental Effects) Act 2012 regulates the environmental effects of mineral mining activities (and other seabed activities) beyond the territorial sea and has a specific requirement that when certain marine consents are considered the consent authority must take into account under Subpart 2D – Decision Section 59(2)¹¹⁶ the following¹¹⁷:

“59(2)(a) any effects on the environment or existing interests of allowing the activity, including—

(i) cumulative effects; and

(ii) effects that may occur in New Zealand or in the waters above or beyond the continental shelf beyond the outer limits of the exclusive economic zone; and

59(2)(d) the importance of protecting the biological diversity and integrity of marine species, ecosystems, and processes¹¹⁸

59(2)(e) the importance of protecting rare and vulnerable ecosystems and the habitats of threatened species¹¹⁹”

Wildlife Act 1953

The Wildlife Act 1953 outlines the protection and control of wild animals and birds and the management of game. Schedule 7A¹²⁰ lists the marine species that are protected¹²¹. In 2010 the Act was amended to protect four coral groups in Aotearoa, these are:

- black corals (all species in the order Antipatharia)

¹¹⁴ Section 11 –allows the Minister to set a sustainability measure and Section 11(3) allows the Minister to limit areas where a stock can be caught and fishing methods used. Section 297(1)(a)(vii) allows for regulating or prohibiting any method of fishing

¹¹⁵ Section 297 – general regulations. Allows for regulating or controlling fishing, specifically (vii) *“regulating or prohibiting any method of fishing”*

¹¹⁶ [Exclusive Economic Zone and Continental Shelf \(Environmental Effects\) Act 2012 No 72 \(as at 08 September 2018\), Public Act 59 Marine consent authority’s consideration of application – New Zealand Legislation](#) Note emphasis added to demonstrate particularly relevant statement for seamounts and similar features and their VME and associated biodiversity

¹¹⁷ Not all 59(2) is listed, the following are relevant towards seamounts and similar features and their associated biodiversity (VMEs).

¹¹⁸ emphasis added

¹¹⁹ emphasis added

¹²⁰ Schedule 7A: [Wildlife Act 1953 No 31 \(as at 07 August 2020\), Public Act – New Zealand Legislation](#)

¹²¹ Non-coral protected marine species: oceanic whitetip shark, basking shark, deepwater nurse shark, white pointer shark, whale shark, manta ray, spinetail devil ray, giant grouper & spotted black grouper

- gorgonian corals (all species in the order Scleractinia)
- stony corals (all species in the order Scleractinia)
- hydrocorals (all species in the family Stylasteridae)¹²²

The protection status means it is illegal to deliberately collect, harm or kill these animals, but under Section 63B a permitted commercial fisher can destroy and kill any protected coral species while fishing in Aotearoa’s waters, as long as they record the event and report it in writing within 48 hours (or as soon as practicable) and return the protected coral to the sea immediately¹²³.

The current Wildlife Act is inadequate as it does not protect these coral groups from being destroyed. It also only protects these specific coral groups, so if new species of corals (outside of those groups listed) are discovered within Aotearoa, they would not have a protection status.

Te Mana o te Taiao, the Aotearoa New Zealand Biodiversity Strategy 2020

Te Mana o te Taiao, the Aotearoa New Zealand Biodiversity Strategy (ANZBS) was updated and released in 2020. ANZBS “sets a strategic direction for the protection, restoration and sustainable use of biodiversity, particularly indigenous biodiversity, in Aotearoa New Zealand”¹²⁴. ANZBS has the following objectives that agencies need to give effect to, including in advice to Ministers:

- “By 2025, the number of fishing-related deaths of protected marine species is decreasing towards zero for all species.”
- “By 2030 the direct effects of fishing do not threaten protected marine species populations or their recovery.”
- “Zero non-target mortality by 2050 and marine fisheries resources are abundant, resilient and managed sustainably to preserve ecosystem integrity”.

Bottom trawling on seamounts and similar features where VMEs occur is not compatible with ANZBS.

National Fisheries Plan for deepwater and Middle-depth Fisheries 2019

The National Deepwater Fisheries Plan for Deepwater and Middle-depth Fisheries (Deepwater Plan) is a Fisheries New Zealand (FNZ) Plan that sets “management objectives, actions, and services required to meet relevant legislative obligations and strategic directions for managing New Zealand’s deepwater fisheries”¹²⁵.

The Deepwater Plan has four specific environmental outcomes:

- “Ensure that maintenance of biological diversity of the aquatic environment and protection of habitats of particular significance for fisheries management are explicitly considered in management
- Manage deepwater and middle-depth fisheries to avoid, remedy or mitigate the adverse effects of these fisheries on associated or dependent and incidentally caught fish species
- Manage deepwater and middle-depth fisheries to avoid, remedy or mitigate the adverse effects of these fisheries on the benthic habitat
- Manage deepwater and middle-depth fisheries to avoid, remedy or mitigate the adverse effects of these fisheries on the long-term viability of endangered, threatened and protected species populations”

¹²² Schedule 7A: [Wildlife Act 1953 No 31 \(as at 07 August 2020\), Public Act – New Zealand Legislation](#)

¹²³ Unless part of an observer collection program

¹²⁴ The New Zealand Government, 2020.

¹²⁵ Fisheries New Zealand, 2019

The Deepwater Plan has specific management action for each objective, the most relevant is “*Explore the role of protecting marine biodiversity as a strategy to build the resilience of marine ecosystems and fish stocks to buffer the effects of climate change*”. The Deepwater Plan does not have any specific management actions to reduce benthic impacts on protected, endangered or threatened VME taxa like corals.



©NIWA. *Flabellum knoxi* (endemic cup coral species)

Section 2: Overview of seamounts and similar features biodiversity found in Aotearoa

Aotearoa has over 800 known¹²⁶ seamounts within the EEZ greater than 100m high¹²⁷. Most vary in water depth, height, geological origin, age, and level of primary productivity¹²⁸. Figure 2 shows the locations of all known seamounts¹²⁹ and existing benthic protection, and Figure 3 is an example of an Aotearoa seamount complex.

¹²⁶ FNZ has commissioned the project BEN2020-07 to update the NZ database of all known seamounts and seamount-like features exceeding 100m elevation above the seafloor. Greenpeace requested a copy of the interim results (locations of the seamounts) of this project to include in this report but was denied. We have been advised the report will be published in January 2022.

¹²⁷ Rowden & Clark, 2004

¹²⁸ Rowden & Clark, 2004

¹²⁹ While this is 2008 it was the most recent data Greenpeace could access from Rowden et al, 2008, seamount table supplied by NIWA via email.

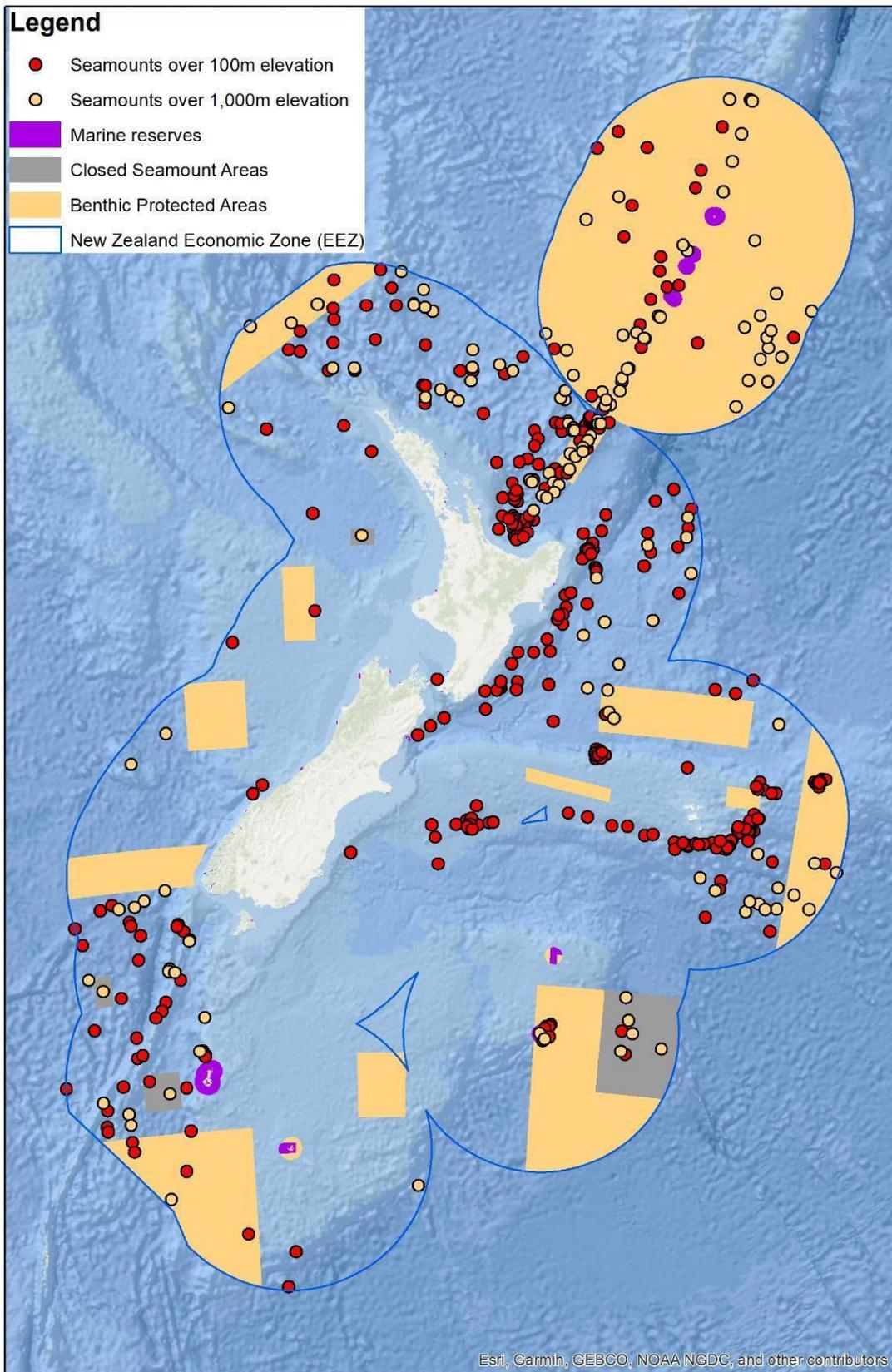


Figure 2: Location of known seamounts within Aotearoa that have a 100m or higher elevation (red) and a 1,000m or higher elevation (orange), along with existing benthic closures . Sources Rowden et al, 2008; Clark, 2021; New Zealand Government, 2021a,b & c.

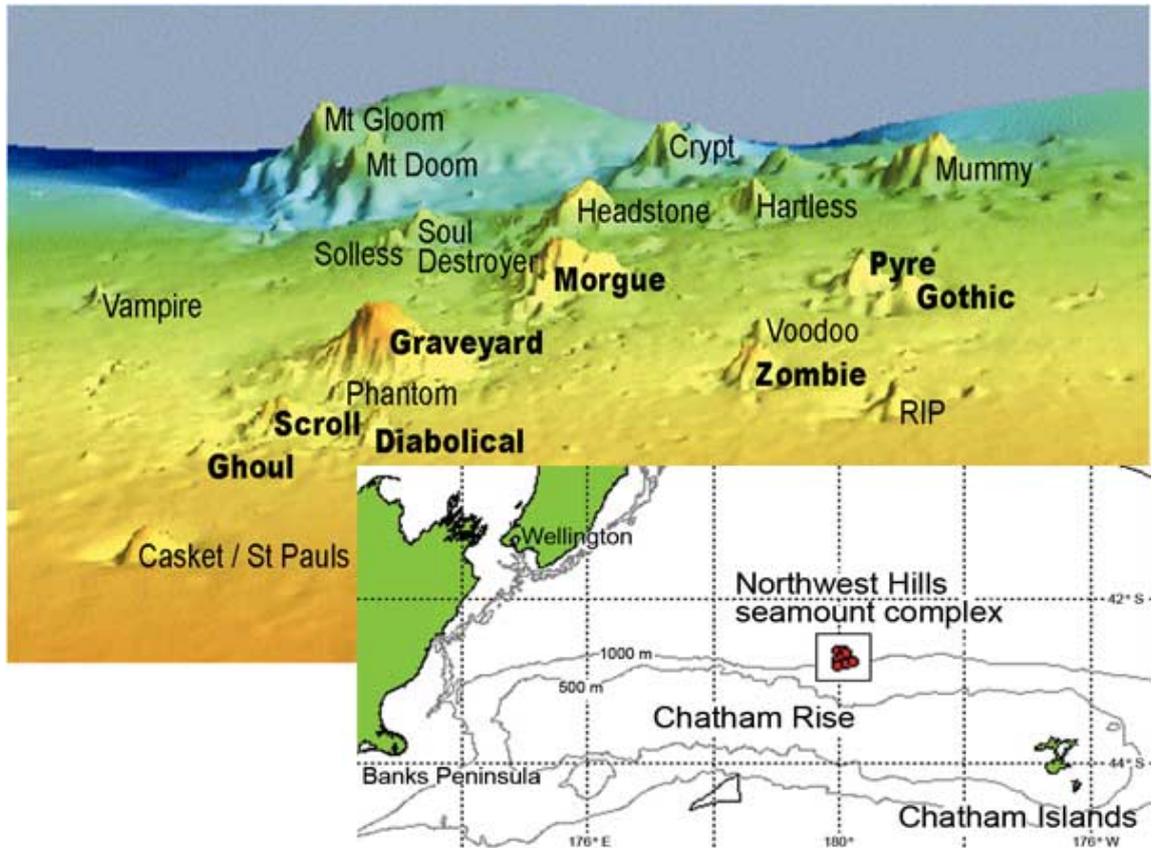


Figure 3: Shows the Northwest Hills seamount complex, located approximately 500 km east of Kaikoura and about halfway to the Chatham Islands. The seamounts in bold have been surveyed in detail and are between 100–400m high, rising from the seafloor at about 1,200m depth. Source: Clark, 2008

Benthic invertebrate biodiversity overall within Aotearoa is still poorly described¹³⁰. However, Watling and Auster (2017) reviewed seamount data and the presence of VME indicator species from the South Pacific¹³¹ and found that “*all seamounts that have so far been surveyed by cameras, either towed or mounted on maneuverable submersible vehicles, have been found to have abundant VME indicator species (including xenophyophores on sandy areas) distributed on their sides and summits*”¹³². This reinforces the assumption that all seamounts that occur within the biological depth ranges of VME indicator species, like deep-sea corals, should be considered to contain VMEs and therefore warrant full protection in order to meet New Zealand’s domestic policies and international commitments.

NIWA has mapped the locations of VME in the wider South Pacific¹³³. Figure 4 shows the known locations of VMEs within Aotearoa’s EEZ.

¹³⁰ Clark et al, 2019b

¹³¹ Including camera tows from some of the Louisville Seamounts from Clark et al, 2015

¹³² Watling and Auster, 2017

¹³³ NIWA, 2016 Data available via OBIS

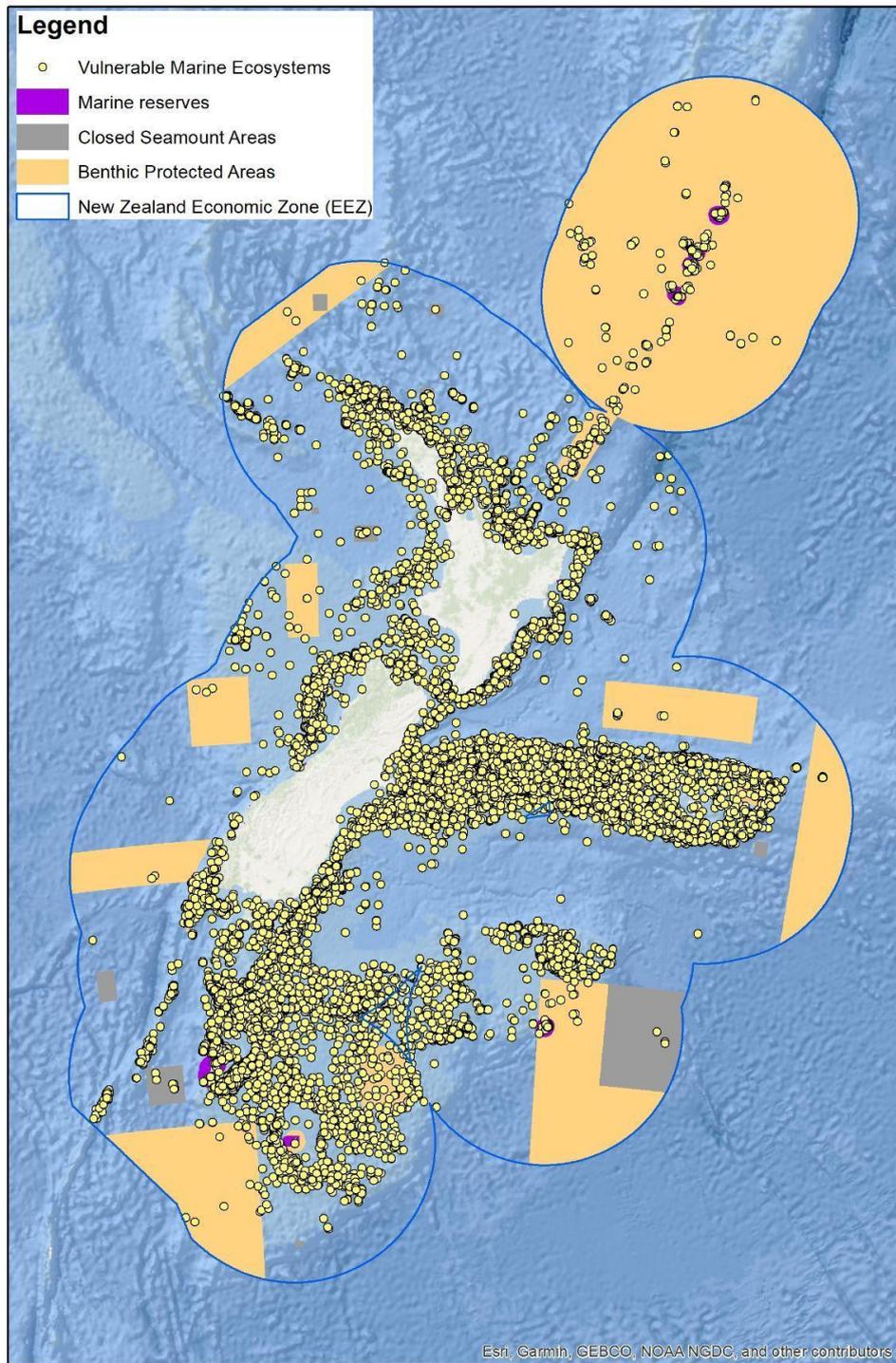


Figure 4: Location of known VMEs within Aotearoa’s EEZ, also showing existing benthic protection (closures).
 Source: NIWA, 2016 & New Zealand Government, 2021a,b&c.

Deep-sea corals

Aotearoa's deep-sea cold-water corals are diverse, often unique^{134,135} and are a key component of Aotearoa's deep-sea ecosystems¹³⁶. Deep-sea corals are important as they can form biogenic hotspots that provide habitat, sanctuary, and nursery areas for many associated species, including various invertebrates and fish, including orange roughy and deep-sea cardinal fish species¹³⁷.

Deep-sea corals are mostly slow growing, have low natural mortality, can have short dispersal distances, and display extreme longevity¹³⁸, some even reaching thousands of years^{139,140,141}.

Scientists have aged many coral samples from Aotearoa, for example black coral *Antipatharia* from the Chatham Rise taken at a depth of 870m was aged between 909 – 2,672 years old¹⁴², and bubblegum coral *Paragorgia arborea* was aged between 300 – 500 years old¹⁴³. These aging studies confirm that many deep-sea coral 'forests' have been growing for hundreds to thousands of years, and this is important given a few bottom trawls can wipe out entire coral communities or 'forests'.



© Paul Caiger. Black coral (*Antipathella fiordensis*), Dusky Sound, Fiordland, New Zealand 2021

¹³⁴ Holland et al. 2020

¹³⁵ Bilewitch & Tracey, 2020

¹³⁶ Clark et al., 2010

¹³⁷ Rowden & Clark, 2004

¹³⁸ Consalvey et al. 2006

¹³⁹ Tracey & Hjørvarsdóttir, 2019,

¹⁴⁰ Marriott et al., 2020

¹⁴¹ Hitt et al., 2020

¹⁴² Hitt et al., 2020; Tracey & Hjørvarsdóttir, 2019

¹⁴³ Tracey et al. 2003 described by Tracey and Hjørvarsdóttir, 2019

Endemism

There is evidence of extensive coral endemism within the waters around Aotearoa¹⁴⁴. Table 1, taken from Tracey & Hjørvarasdóttir (2019) summarises the state of knowledge of endemic coral species within Aotearoa’s EEZ based on Cairns *et al* in Gordon 2009. It’s important to note that this table has only been partially updated and that a full literature review is needed. The table does not include recent unpublished benthic invertebrate research commissioned by Fisheries New Zealand, which may have identified an undescribed (and unprotected) new coral species, which could be potentially endemic¹⁴⁵. Regardless, what Table 1 shows is that Aotearoa has at least 196 known endemic coral species, and this number will definitely increase.

Table 1: Summary of endemic coral species from Aotearoa EEZ updated in part (from Cairns et al. in Gordon 2009; Opresko pers. comm). Source: Tracey & Hjørvarasdóttir, 2019 (Table 2).

Taxon Cnidaria	Described living species	Known undescribed, undet. species	Estimate unknown species	Endemic species	Endemic genera
Anthozoa	279	270	140	~111	5
Alcyonacea	46	166	70	~58	4
Pennatulacea	12	19	5	0	0
Scleractinia	124	5	10	16	4
Corallimorpharia	5	0	3	2	0
Antipatharia	28	~80	~52	~20	1
Hydrozoa	454	28	75	121	7
Leptothecata	234	14	30	77	2
Anthoathecata	138	12	20	43	5
Total	~1320	~594	~363	~196	~28

Research has also demonstrated that Aotearoa’s seamounts are incredibly unique as the assemblages of macroinvertebrates (like corals and sponges etc.) can be distinct from neighbouring seamounts separated by relatively small distances (tens of kilometres)¹⁴⁶. This means a small area of ocean with several seamounts could have entirely unique and even endemic macroinvertebrate communities.

Protected deep-sea coral distributions

Not all corals are protected in Aotearoa, only the four groups listed under schedule 7A of the Wildlife Act 1953¹⁴⁷ are. The protection status means it is illegal to deliberately harm or kill these animals, but under Section 63B a permitted commercial fisher can destroy and kill any protected coral species while fishing in Aotearoa’s waters, as long as they record the event and report it in writing within 48 hours (or as soon as practicable), and return the protected coral to the sea

¹⁴⁴ Tracey and Hjørvarasdóttir, 2019

¹⁴⁵ Schnabel et al. (2019) [Unpublished Final Research Report for Fisheries New Zealand Project DAE2018-04, from Schnabel et al. 2021] states that scientists “*reported for the first time the zoanthid genus Bullagummi^zoanthus in the New Zealand region with two samples identified from the EEZ*” and that work is “*underway to determine whether the New Zealand specimens belong to an undescribed species*” Schnabel et al. 2021.

¹⁴⁶ Rowden and Clark, 2004

¹⁴⁷ Schedule 7A: [Wildlife Act 1953 No 31 \(as at 07 August 2020\), Public Act – New Zealand Legislation](#)

immediately¹⁴⁸. The current Wildlife Act is inadequate as it does not protect these coral groups from being destroyed and it excludes any other groups of corals such as hexacorals *Zoanthia* as they are not listed on schedule 7A.

Protected corals groups:

- black corals (all species in the order Antipatharia)
- gorgonian corals (all species in the order Gorgonacea)
- stony corals (all species in the order Scleractinia)
- hydrocorals (all species in the family Stylasteridae)

The following deep-sea coral diversity and distributions are described based primarily on the review by Tracey and Hjørvarsdóttir (2019)¹⁴⁹, but also Consalvey et al. (2006), Owen et al. (2016), Baird, et al. (2013), Tracey et al. (2011), Bilewitch and Tracey (2020a) and (2020b) and Owen et al. (2020).

A large majority of the information used to underpin our understanding of deep-sea coral diversity (such as species identification) and distributions comes primarily from observed bycatch samples from commercial deep-sea fisheries (such as bottom trawling for orange roughy). Additional data sources include limited research trawls or underwater surveys, such as those carried out on seamounts by scientists.

The level of reporting from un-observed commercial vessels is often poor and unreliable¹⁵⁰. Consequently, the only source of accurate information comes from observed vessels bycatch. The problem with this is that coverage across all New Zealand vessels and locations (known as fisheries management areas (FMAs)) can be highly variable ranging from 0% coverage to 100%¹⁵¹, and this data is not generally available to the public¹⁵².

Table 2 shows the variation in observer coverage (percentage of fishing effort observed) and the total reported protected coral bycatch for the latest published fishing year (2018-2019). The squid trawl fishery had the highest observed coral bycatch in the 2018/19 fishing year with over 8,000kg of coral. The deep bottom trawl fisheries (orange roughy, cardinal and oreo species) were reported to have 526.9kgs of coral bycatch based on a total of 24.8% observer coverage.

¹⁴⁸ Unless part of an observer collection program

¹⁴⁹ Tracey et al. 2019 (Chapter 12)

¹⁵⁰ A camera trial in the Hauraki Gulf found that inshore fishers report twice as many protected seabirds captures when a camera was on board Source: Tremblay-Boyer & Abraham, 2020. Similar patterns of increased reporting of endangered, protected, or threatened species by commercial fishers when observers are present have been found in other New Zealand fisheries.

¹⁵¹ FNZ is transitioning some fisheries within New Zealand to use electronic monitoring (cameras), once implemented this program should increase New Zealand's coverage.

¹⁵² Observed bycatch data of protected species is reported by DOC in an annual Conservation Services Program report (Weaver, 2020), but the data is grouped, not spatially specific, not available in real time, and not publicly available. The author requested a copy of all the observed bycatch of protected species data broken down by fisheries and location in order to map it. This protected species observed bycatch data is not held by DOC but by FNZ. This request was turned into an Official Information Request and has not been delivered at the time of publishing this report.

Table 2: Summarises the latest report on observed protected coral bycatch in Aotearoa for the 2018/19 fishing year. All data comes from Weaver (2020)¹⁵³

Fishery	Observer coverage ¹⁵⁴	Reported total protected coral bycatch for fishery (kgs)
Middle depth trawl fisheries (hoki, hake, ling and warehouse species)	Range: 0 - 41.4% Total: 31% ¹⁵⁵	45.9
Scampi	Range: 0 – 17.7 % ¹⁵⁶ Total: 14.3%	135.7
Squid	Range: 0 – 93.6% ¹⁵⁷ Total: 82.4%	8,015.7
Pelagic trawl fisheries (mackerel and barracouta)	Range: 0 – 95.4% ¹⁵⁸ Total: 48.2%	11
Deepwater bottom trawl fisheries (orange roughy, cardinal and oreo species)	Range: 0 – 59.2% ¹⁵⁹ Total: 24.8%	526.9
Inshore trawl	Range: 0 – 57.2% ¹⁶⁰ Total: 4%	6
Inshore bottom longline	Range: 0 – 12.2% ¹⁶¹ Total: 3.3%	4
Precision Seafood Harvesting	Range: 0 – 100% ¹⁶² Total: 15.3%	138.2

Even the limited observer bycatch data can under-estimate the diversity of deep-sea coral species as not all observers are capable of accurately identifying deep-sea coral bycatch to a species level while at sea, not all bycatch comes back to shore for genetic testing or expert identification, and consequently we do not know what levels of endemism or cryptic benthic diversity exists in Aotearoa waters.

Bilewitch & Tracey (2020) genetically analysed 91 bycatch specimens of gorgonian corals and was able to identify viable DNA data for 62 specimens which led to a minimum of 34 different species to be identified¹⁶³. This study found their “*rate of discovery of unique species indicates that many more species remain unsampled and that we have not yet documented the limits of gorgonian coral diversity within the sampled bycatch community*”¹⁶⁴. Based on genetic analysis, this study found the orange roughy fishery alone had “*at least 23 species of protected gorgonian coral*” bycatch. Bilewitch & Tracey (2020) also concluded that “*we would expect similarly high numbers within the smooth oreo fishery as well, if*

¹⁵³ Note at the time of finalising this report in early October the latest CSP bycatch report had not been released and requests for the latest information were denied. We note this report was released Wednesday 27th October but was unable to be included.

¹⁵⁴ Range depends on the coverage within different locations / FMAs. Total is the overall observer coverage of fishing effort for the whole fishery. Note zero reflects that some trawls / lines in some FMAs did not have any observer coverage

¹⁵⁵ Weaver, 2020 Table 1.

¹⁵⁶ Weaver, 2020 Table 7

¹⁵⁷ Weaver, 2020 Table 10

¹⁵⁸ Weaver, 2020 Table 13

¹⁵⁹ Weaver, 2020 Table 16

¹⁶⁰ Weaver, 2020 Table 22

¹⁶¹ Weaver, 2020 Table 34

¹⁶² Weaver, 2020 Table 43

¹⁶³ Bilewitch & Tracey, 2020a

¹⁶⁴ Bilewitch & Tracey, 2020a

additional bycatch was sampled to the same extent as for orange roughy”¹⁶⁵. This review highlights that many species of gorgonian octocoral are yet to be discovered and described from within Aotearoa¹⁶⁶.

NIWA has a significant invertebrate database which can be used to highlight the locations of known VMEs¹⁶⁷ (Figure 4) and the specific distributions of some protected corals. But given the nature of the deep-sea, it isn't possible to survey all seamounts and similar features to determine the diversity and location of VME indicator species, like protected coral species. NIWA scientists have developed a habitat suitability model based on key protected coral taxa to predict where they are likely to occur. This habitat suitability model has multiple applications and was used to predict the present and future distributions of 12 protected coral taxa¹⁶⁸. Owen et al. (2020) found there was variability among the 12 coral taxa in the overlap of predicted distributions with current bottom trawling effort. Owen et al. (2020) found the taxa currently most vulnerable to trawling based on the habitat suitability modeling are the thicket-forming corals *Goniocorella dumosa* and *Madrepora oculata*, and the hydrocorals *Errina* species and *Stylaster* species¹⁶⁹.

Stony Corals:

Aotearoa has about 110 known anthellate (lacking symbiotic algae) stony corals (Order Scleractinia). These branching deep-sea corals are predominantly found between 800m – 1,200m water depth (Figure 5,) and are widely distributed around Aotearoa (Figure 6)¹⁷⁰. To date no branching stony corals are listed as endemic, however there are various deep-sea stony cup corals that are endemic to Aotearoa that belong to the genus *Flabellum*¹⁷¹. Figure 7 shows the known distribution of cup corals from the Family Flabellidae and shows that they are widely distributed throughout Aotearoa's known seamounts.

Stony corals found in some regions such as the Chatham Rise are known to have unique genetic diversity¹⁷². This is likely because of the prevailing currents that converge from both the north and the south and the isolation of these stony coral populations¹⁷³.

¹⁶⁵ Bilewitch & Tracey, 2020a

¹⁶⁶ Bilewitch & Tracey, 2020a

¹⁶⁷ NIWA, 2016

¹⁶⁸ Owen et al. (2020)

¹⁶⁹ Owen et al. (2020)

¹⁷⁰ Tracey & Hjørvarsdóttir, 2019

¹⁷¹ Tracey & Hjørvarsdóttir, 2019

¹⁷² Holland et al. (2020)

¹⁷³ Holland et al. (2020)

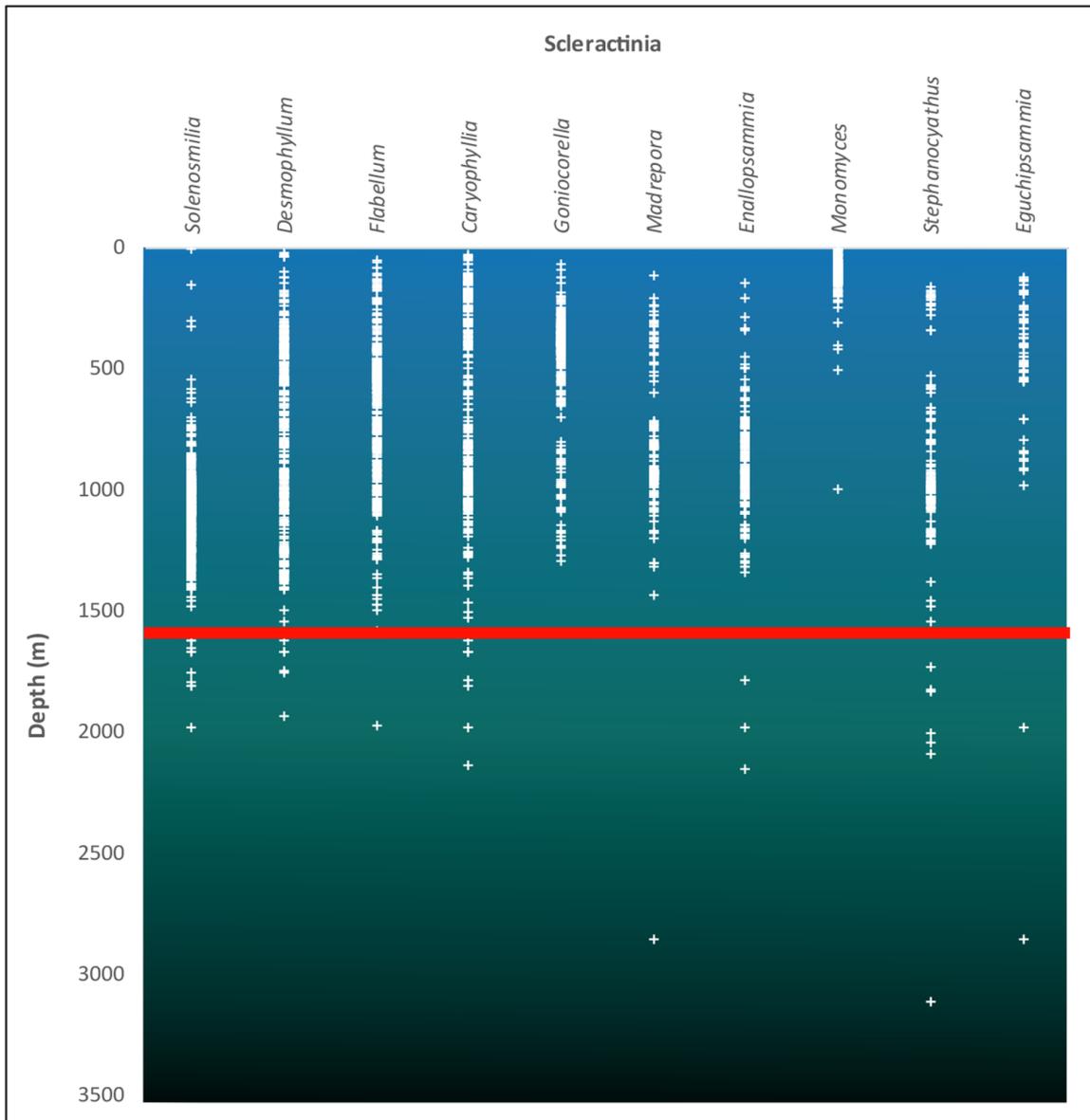


Figure 5: Depth distribution plot of the most abundant reef-forming and cuplike scleractinian stony corals. Source: Tracey & Hjørvarsdóttir, 2019¹⁷⁴. The red line represents the maximum depth New Zealand deep-sea trawlers operate, 1,600m (generally deep-sea trawlers operate between 200m - 800m)¹⁷⁵.

¹⁷⁴ Figure 30

¹⁷⁵ Fisheries New Zealand, 2020

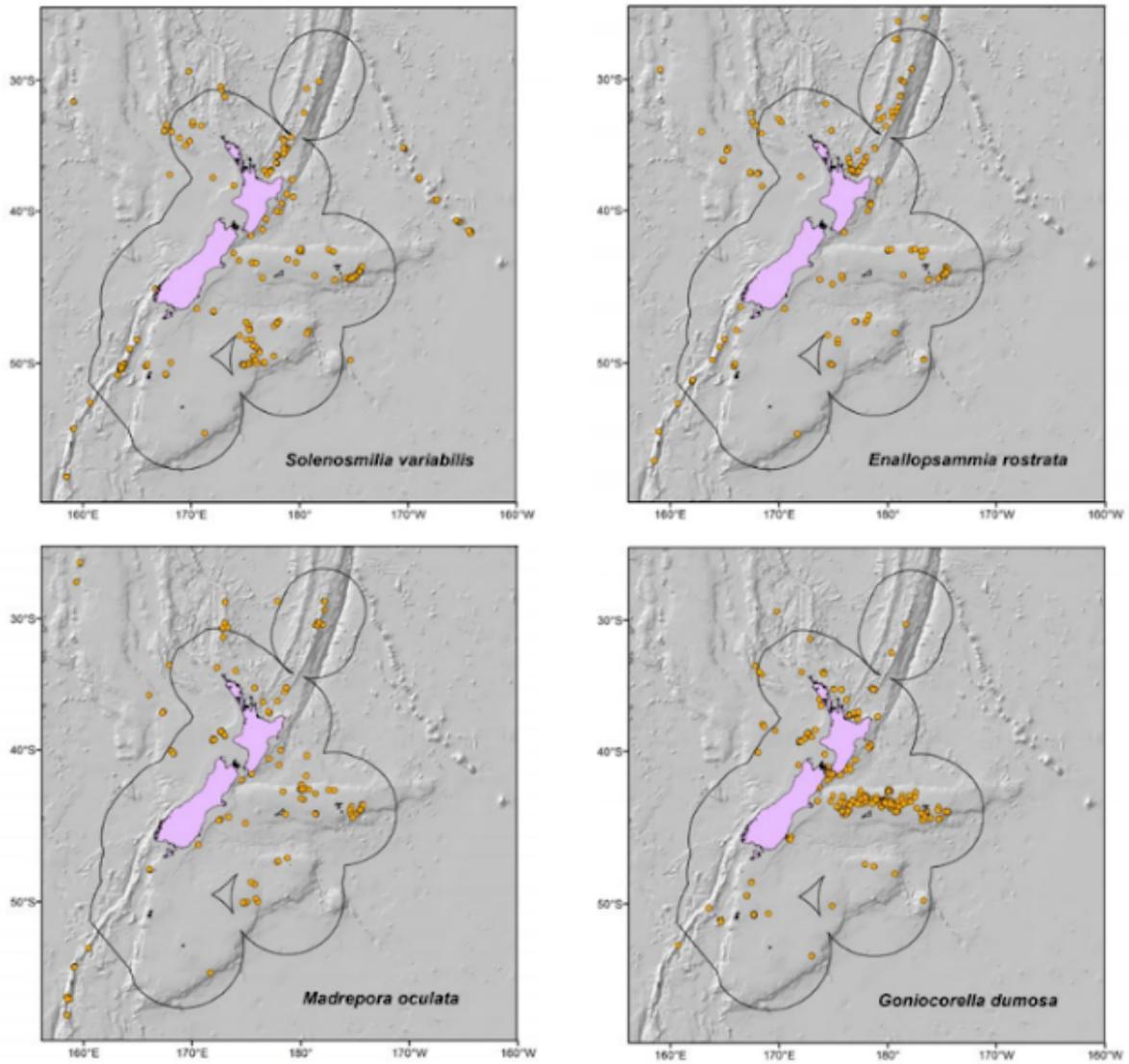


Figure 6: Distribution maps of four key habitat-forming branching stony corals (Scleractinia) found in Aotearoa waters. Published as supplementary material in Anderson et al. (2016). Source Tracey & Hjørvarsdóttir, 2019¹⁷⁶

¹⁷⁶ Figure 18

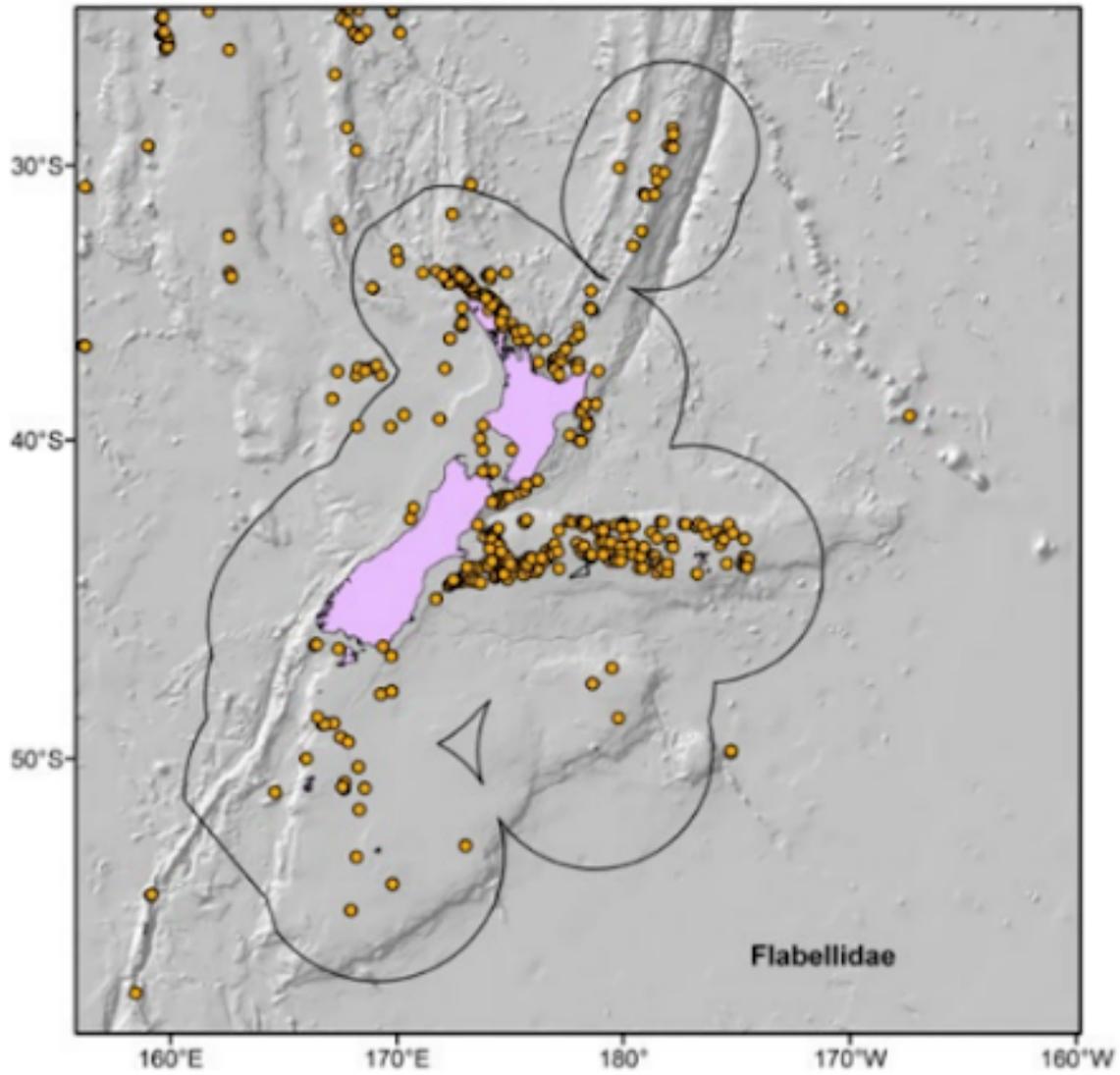


Figure 7: Distribution map of cup coral Family Flabellidae. Published as supplementary material in Anderson et al. (2016). Source Tracey & Hjørvarsdóttir, 2019¹⁷⁷

¹⁷⁷ Figure 34



©NIWA. Flabellum knoxi (endemic cup coral species), New Zealand

Black corals:

Aotearoa is known to have about 80 species of black corals (Order Antipatharia), of which seven have recently been described as new to science¹⁷⁸. Black corals are widely distributed and found throughout the EEZ (Figure 8). Black corals are predominantly found between 200 – 1,000m deep, however some species are known to live deeper over 2,000m deep (Figure 9)¹⁷⁹.

It is not known how many black coral species are endemic. Scientists estimated that approximately 20 will be identified as endemic once all the new species and samples have been fully described¹⁸⁰. A genetic review of some bycatch samples of black coral (*Bathypathes patula*) in Aotearoa in 2020 found that what was thought to be a single species was in fact a cryptic complex of at least five different genera¹⁸¹. It is highly likely that Aotearoa has more unknown species of black coral and that some of these will be endemic and located on seamounts and similar features.

¹⁷⁸ Opresko, 2016 described in Tracey & Hjørvarsdóttir, 2019

¹⁷⁹ Tracey & Hjørvarsdóttir, 2019

¹⁸⁰ Tracey & Hjørvarsdóttir, 2019

¹⁸¹ Bilewitch & Tracey, 2020b

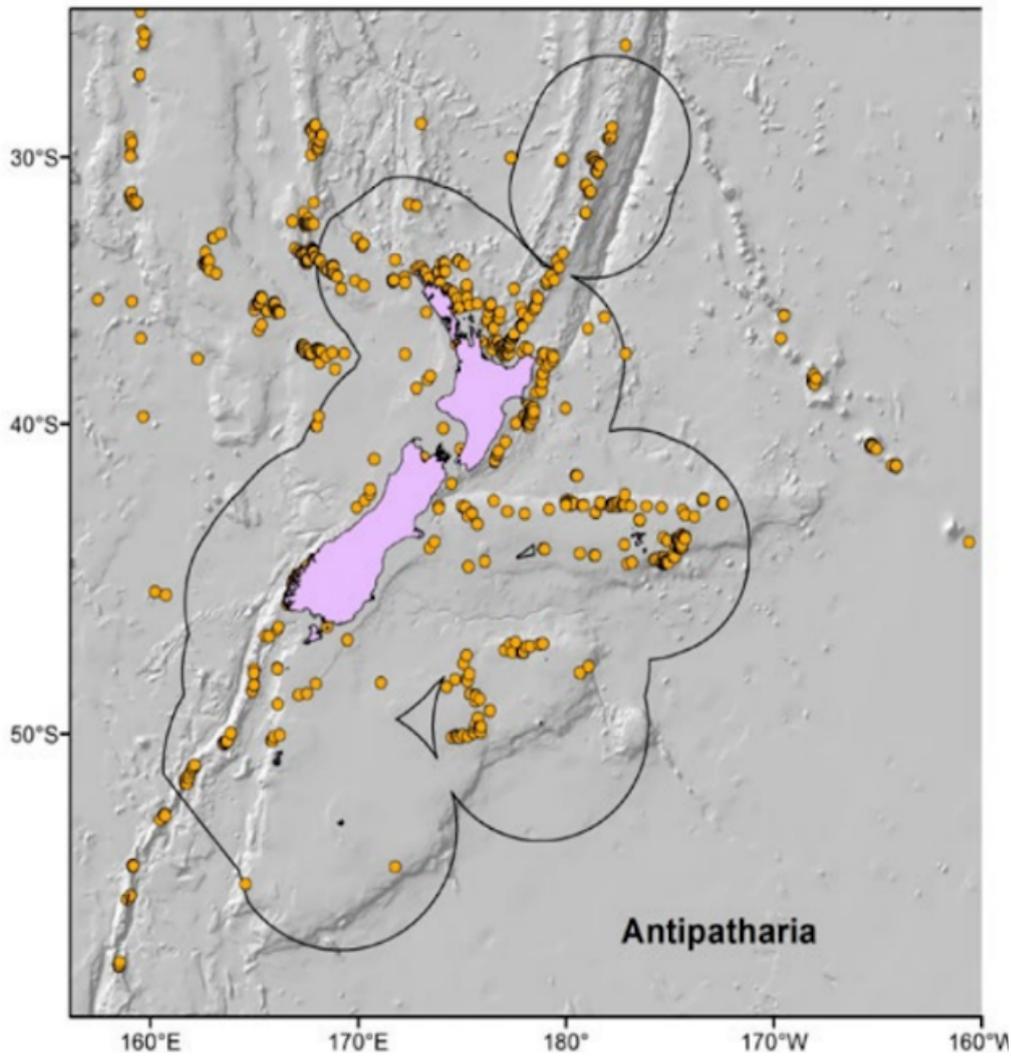


Figure 8: Distribution map of black coral group Antipatharia. Published as supplementary material in Anderson et al. (2016). Source Tracey & Hjørvarsdóttir, 2019¹⁸²

¹⁸² Figure 36



© Greenpeace / Roger Grace, 2004. Protected black coral retrieved from bycatch from a deep-sea trawler in international waters 350 miles west of New Zealand in the Tasman Sea.

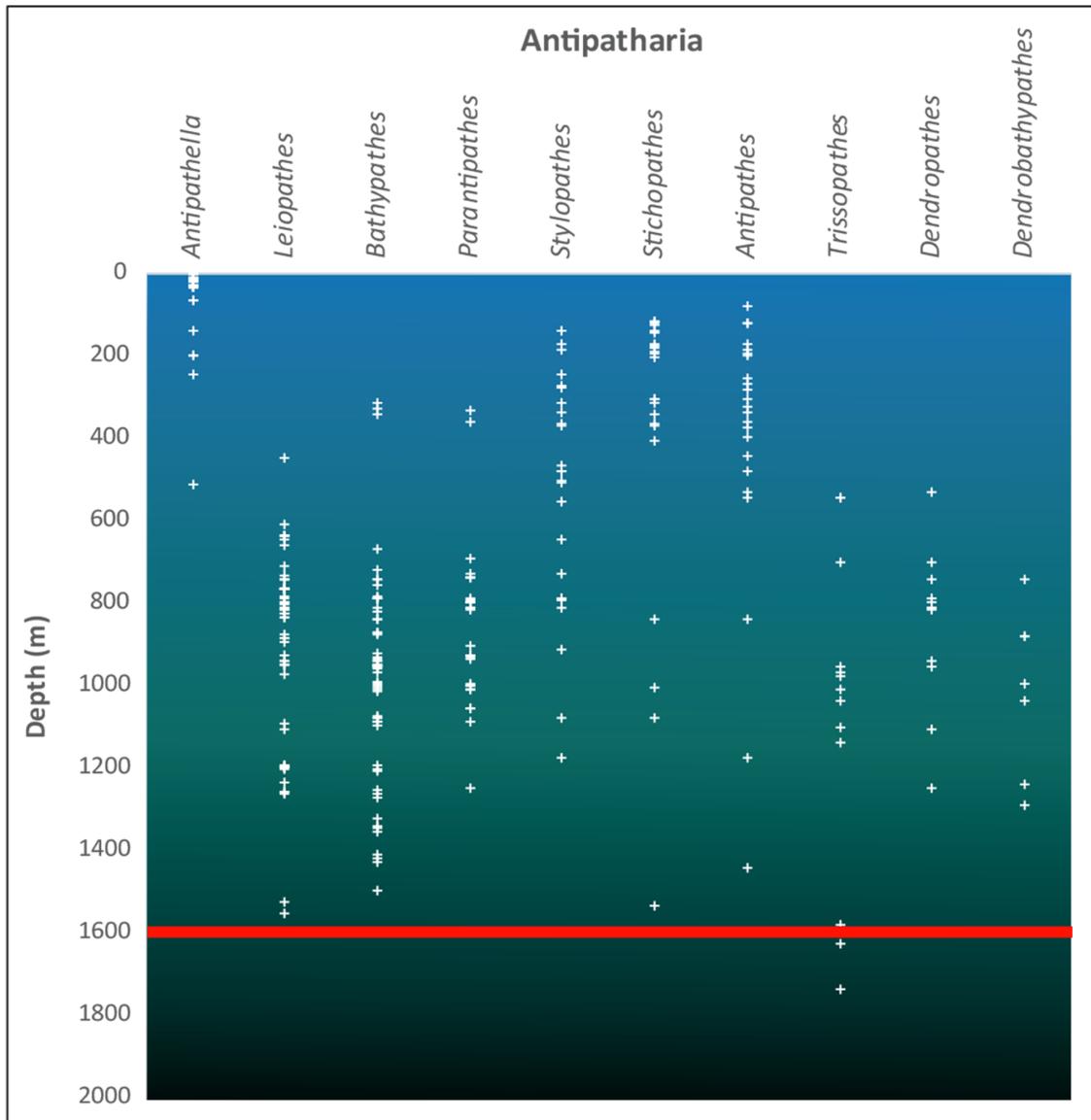


Figure 9: Depth distribution of the ten most abundant black coral genera. Source: Tracey & Hjørvarsdóttir, 2019¹⁸³. The red line represents the maximum depth New Zealand deep-sea trawlers operate, 1,600m (generally deep-sea trawlers operate between 200m - 800m)¹⁸⁴.

Gorgonian Octocorals:

There are over 250 known species of gorgonian octocorals (Order Alcyonacea) found within Aotearoa’s EEZ, with varying distributions (Figure 10). Some gorgonian octocorals are well represented in the deep-sea and can commonly occur at depths of 1,500m or more while others commonly occur between 200m – 1,200m water depth (Figure 11)¹⁸⁵.

¹⁸³ Figure 37

¹⁸⁴ Fisheries New Zealand, 2020

¹⁸⁵ Tracey & Hjørvarsdóttir, 2019

Eight species of bubblegum corals are endemic (*Paragorgia spp*) and one octocoral that lives as a single solitary polyp is also endemic (*Taiaroa taubou*)¹⁸⁶. This endemic octocoral is found widespread on soft sediment on the Chatham Rise. Plexaura is a cosmopolitan family of gorgonian octocorals and is poorly described in Aotearoa but known to occur from nearshore to the deep-sea. It is likely that Aotearoa has endemic plexaurid species, but these have yet to be fully identified¹⁸⁷.

A recent genetic study of octocorals highlights the evolutionary knowledge of the group. It found that “the assemblage of octocoral species contained within bottom trawling bycatch are genetically distinct from each other in the contemporary sense, but the evolutionary processes that generated this current breadth of diversity span a period of at least 140 million years”¹⁸⁸. This supports that Aotearoa is home to a vast number of genetically unique protected corals.

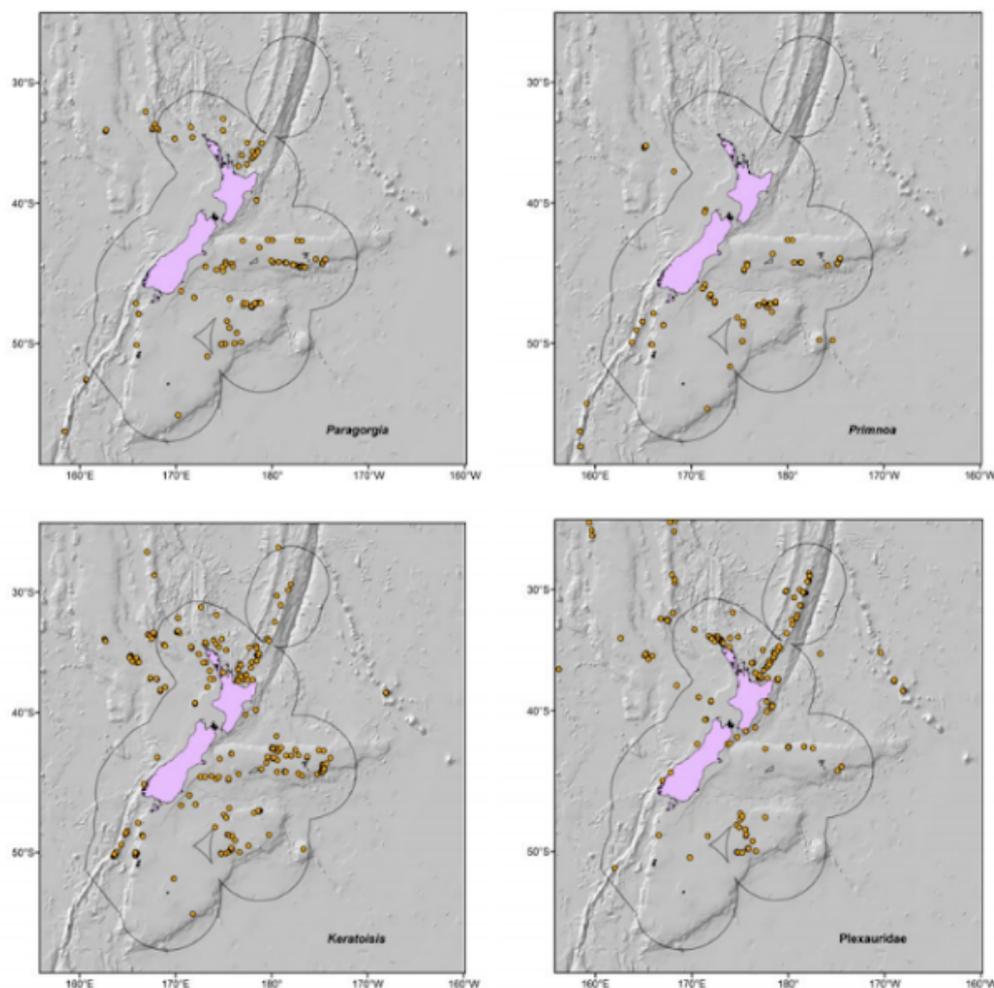


Figure 10: Distribution maps for key gorgonian octocorals located in Aotearoa. Top L to R: Bubblegum coral *Paragorgia* and sea fan *Primnoa*. Bottom L to R: Bamboo coral *Keratoisis* and Family *Plexauridae*. Published as supplementary material in Anderson et al. (2016). Source: Tracey & Hjørvarsdóttir, 2019¹⁸⁹

¹⁸⁶ Tracey & Hjørvarsdóttir, 2019

¹⁸⁷ Tracey & Hjørvarsdóttir, 2019

¹⁸⁸ Park et al. 201 as described in Bilewitch & Tracey, 2020a

¹⁸⁹ Figure 44

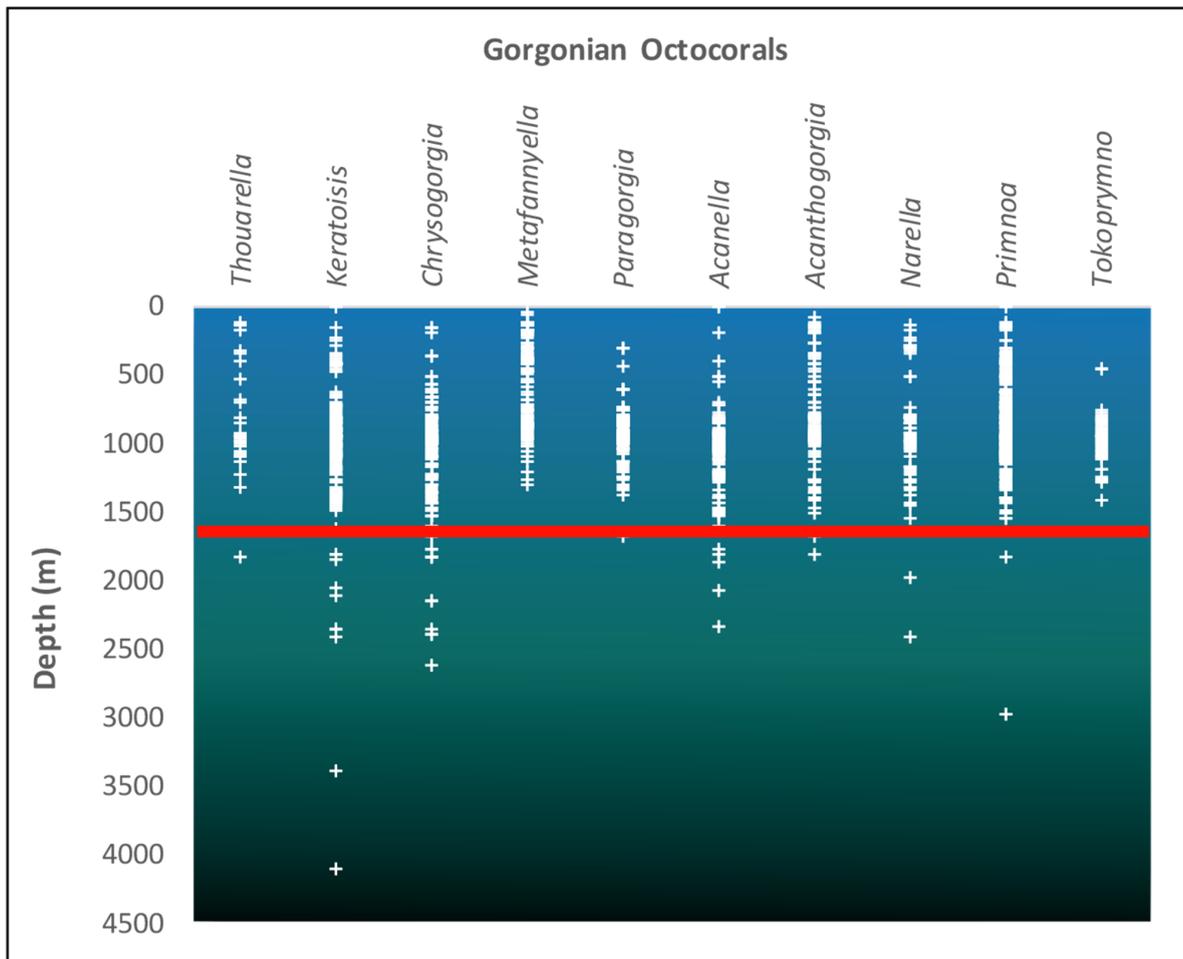


Figure 11: Depth distribution of commonly occurring gorgonian octocorals. Source: Tracey & Hjørvarsdóttir, 2019¹⁹⁰. The red line represents the maximum depth New Zealand deep-sea trawlers operate, 1,600m (generally deep-sea trawlers operate between 200m - 800m)¹⁹¹.

Hydrocorals:

Several species of hydrocorals (Family Stylasteridae) are found throughout Aotearoa’s EEZ (Figure 12), one of which is known to be endemic (*Errina novaezelandiae*). Stylasterid hydrocorals have a large depth range¹⁹², but most genera commonly occur within 200m – 1,000m depth (figure 13).

¹⁹⁰ Figure 41

¹⁹¹ Fisheries New Zealand, 2020

¹⁹² Tracey & Hjørvarsdóttir, 2019

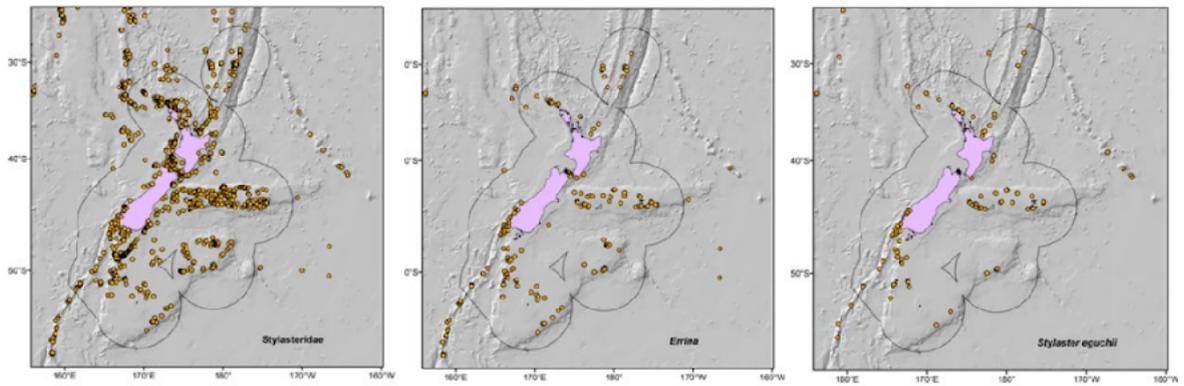


Figure 12: Distribution map of all Stylasteridae hydrocorals (left), and most common species *Errina novaezelandiae* (middle), and *Stylaster eguchii* (right). Published as supplementary material in Anderson et al. (2016). Tracey & Hjørvarsdóttir, 2019¹⁹³



© Paul Caiger. Hydrocoral, Kermadec Islands, New Zealand 2020

¹⁹³ Figure 46

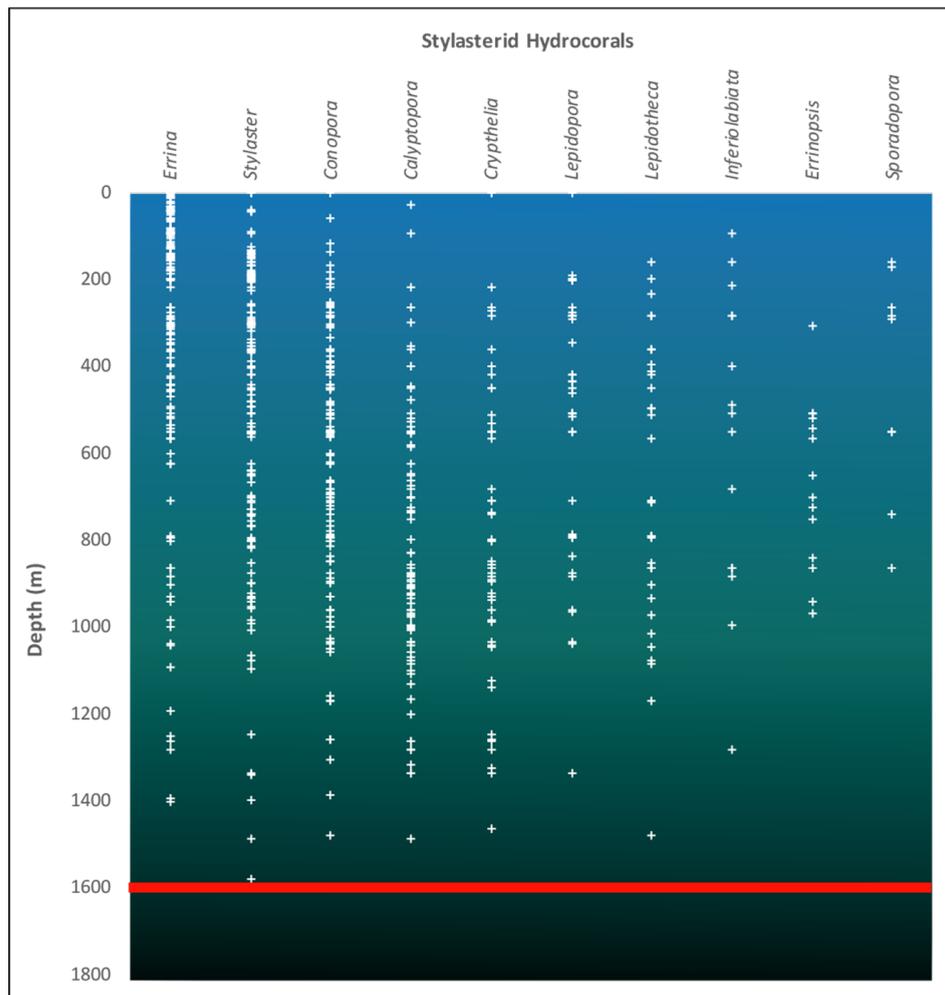


Figure 13: Depth range plot for the ten most abundant stylasterid hydrocoral genera. Source: Tracey & Hjørvarsdóttir, 2019¹⁹⁴. The red line represents the maximum depth New Zealand deep-sea trawlers operate, 1,600m (generally deep-sea trawlers operate between 200m - 800m)¹⁹⁵.

Unprotected Coral and Non-coral Invertebrate Diversity and Distribution

True soft corals, zoantharians and sea pens are not protected in Aotearoa under the Wildlife Act 1953, despite some species being described as ecologically important benthic communities found from shallow waters down to the deep-sea¹⁹⁶.

True soft corals

There is a lack of knowledge on the distribution and ecology of soft corals. Scientists estimate there are at least four endemic species of shallow-water soft corals¹⁹⁷. Some species of non-gorgonian octocorals are found in relatively shallow waters and others can live very deep

¹⁹⁴ Figure 47

¹⁹⁵ Fisheries New Zealand, 2020

¹⁹⁶ Tracey & Hjørvarsdóttir, 2019

¹⁹⁷ Tracey & Hjørvarsdóttir, 2019

beyond 1,300m depth (Figure 14)¹⁹⁸. A large majority of common non-gorgonian octocorals are found between 500m – 1,200m depth (Figure 14).

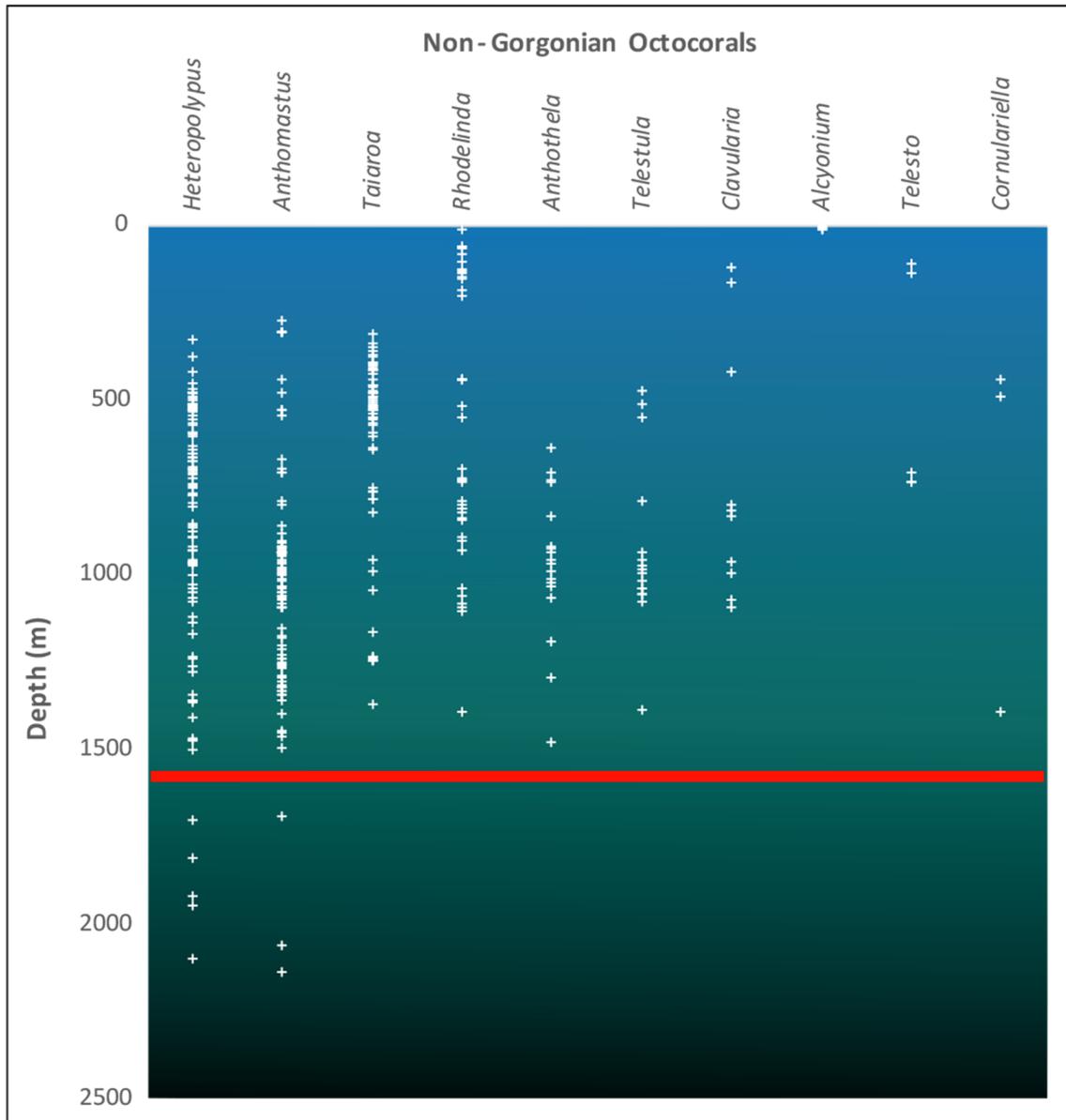


Figure 14: Depth distribution of the most commonly occurring soft corals. Source: Tracey & Hjørvarsdóttir, 2019¹⁹⁹. The red line represents the maximum depth New Zealand deep-sea trawlers operate, 1,600m (generally deep-sea trawlers operate between 200m - 800m)²⁰⁰.

¹⁹⁸ Tracey & Hjørvarsdóttir, 2019
¹⁹⁹ Figure 49
²⁰⁰ Fisheries New Zealand, 2020

Sea pens

Sea pens belong to the order Pennatulacea and are classified as VME indicator species²⁰¹. Sea pens are widely distributed (Figure 15) and are an important soft-sediment stabilizer species that also provides important three-dimensional habitat for fish and other invertebrates²⁰².

Scientists have identified 31 species of sea pens from Aotearoa which equates to about 15% of the global fauna²⁰³. Some are found in shallower waters, usually greater than 50m, while others live in deeper waters down to the abyssal plain. There is no data available from Aotearoa to determine if any species found here are endemic.

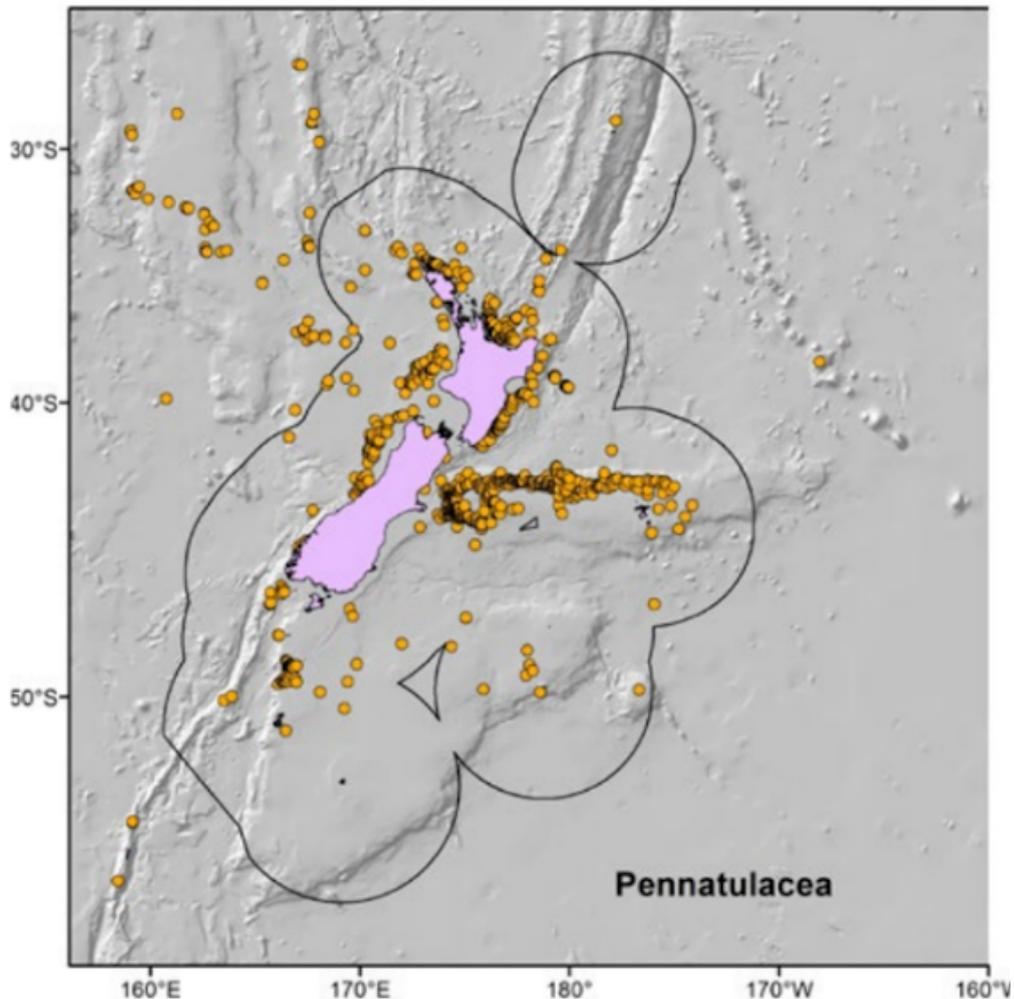


Figure 15: Distribution map of all sea pens (Order Pennatulacea) (Published as supplementary material in Anderson et al. (2016). Source: Tracey & Hjørvarsdóttir, 2019²⁰⁴

²⁰¹ Tracey & Hjørvarsdóttir, 2019

²⁰² Baillon et al, 2012 described in Tracey & Hjørvarsdóttir, 2019

²⁰³ Tracey & Thorvaldsdóttir, 2019

²⁰⁴ Figure 52

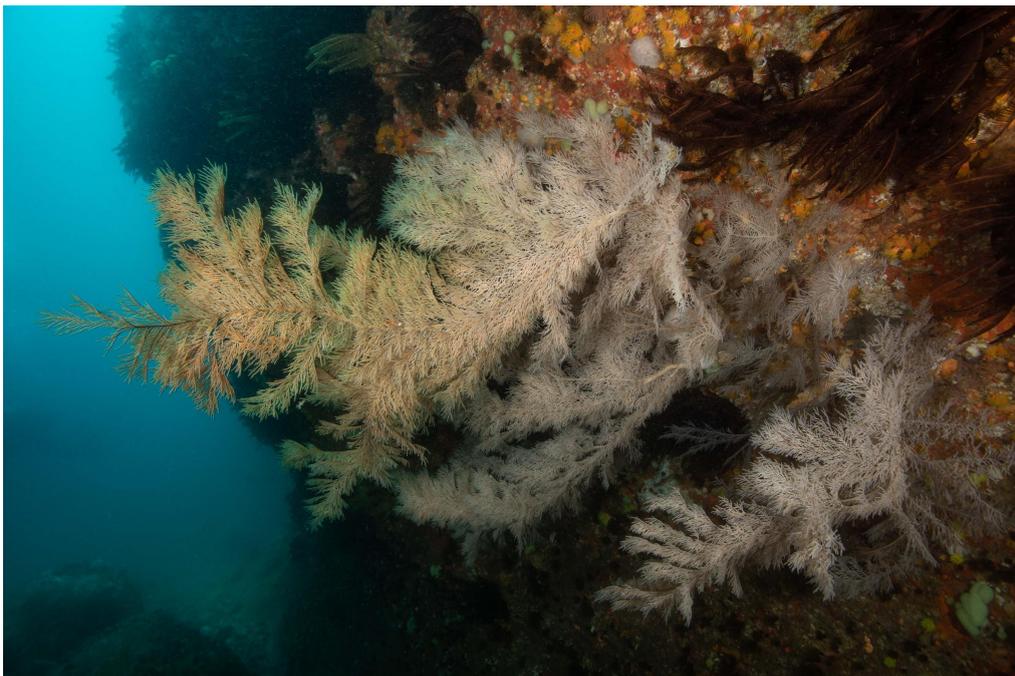
Zoantharia

Scientists have discovered two species of a unique ‘gold coral’ which is actually rare zoantharians. These gold corals are unprotected in Aotearoa waters as they are not listed on Schedule 7A under the Wildlife Act 1953. One species (*Savali sp*) is found between 10 - 300m, whilst the other (*Kulamamanamana haumeaae*) is found in deeper waters²⁰⁵.

Coral core depth distributions

Deep water is classified as 200m and beyond.

In summary, stony corals are predominantly found between 800m - 1,200m deep, black corals are generally found between 200 - 1000m, gorgonian octocorals and hydrocorals generally have a wide depth distribution and can be found down to 1,500m or deeper, while most soft corals (non-gorgonian) also have a broad range from shallow to deeper waters with a max depth of around 1,300m²⁰⁶. Based on the depth distribution plots (Figures 5, 9, 11, 15 and 14) from Tracey & Hjørvarsdóttir (2019), and what is known about the depths different corals live, it can be generalised that most Aotearoa deep-sea coral taxa can be found between 200 - 1,000m, we refer to this as the ‘core depth distribution’ in this report.



© Paul Caiger. Gorgonian Fan (Alcyonacea), Kermadec Islands, New Zealand 2020

Non-coral deep-sea invertebrates diversity

Aotearoa has a diverse range of non-coral deep-sea invertebrates that live on or around seamounts and are often caught as bycatch by deep-sea bottom trawl vessels, these include sponges, sea-stars, starfish, sea-lilies, barnacles, limpets, squid, crabs, prawns, sand dollars, sea

²⁰⁵ Tracey & Hjørvarsdóttir, 2019

²⁰⁶ Tracey & Hjørvarsdóttir, 2019

urchins and deep-sea fish species²⁰⁷. It is highly likely some of these non-coral deep-sea invertebrates found within Aotearoa’s EEZ are endemic, but there is a lack of published literature on this.

Over the past three years, scientists in Aotearoa have identified 135 taxa new to science from fisheries bycatch samples collected between 1994 to 2020 (Figure 16 and 17), including 128 new species, 6 new genera and one new family, as part of an invertebrate identification project²⁰⁸.

Most of the samples containing new taxa were collected on research trawl surveys (219 of 318). Samples collected by observers on commercial fishing vessels, which contained new taxa, and are currently able to be attributed to a fishery, are shown in Table 3²⁰⁹.

This research shows that some of the very areas that scientists are discovering new species from are also the same areas being bottom trawled. This means there is the potential for New Zealand vessels to destroy endemic or rare invertebrate species before they can be discovered and identified. This highlights the importance of taking a precautionary approach and protecting these important deep-sea ecosystems and associated biodiversity, like those around seamounts.

Table 3: The number of new invertebrate taxa identified in Schnabel et al. (2021) broken down by target fishery samples were collected in, Source: Unpublished data from FNZ, 2021.

Target fishery species	Total number of samples containing new taxa
Barracouta	7
Bass groper	6
Hoki	8
Hapuka/Bass	3
Jack mackerel	1
Ling	1
Orange roughy	13
School shark	4
Scampi	5
Snapper	1
Squid	20
Oreos (SSO)	3
Tarakihi	1
Grand Total	73

²⁰⁷ Ministry of Fisheries, 2011

²⁰⁸ Schnabel et al 2021

²⁰⁹ Unpublished data from Fisheries New Zealand, 2021

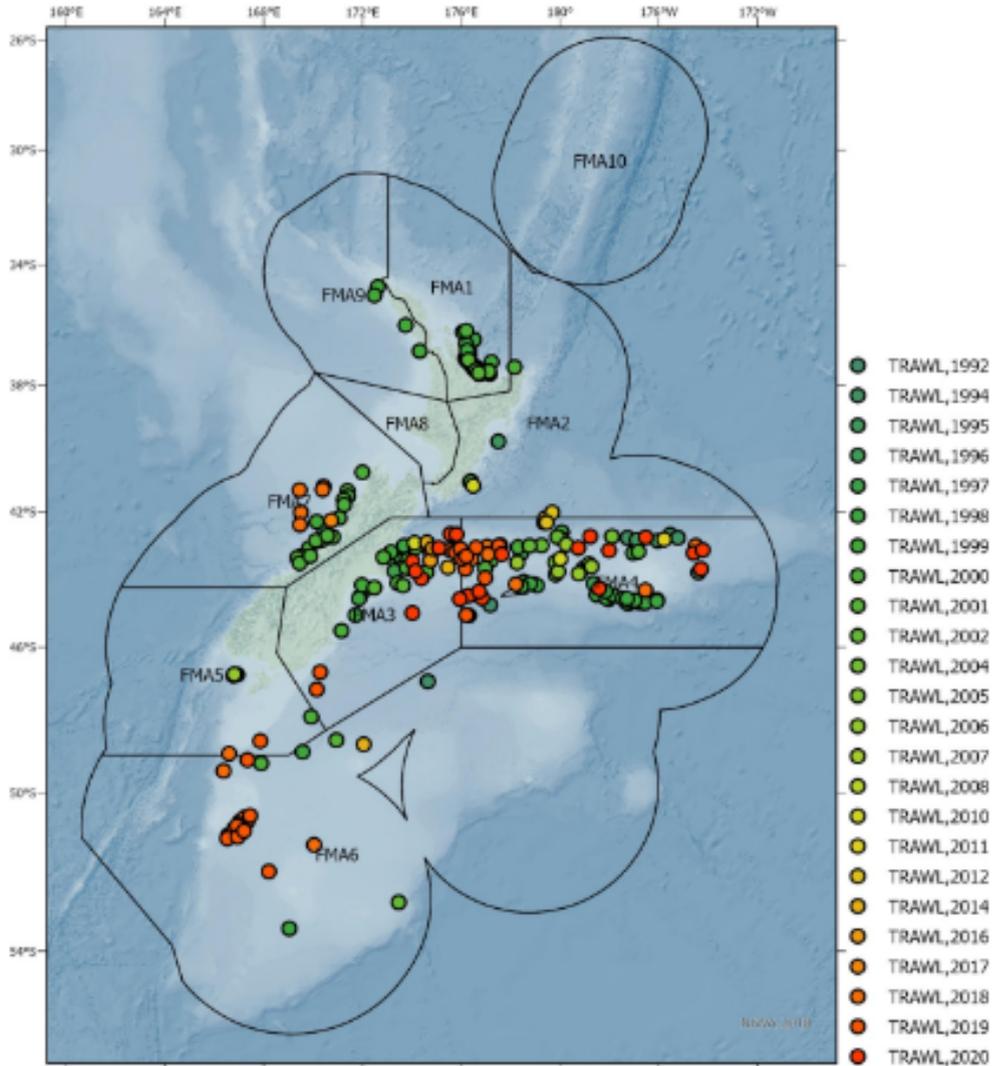


Figure 16: A plot of the location of invertebrate samples processed in Schnabel et al (2021) collected from research trawl surveys. Coloured dots represent collection year with the oldest samples in dark green, and the most recent in dark red. Source: Schnabel et al (2021)

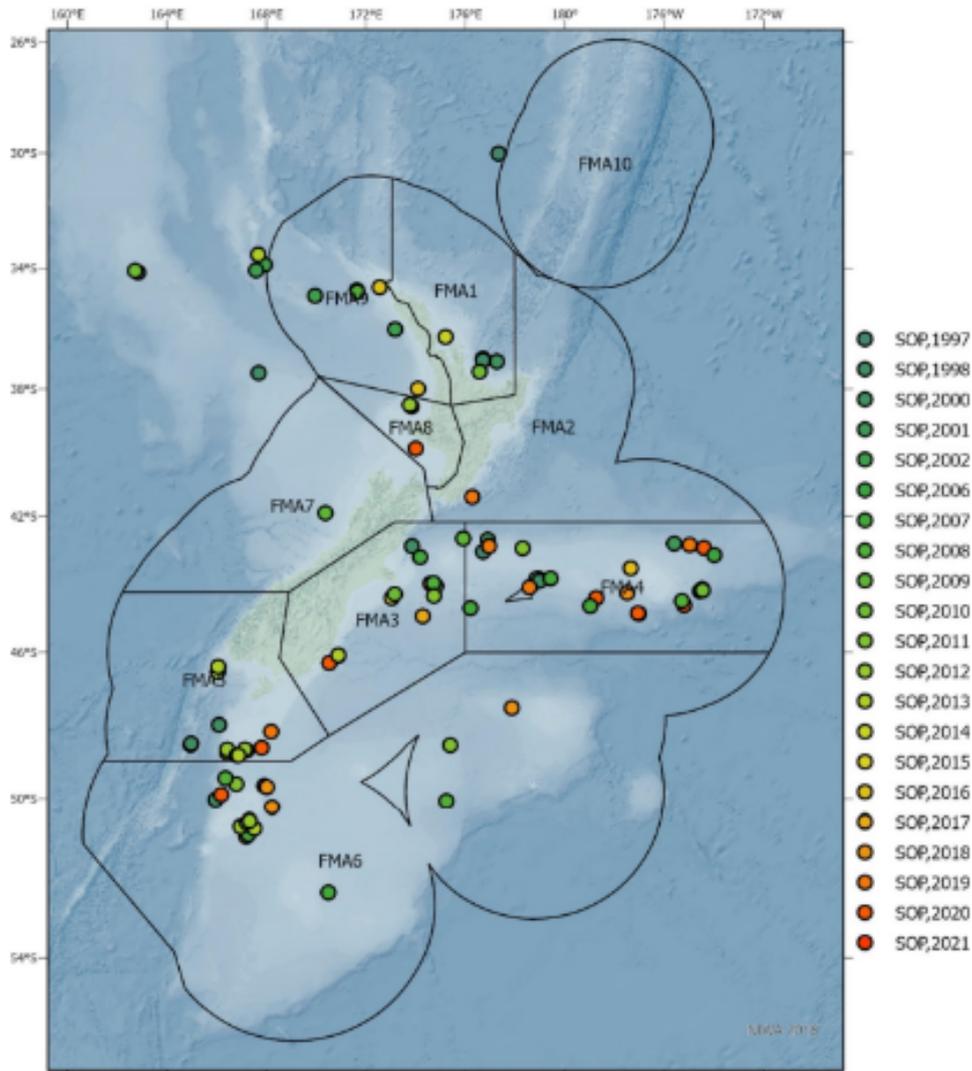


Figure 17: A plot of the location of invertebrate samples processed in Schnabel et al (2021) collected from observers on commercial vessels. Coloured dots represent collection year with the oldest samples in dark green, and the most recent in dark red. Source: Schnabel et al (2021)

More research is needed to describe deep-sea soft sediment fauna. Researchers have looked at the effects of trawling on benthic mega-epifaunal and macro-infaunal communities on the Chatham Rise and found that bottom trawling can significantly modify the structure and functional composition of epifaunal communities in deep-sea soft sediment environments, by reducing the density of taxa and diversity²¹⁰. This modification of deep-sea soft sediment such as the loss of epifaunal diversity is concerning as this could result in a loss of ecosystem function such as reduction in the availability of species for benthic-feeding demersal fish²¹¹.

²¹⁰ Bowden & Luduc, 2017

²¹¹ Bowden & Luduc, 2017

Section 3: Overview of seamount biodiversity in the South Pacific

The Pacific Ocean has the highest number of seamounts of all the world's oceans²¹² (Figure 18).

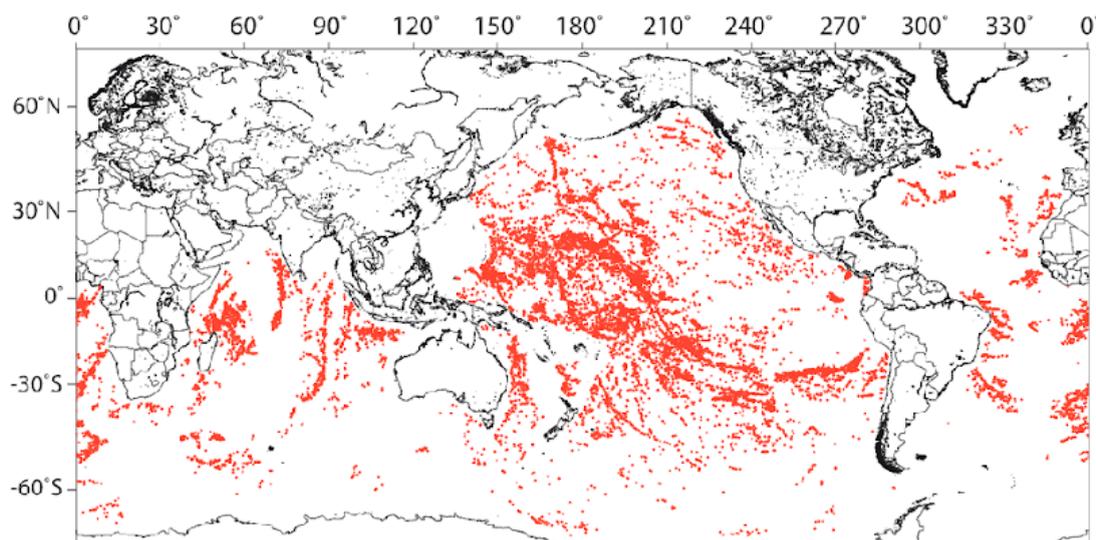


Figure 18: Global distribution of 118,882 seamounts described by Wessel *et al.*, (2010) from Lyer *et al.* (2013).

The high seas, the area of ocean beyond national jurisdiction, comprises two-thirds of the global ocean and covers nearly half the planet. These areas beyond national jurisdiction are divided up into regional fisheries management areas. The South Pacific Regional Fisheries Management Organisation (SPRFMO) area (Figure 19) extends all around Aotearoa's EEZ.

The United Nations General Assembly (UNGA) has set out resolutions²¹³ to globally minimise the effects of seafloor fisheries on the high seas. To meet these resolutions, in some oceans around the world little or no bottom trawling occurs on seamounts in the high seas. For example, in September 2021, at the 43rd annual meeting of the Northwest Atlantic Fisheries Organisation (NAFO) NAFO agreed to fully protect all seamounts and other features less than 4,000m depth from bottom trawling²¹⁴. NAFO seamount closures cover an area approximately 100,000 square kilometres²¹⁵, that's almost the same size as the whole of the North Island²¹⁶.

²¹² Kitchingman *et al.* 2008

²¹³ UNGA Resolution 61/05: The UN General Assembly, in Resolution 61/105, adopted in 2006, called on "States to take action immediately, individually and through regional fisheries management organizations and arrangements, and consistent with the precautionary approach and ecosystem approaches, to sustainably manage fish stocks and protect vulnerable marine ecosystems, including seamounts, hydrothermal vents and cold water corals, from destructive fishing practices, recognizing the immense importance and value of deep sea ecosystems and the biodiversity they contain" (Paragraph 80)

²¹⁴ NAFO, 2021. Press release: NAFO agrees to further measures to protect Vulnerable Marine Ecosystems (VMEs) at its 42rd Annual Meeting [pressrelease_AM2021.pdf \(nafo.int\)](https://www.nafo.int/pressrelease_AM2021.pdf). Retrieved 26 September 2021

²¹⁵ DSCC, 2021. Press release 24/9/21: New Protections for Fragile Deep-sea Ecosystems Agreed by Northwest Atlantic Fisheries Organisation. [New protections for fragile deep-sea ecosystems agreed by Northwest Atlantic Fisheries Organisation - Deep Sea Conservation Coalition \(savethehighseas.org\)](https://www.savethehighseas.org/news/new-protections-for-fragile-deep-sea-ecosystems-agreed-by-northwest-atlantic-fisheries-organisation). Retrieved 26 September 2021

²¹⁶ North Island Area 113,729km² – Source: New Zealand Official Yearbook, 2000 from Statistics New Zealand [Quick Facts - Land and Environment : Geography - Physical Features \(archive.org\)](https://www.stats.govt.nz/quick-facts-land-and-environment-geography-physical-features). Retrieved 26 September 2021

In the South Pacific, largely due to the New Zealand Government’s influence at the SPRFMO the vulnerable deep-sea communities and ecosystems on seamounts and similar features are still at risk from benthic disturbance, as a fleet of New Zealand owned trawlers are actively bottom trawling (Figure 19).

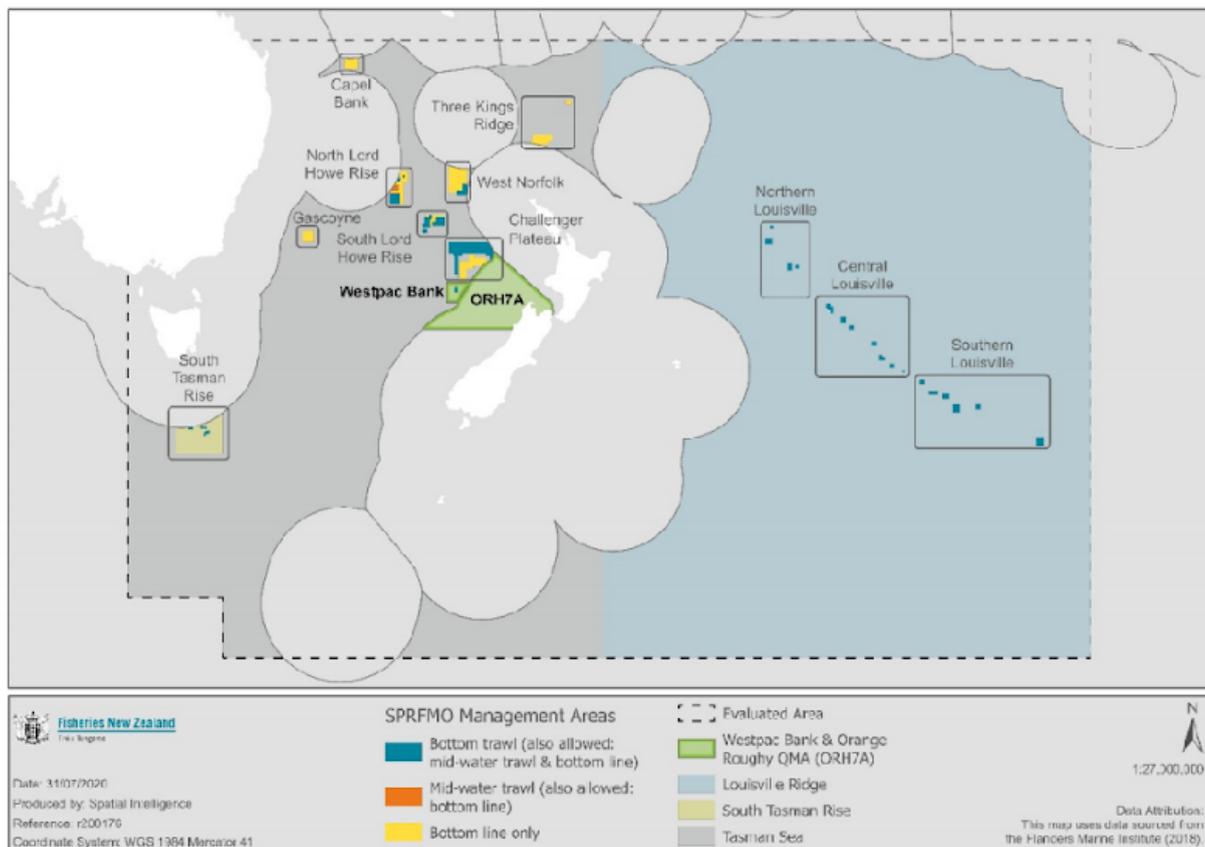


Figure 19: SPRFMO management area showing areas New Zealand bottom trawlers operate since 2006 overlaid with the CMM03-2019 management. Source: Fisheries New Zealand, 2021

South Pacific Regional Fisheries Management Organisation:

Within the SPRFMO area New Zealand is the only country currently bottom trawling on seamounts primarily targeting orange roughy²¹⁷ and alfoncino. As of 1 July 2021, Fisheries New Zealand authorised six New Zealand owned bottom trawl vessels to fish in this predominantly seamount-focused fishery²¹⁸. Figure 20 shows the SPRFMO regulatory area (blue) and known seamounts (red dots). Figure 21 shows SPRFMO regulatory area (purple line) and known ‘fishable’²¹⁹ seamounts (yellow dots)²²⁰.

²¹⁷ Orange roughy is the main target species and has made up 67-99% of the total New Zealand bottom trawl catch since 2002 with tonnages ranging from 301 to 2 578 tonnes: Source Fisheries New Zealand, 2021

²¹⁸ All six vessels belong to companies or skippers that have been convicted in the past year of trawling illegally in closed areas

²¹⁹ ‘fishable’ means the seamount is within the fishable depth of 0 - 1,500m

²²⁰ Fuller et al. 2020

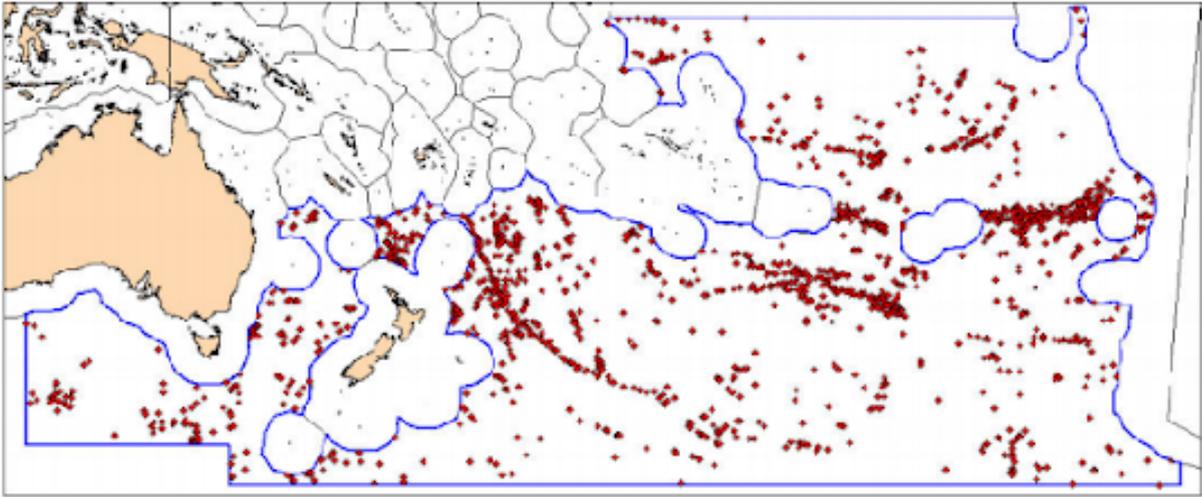


Figure 20: Geographic distribution of validated seamounts and similar under-water features within the SPRFMO regulatory area. Source: Allain & Clark, 2008



© CSIRO, 2018. A high diversity of stony corals and sponges. From seamount biodiversity study in the South West Pacific, on Tasmanian seamounts.

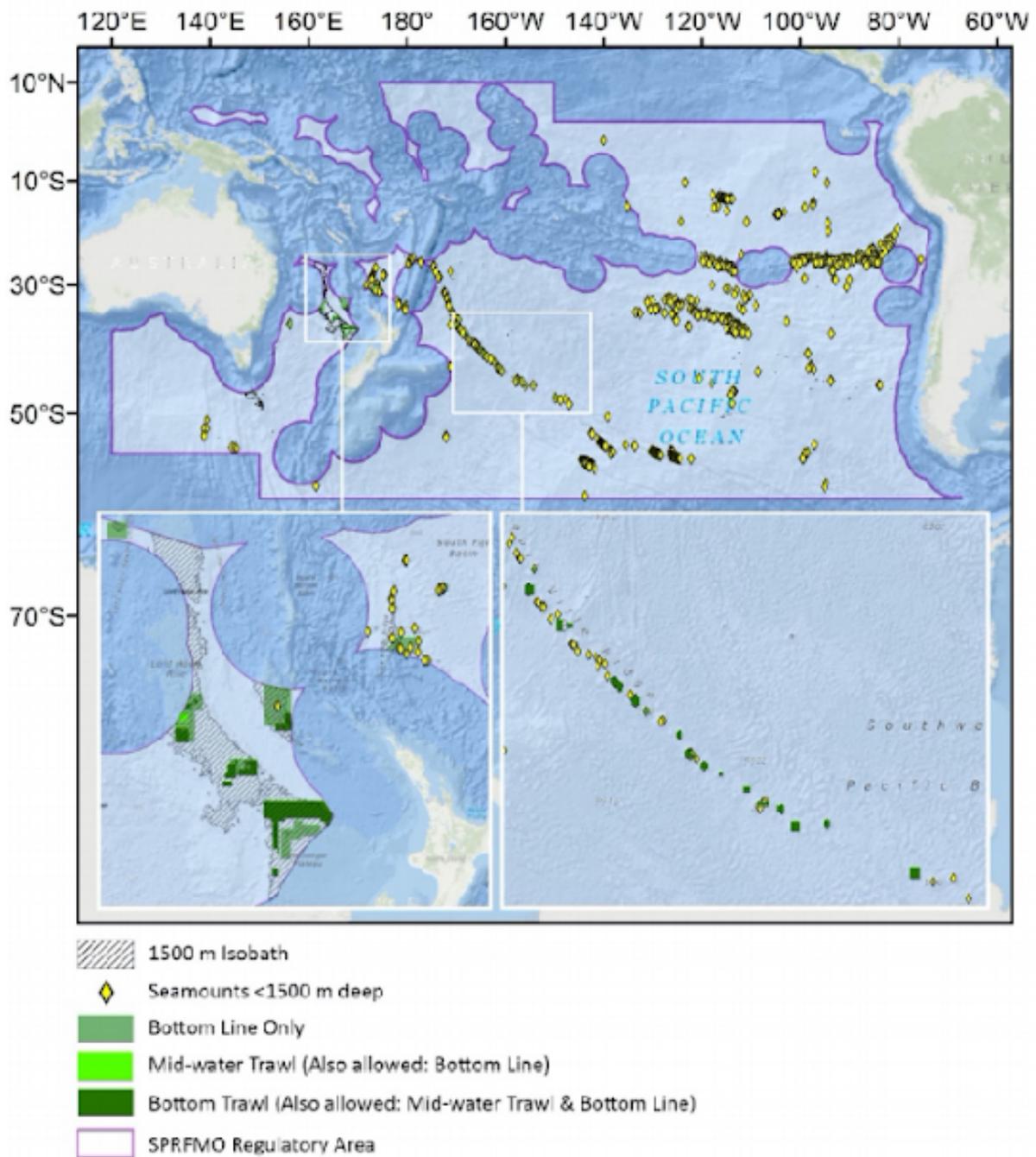


Figure 21: Map of the SPRFMO regulatory area showing seamounts within 'fishable' depth (<1,500m). Source: Fuller et al., 2020.

South Pacific Seamounts Biodiversity

Identifying the locations of seamounts within SPRFMO is important. Watling and Auster (2017) reviewed seamount locations and the presence of VME indicator species from within the SPRFMO area and found that “*all seamounts that have so far been surveyed by cameras, either towed or mounted on maneuverable submersible vehicles, have been found to have abundant VME indicator species*”

(including xenophyophores on sandy areas) distributed on their sides and summits²²¹. Watling and Auster (2017) also conclude “the distribution of VME indicator species is far more extensive than fishery bycatch data would suggest. The lack of VME species in virtually all trawl catches on seamounts is due entirely to their fragility and brittleness. While some large pieces of VME species encountered might be retained in the coarse meshes of a commercial trawl net or snagged on fixed gears such as longlines, the biomass would rarely be high enough to trigger an official “encounter” as most are surely broken into small pieces and fall to the seafloor before reaching the deck of the ship²²². This reinforces the assumption that all seamounts, even acknowledging that distributions across seamounts can vary, that occur within the biological depth ranges of VME indicator species, like deep-sea corals, should be considered to contain VMEs and therefore warrant full protection in order to meet international obligations such as UNGA resolutions²²³

SPRFMO has some controls in place which New Zealand trawlers must follow (refer to Figure 19 & Figure 21) to reduce the impact on VMEs from bottom trawling including 100% observer coverage, move-on rules²²⁴, and requiring pre-trawling impact assessments. New Zealand has argued for ‘spatial management’, which by that mean that SPRFMO allows the destruction of VMEs on the basis that there are other VMEs found elsewhere²²⁵. These measures are not adequate as they do not comply with the UNGA resolutions to fully prevent all significant adverse impacts (SAI) on VMEs.

There has been no comprehensive assessment of the deep-sea benthic biodiversity found within SPRFMO and specifically associated with SPRFMO ‘fishable’ seamounts besides limited research, observer bycatch data and predictive habitat suitability modelling. This means it is impossible to determine the level of endemism and presence of rare or unique species. What is known is that the bycatch of corals and other vulnerable deep-sea species by New Zealand bottom trawl vessels on seamounts in the South Pacific is often high.

New Zealand vessels reported benthic bycatch exceeding 1,000 kg per tow in places like the Three Kings Ridge and Wanganella Bank of bushy hard corals²²⁶. One vessel was reported to have taken 5,000 kg of coral in a single tow while bottom trawling on the Louisville Ridge Seamount Chain²²⁷. As described earlier the bycatch that ends up at the surface observed is not the total destroyed, much more is destroyed on the seabed²²⁸. This means in some of the highest bycatch incidents by New Zealand trawlers in the South Pacific, these incidents may have actually destroyed between 122 and 1,700 tonnes of deep-sea corals and other vulnerable benthic species from single trawls²²⁹.

²²¹ Watling and Auster, 2017

²²² Watling and Auster, 2017

²²³ e.g.61/50 in 2006 called “upon States to take action immediately, individually, and through regional fisheries management organizations and arrangements, and consistent with the precautionary approach and ecosystem approaches, to sustainably manage fish stocks and protect vulnerable marine ecosystems [VMEs], including seamounts, hydrothermal vents and cold water corals, from destructive fishing practices, recognizing the immense importance, and value of deep-sea ecosystems and the biodiversity they contain?”

²²⁴ Move-on rules require a vessel to move one nautical mile away from where it is trawling if a certain amount of vulnerable indicator species are observed. VME indicator taxa include stony corals, soft corals, black corals

²²⁵ DSCC, 2020

²²⁶ Cryer & Geange, 2018

²²⁷ Maximum bycatch from Cryer and Geange, 2018 and Cryer et al. 2018.

²²⁸ Pitcher et al. 2019

²²⁹ A trawl bringing up 1.133 mt of bycatch (Three Kings Ridge and Wanganella Bank) indicates 122 - 385 mt destroyed on the seabed, while 5.001 mt bycatch trawled up from Louisville Ridge indicates 540 - 1,700 mt destroyed on the seabed. Sources: Cryer and Geange, 2018 (bycatch) and Pitcher et al. 2019 (destruction on the seabed)

New Zealand owned bottom trawlers fishing for orange roughy in the South Tasman Rise have reportedly caught a large diversity of coral species, often dominated by reef-forming stony corals such as *Solenastrea variabilis*²³⁰.

One study looked at fish and macroinvertebrates diversity from seamounts in the Tasman and Coral Seas in the Southwest Pacific (taxa samples were taken from seamounts within and outside of SPRFMO), Figure 22²³¹. Of the 516 taxa samples taken from 6 seamounts along the Norfolk Ridge, 36% of invertebrates were species new to science and were not found in the surrounding areas indicating high levels of endemism²³². Of the 108 taxa samples taken from 4 seamounts on the Lord Howe Rise, 31% were new to science and of the 297 taxa samples taken from 14 seamounts south of Tasmania (within Australia's EEZ) between 16 - 33% were species new to science²³³.

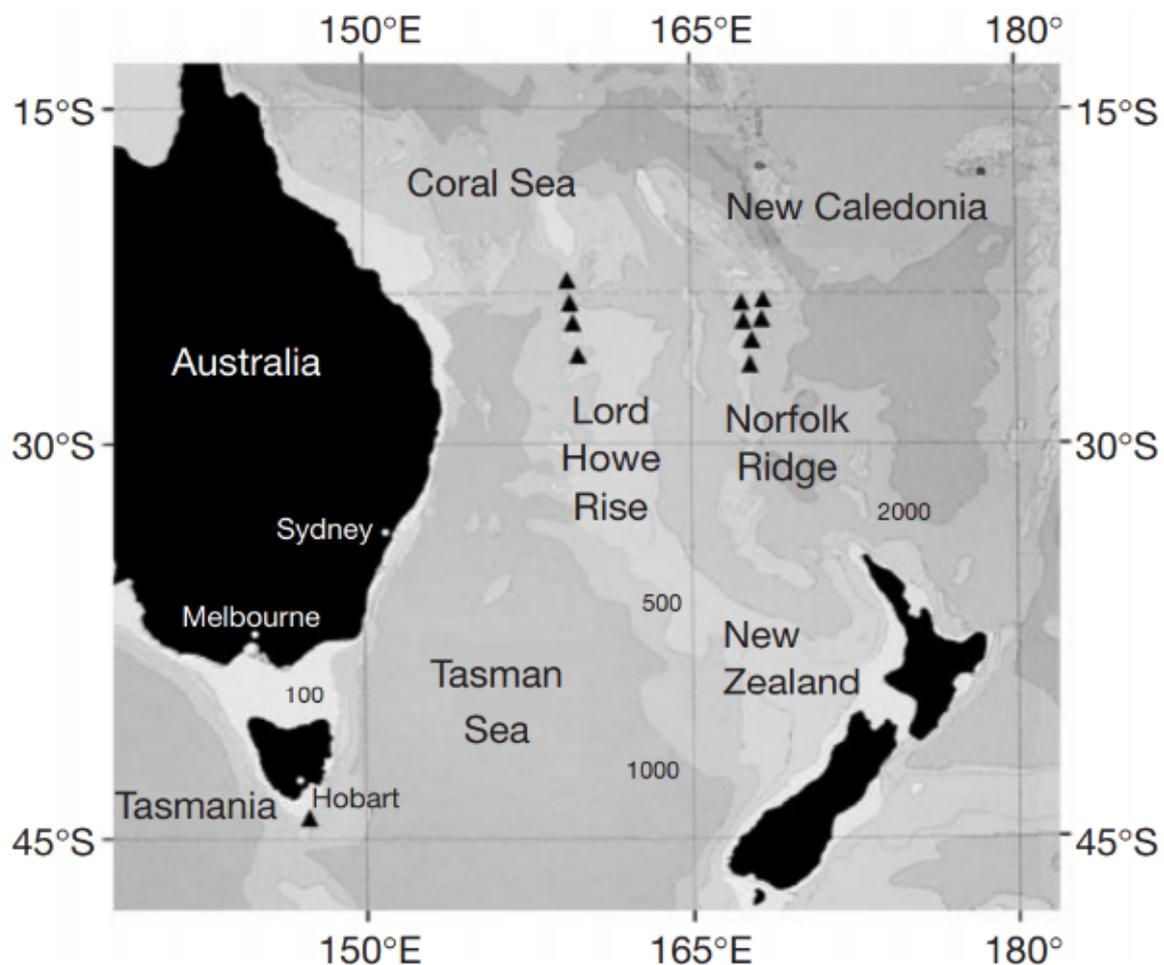


Figure 22: Location of seamount sampling sites (triangles) from Richer de Forges *et al.* (2000)

VME bycatch by New Zealand fishers is not a historic issue. New Zealand is still reporting high bycatch rates of VME indicator species to SPRFMO. In 2020, a New Zealand vessel exceeded

²³⁰ Anderson & Clark, 2003
²³¹ Richer de Forges et al 2000
²³² Richer de Forges et al 2000
²³³ Richer de Forges et al 2000

the amount of VME indicator species that can be trawled up in a single trawl within the North Lord Howe region²³⁴. It's reported the incident took place on the Lord Howe Rise plateau area (Figure 23) and that between 18 - 20kgs²³⁵ of Alcyonacea Gorgonian coral was brought up²³⁶. New Zealand has not provided the 2021 meeting of the Scientific Committee²³⁷ with any further details around the encounter, such as vessel details²³⁸, if the bycatch was identified to a lower taxonomic level²³⁹ and whether or not the bycatch represented multiple species, or if a sample of the bycatch was brought back to New Zealand for any further identification²⁴⁰. New Zealand is recommending to reopen the area to bottom trawling and states based on their assessment that “bottom trawling is unlikely to cause SAI [significant adverse impacts] to VMEs at the FMA [fisheries management area] scale. While it cannot be completely excluded that reopening the area to trawling could result in encounters with other VMEs at the sub-tow spatial scale, this risk is considered to be low”²⁴¹.

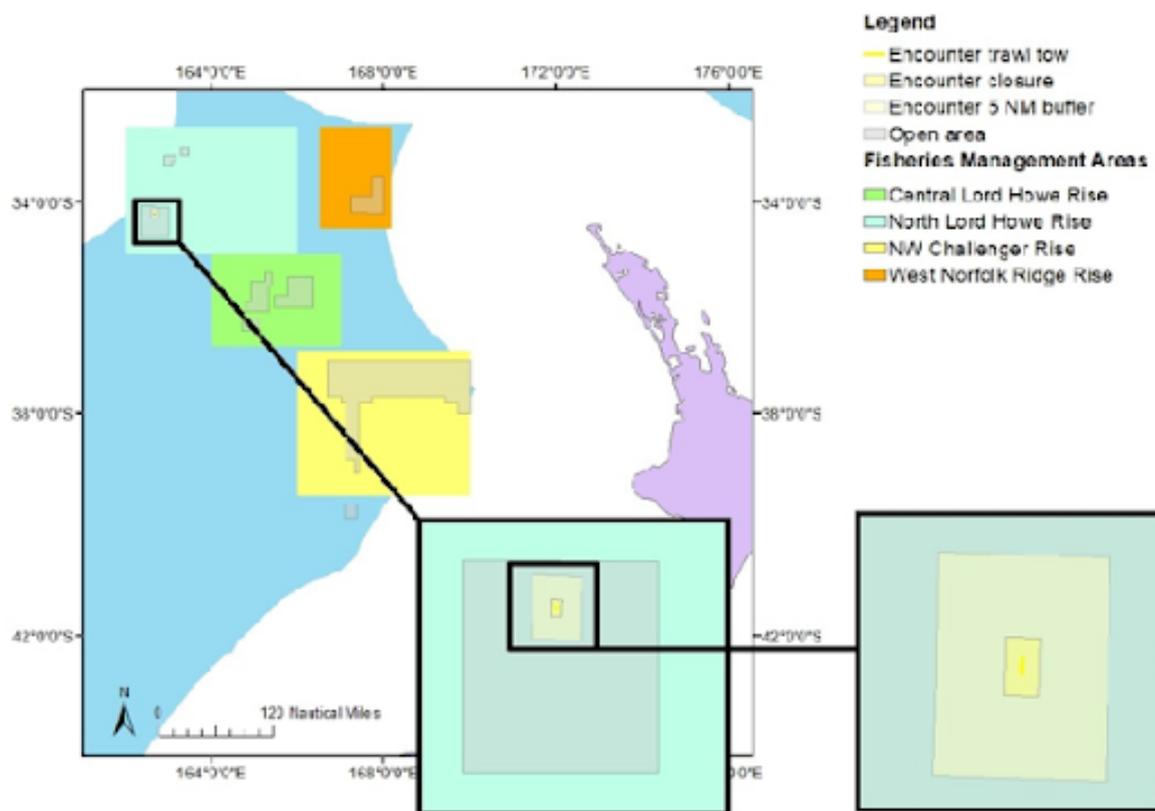


Figure 23: Location of the 2020 VME encounter by New Zealand bottom trawler within the North Lord Howe Rise FMA (teal polygon). Source: Milardi et al. 2021

Like in Aotearoa, there is a critical information gap which is that most New Zealand fisheries observers can't accurately identify benthic species, as they simply are not taxonomic experts. This

²³⁴ Milardi et al. 2021

²³⁵ Threshold weight is 15kgs

²³⁶ Milardi et al. 2021

²³⁷ The 9th meeting of the SPRFMO Scientific Committee held virtually 27th September to 2nd October, 2021

²³⁸ New Zealand states there is an ongoing compliance investigation, see Milardi et al. 2021

²³⁹ Most bycatch is reported at the taxonomic level of Order or higher

²⁴⁰ Greenpeace has requested further information from FNZ around the details of the SPRFMO VME encounter, but so far all requests have been denied.

²⁴¹ Milardi et al. 2021b – note this reference is now referring to the updated 'SCDW09_rev1' New Zealand 2020 VME encounter review paper

means often benthic-bycatch taxa is identified at the Order (or higher) taxonomic level rather than species level. The consequence of this is that we do not know what levels of endemism or cryptic benthic diversity is being caught as bycatch by New Zealand trawlers.

Based on the benthic bycatch from New Zealand bottom trawlers operating within the SPRFMO area that could be identified, it is estimated that New Zealand vessels have destroyed samples of at least 281 genera and 231 species of VME taxa²⁴². The last five years of observed benthic bycatch is shown in Table 4.

The bycatch from New Zealand vessels that has been identified is comparable to the list of known VME taxa²⁴³ identified from a study area within SPRFMO. Of the 5,300 taxa records assessed from within the study area, 281 genus and 231 species of VME taxa were identified²⁴⁴. These records are from multiple data sets, including research trawls and fisheries bycatch. Of this, 173 genera and 182 species of VME taxa were identified from fishable depths (<1,400m)²⁴⁵. Importantly, this highlights that most of the genera (63%) and species (79%) identified as VME taxa found within the SPRFMO area occur within fishable depths (< 1400m)²⁴⁶.

This review of VME taxa is likely to be an underestimate of the VME species that live within the entire SPRFMO region because a) only a subset of the region was included in the sample and research has found that seamounts even over short distances can have distinct and unique macroinvertebrate communities (e.g. corals and sponges)^{247,248}, b) not all of the study samples were able to be identified and c) not all samples had depths recorded. It is important to note that this study is likely to significantly underestimate naturally rare VME species as evidenced by the high percentage (41%) of taxa with only a single observation in the dataset²⁴⁹. Additionally, Richer de Forges et al. (2000) found a linear relationship between the number of species recorded from each seamount and the number of samples obtained, which supports that greater sampling effort will result in greater species richness²⁵⁰.

These reports confirm within the SPRFMO area there are likely hundreds to thousands of VME species, many likely to be rare and endemic, which supports that this area is biologically very important and warrants full protection from benthic disturbance methods, such as bottom trawling in order to comply with the UNGA resolutions to fully prevent all SAI on VMEs.

²⁴² Fisheries New Zealand, 2021

²⁴³ The review only reported on the 15 VME groups identified in SPRFMO SC7-DW13

²⁴⁴ Geange et al. 2020

²⁴⁵ Geange et al. 2020

²⁴⁶ Geange et al. 2020

²⁴⁷ Rowden and Clark, 2004

²⁴⁸ Richer de Forges et al 2000

²⁴⁹ Geange et al. 2020.

²⁵⁰ Richer de Forges et al 2000

Table 4: Benthic bycatch reported by observers on New Zealand bottom trawlers within the SPRFMO area between 2016 – 2020. Source)²⁵¹ Source: Fisheries New Zealand, 2021

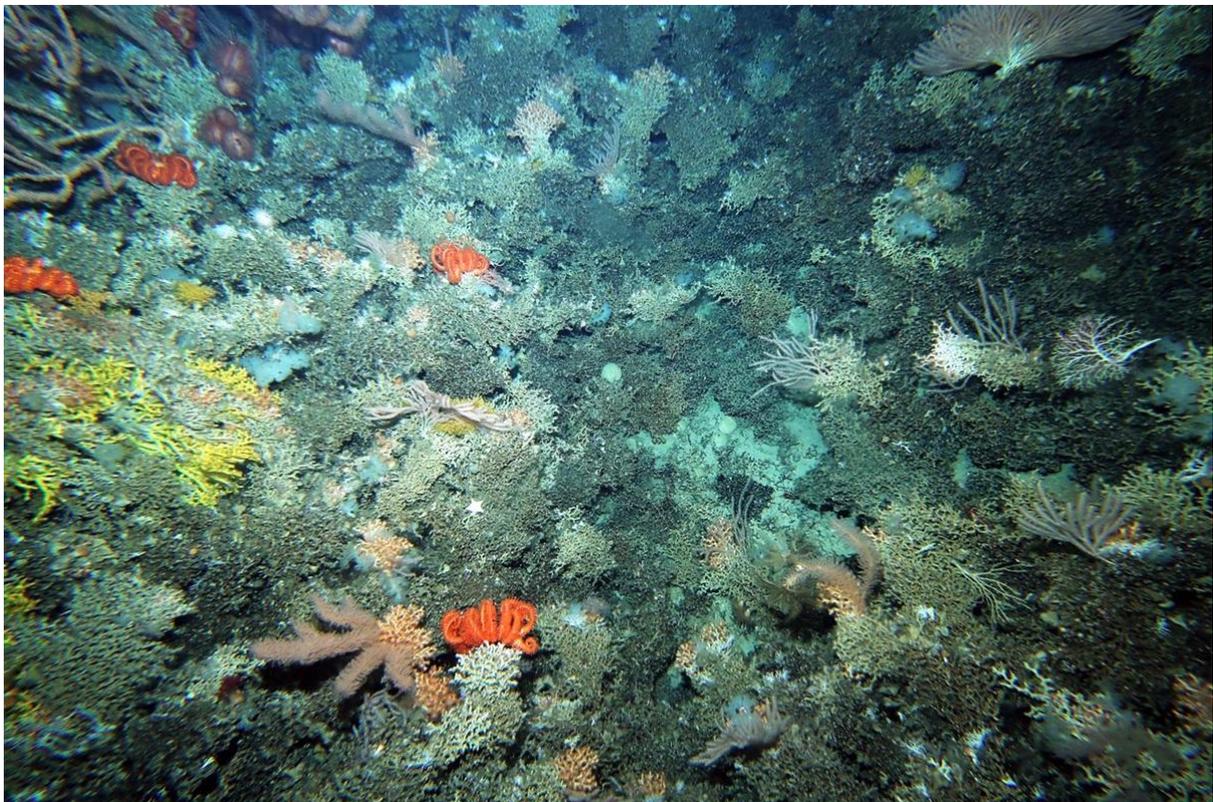
Taxon	2016	2017	2018	2019	2020
BOTTOM TRAWL					
Arthropoda					
Hexanauplia (Barnacles)	1.7 (4)	0 (0)	0.6 (3)	1.9 (3)	0 (0)
Malacostraca (Crabs, prawns)	6.1 (5)	35.9 (18)	2.9 (4)	8 (3)	3 (2)
Pycnogonida (Sea spiders)	0 (0)	1.0 (1)	0 (0)	1.0 (1)	0 (0)
Brachiopoda (Lamp shells)	0 (0)	1.0 (1)	0 (0)	0 (0)	0 (0)
Bryozoa (Lace corals)	0 (0)	0 (0)	0.4 (2)	0 (0)	0 (0)
Chordata					
Ascidiacea (Sea squirts)	0 (0)	20 (4)	0 (0)	0 (0)	0 (0)
Thalacea (Tunicates)	4.7 (8)	12.1 (13)	0 (0)	0 (0)	14 (4)
Cnidaria					
Anthozoa (Anemones, corals, sea pens) ¹	546.7 (164)	3369.5 (104)	1252.53 (139)	36.2 (23)	24 (24)
Actinaria (Anemones)	906.3 (158)	902.5 (159)	989.1 (80)	107.8 (29)	83.5 (34)
Alcyonacea (Soft corals)	0 (0)	0 (0)	0 (0)	0 (0)	0.2 (2)
Antipatharia (Black corals)	43.7 (67)	37.9 (58)	46.9 (69)	13.7 (31)	2.5 (13)
Gorgonian Alcyonacea (Tree-like forms, sea fans, sea whips, bottlebrush) ²	33.6 (51)	78.9 (73)	45.4 (64)	33.5 (29)	4.3 (7)*
Pennatulacea (Sea pens)	3.3 (15)	18.3 (28)	8.1 (11)	2.1 (10)	1.2 (8)
Scleractinia (Stony corals) ³	32 (18)	840.1 (61)	47.5 (31)	8.4 (6)	62.8 (40)
Zoantharia (Hexacorals)	194.4 (131)	93.3 (53)	100.9 (57)	1 (1)	9.9 (7)
Hydrozoa (Hydroids) ⁴	3 (2)	0.3 (2)	17.3 (11)	21 (21)	0 (0)
Stylasteridae (Hydrocorals)	0 (0)	3.7 (4)	2.3 (3)	0 (0)	0.3 (1)
Echinodermata					
Asterocidea (Starfish)	1.0 (2)	11.7 (8)	4.0 (3)	30.0 (12)	3.3 (5)
Brisingida (Armless' stars)	2.1 (5)	0 (0)	2.3 (2)	0 (0)	0.1 (1)
Crinoidea (Sea lillies)	3.8 (7)	1.7 (13)	3.5 (6)	20 (20)	0 (0)
Echinoidea (Sea urchins)	1037.2 (38)	215.9 (45)	52.5 (6)	80 (3)	19.3 (13)
Holothuridea (Sea cucumbers)	17.5 (14)	19.6 (7)	0 (0)	0 (0)	7.6 (03)
Ophiuroidea (Brittle stars)	253.0 (70)	11.3 (16)	2.0 (1)	3.0 (3)	1.6 (3)
Mollusca					
Bivalvia (Mussels, clams)	0.2 (2)	0.7 (3)	0.1 (1)	0 (0)	0 (0)
Gastropoda (Snails, whelks, tritons)	1.8 (5)	6.1 (7)	0 (0)	0 (0)	0.2 (1)
Phaeophyceae (Brown algae)	10.0 (6)	0 (0)	0 (0)	0 (0)	0 (0)
Porifera (Sponges) ⁵	191.9 (125)	428.5 (97)	168.5 (73)	15.4 (16)	22.9 (27)
Sipuncula (Peanut worms)	0 (0)	3 (1)	0 (0)	0 (0)	0 (0)
Unidentified	15.7 (7)	5.7 (5)	4 (4)	0 (0)	0 (0)

²⁵¹ Source: Fisheries New Zealand, 2021: “Weight in kg (and number of positive reports). Where taxonomic resolution allows, bycatch is presented at the Order level, otherwise at the Class or Phylum level. Row colours refer to VME indicator taxa included in CMM 03-2019 (purple), other VME indicator taxa added to CMM02-2021 (orange), and other benthic bycatch taxa (white)* This does not include the encounter event or weight. 1 includes taxa other than Actinaria, Gorgonian, Alcyonacea, Scleractinia, Antipatharia, Pennatulacea, Zoanatharia, 2 Includes all Gorgonacea within the sub-orders Halaxonia, Calcaxonia and Scleraxonia, 3 Includes all taxa within the following genera: Solenosmilia; Goniocorella; Oculina; Enallopsammia; Madrepora; Lophelia, 4 Includes taxa other than Stylasteridae, and 5 Includes all Porifera within the classes Demospongiae and Hexactinellidae”

Australian waters and the high seas:

New Zealand-owned vessels also bottom trawl in the Indian Ocean under a Cook Island flag, and seasonally in Australia under a New Zealand flag, predominantly around Tasmania.

Benthic macrofauna research of small seamounts (300m - 600m high, with peaks ranging between 660m - 1,700m depth) south of Tasmania found high levels of fauna diversity²⁵². One study found 262 species of invertebrates and 37 species of fishes and that the seamounts surveyed that had not been heavily fished and were shallower than 1,400m had dense and diverse invertebrate fauna including scleractinian corals, gorgonian and antipatharian corals, hydroids, sponges and sea stars²⁵³. This study supports similar findings to New Zealand that many seamounts have endemic, rare or even undiscovered species. Koslow et al (2001) found 24 - 43% of the invertebrate species sampled south of Tasmania were new to science and between 16 - 33% were restricted to the sampled seamount environment.



© CSIRO, 2018. Three kinds of stony corals and orange brisingid seastars. From seamount biodiversity study in the South West Pacific, on Tasmanian seamounts.

It is difficult to determine the exact impact New Zealand owned vessels are having in Australian waters, like the number of deep-sea coral species and other rare or endemic invertebrates that are being destroyed, as these vessels are not required to report coral bycatch in a similar way to within Aotearoa's EEZ.

²⁵² Koslow et al. 2001

²⁵³ Koslow et al. 2001

What is known, is that seamounts off Tasmania support a diverse fauna associated with deep-sea reefs, formed by scleractinian corals like *Solemosmilia variabilis*²⁵⁴. These deep-sea scleractinian coral reefs are vulnerable to physical disturbance like bottom-trawling²⁵⁵ and constitute VMEs. Most deep-sea corals surveyed around these Tasmanian seamounts live in water depths shallower than 1,500m²⁵⁶. This depth distribution makes them particularly vulnerable to deep-sea bottom trawling, which also occurs in waters shallower than 1,500m²⁵⁷.

Deep-sea bottom trawlers operating on and near these Tasmanian seamounts, including New Zealand owned vessels are primarily targeting orange roughy²⁵⁸. Orange roughy are typically found between 700m – 1,200m²⁵⁹ on or around seamounts in the region²⁶⁰. Williams et al. (2020) looked at the impact of bottom trawling on deep-sea coral reefs on 51 different seamounts and detected trawling damage on 88% (45 of 51) of seamounts²⁶¹. This study found that depth of the seamount significantly affected the severity of the trawl damage. Shallower seamounts (those peaking < 950m depth) showed the greatest impact and deep-sea scleractinian coral reefs were reduced to rubble, comparatively to deep seamounts (where the summit depth is >1,500m) which generally exceeded the depth range of living deep-sea scleractinian corals and were also beyond the typical depth a bottom trawler operates showed the least signs of impact²⁶².

Section 4: Seamount protection

Almost half of Aotearoa's known seamounts (> 1,000m elevation) and associated biodiversity are at risk from bottom trawling (48% are open to bottom trawling while 52% are protected within Benthic Protected Areas (BPAs)²⁶³) and most other similar underwater features²⁶⁴ are vulnerable to damage from bottom trawling as 72% of all such features (including seamounts >100m), are open to bottom trawling while only 28% are protected²⁶⁵.

Is existing protection adequate?

The highest level of marine protection in Aotearoa are type 1 Marine Protected Areas (MPAs), known as no-take marine reserves. Aotearoa has 44 type 1 MPAs, which equates to less than 0.5%²⁶⁶ of Aotearoa's entire EEZ²⁶⁷ (Figure 24). No type 1 MPAs can be established beyond the territorial sea (12 nm) into the EEZ where the majority of seamounts occur.

²⁵⁴ Duncan, 1873 described by Williams et al.2020.

²⁵⁵ Clark & Rowden, 2009

²⁵⁶ Williams et al. 2020

²⁵⁷ Williams et al. (2020) reviewed commercial trawl logbook data and found there was no effort in waters deeper than about 1,500 m and on seamounts with peaks deeper than about 1,200 m

²⁵⁸ Collette, 1889 described by Williams et al. 2020

²⁵⁹ Edmonds et al. 1991

²⁶⁰ Williams et al. 2020

²⁶¹ Williams et al. 2020

²⁶² Williams et al. 2020

²⁶³ Ministry for Primary Industries, 2021

²⁶⁴ Underwater topographic features include seamounts in many forms including hills and knolls.

²⁶⁵ Ministry for Primary Industries, 2021

²⁶⁶ 17,430km²

²⁶⁷ Department of Conservation, 2021

Aotearoa has closed approximately 1.1 million km² (~30%) of the EEZ to bottom trawling, known as BPAs and seamount closed areas²⁶⁸, Figure 25 shows the BPAs. Fisheries New Zealand (2020) state “*Much of the seabed within BPAs is below trawlable depth (maximum trawlable depth is about 1600m) and all are outside the Territorial Sea. In combination, the seamount closures and the BPAs include: 28% of underwater topographic features (a term that includes underwater hills, knolls, and seamounts); 52% of seamounts over 1000m high; and 88% of known active hydrothermal vents*”²⁶⁹.

BPAs protect seamounts and similar features from the impacts of bottom trawling but other fishing methods that do not contact the seafloor are permitted, such as long-lining and midwater trawl²⁷⁰. BPAs do not meet the protection standard for the New Zealand government’s MPA Policy²⁷¹, and they would also fail to meet the lowest category in the internationally recognised IUCN marine protected area categories as they do not meet several of the criteria such as preventing mining or industrial fishing^{272, 273}.

Biodiversity knowledge from BPAs is limited, most BPAs have been sampled less than 100 times and are “*typically poorly described*”²⁷⁴. New species of invertebrates have recently been described from the Bounty Heritage BPA²⁷⁵, which highlights that further work is needed to identify historic specimens, as there are likely to be new undescribed species. Clarke et al. (2019) recently compared certain species compositions inside and outside BPAs for the Chatham Rise between depths of 100m - 1000m and found that the BPAs had 27% of the known benthic invertebrate species from the Rise²⁷⁶. The same study stated that the BPAs effectiveness could be improved for biodiversity protection and also referred to similar findings from Leathwick et al (2008), Bors et al (2012) and Zeng et al (2017)²⁷⁷.



©NIWA. Di Tracey and a large specimen of bubblegum coral (Paragorgia Arborea), New Zealand

²⁶⁸ In 2001 the Government prohibited trawling on 17 seamounts, these are known as the seamount closed areas

²⁶⁹ Fisheries New Zealand, 2020

²⁷⁰ requires two observers are on board and an approved net monitoring system is used

²⁷¹ Lundquist et al. 2015

²⁷² Day et al. 2019

²⁷³ In 2019 the then Minister of Conservation confirmed that the BPAs do not constitute marine protected areas nor count towards Aotearoa New Zealand’s goals or commitments for marine protection. Vance, 2019

²⁷⁴ Clark et al, 2019b

²⁷⁵ Clark et al, 2014 found a new genus and 13 new species records of Porifera and 21 new species of Bryozoa described in Clark et al, 2019b

²⁷⁶ Clark et al, 2019b

²⁷⁷ Clark et al, 2019b

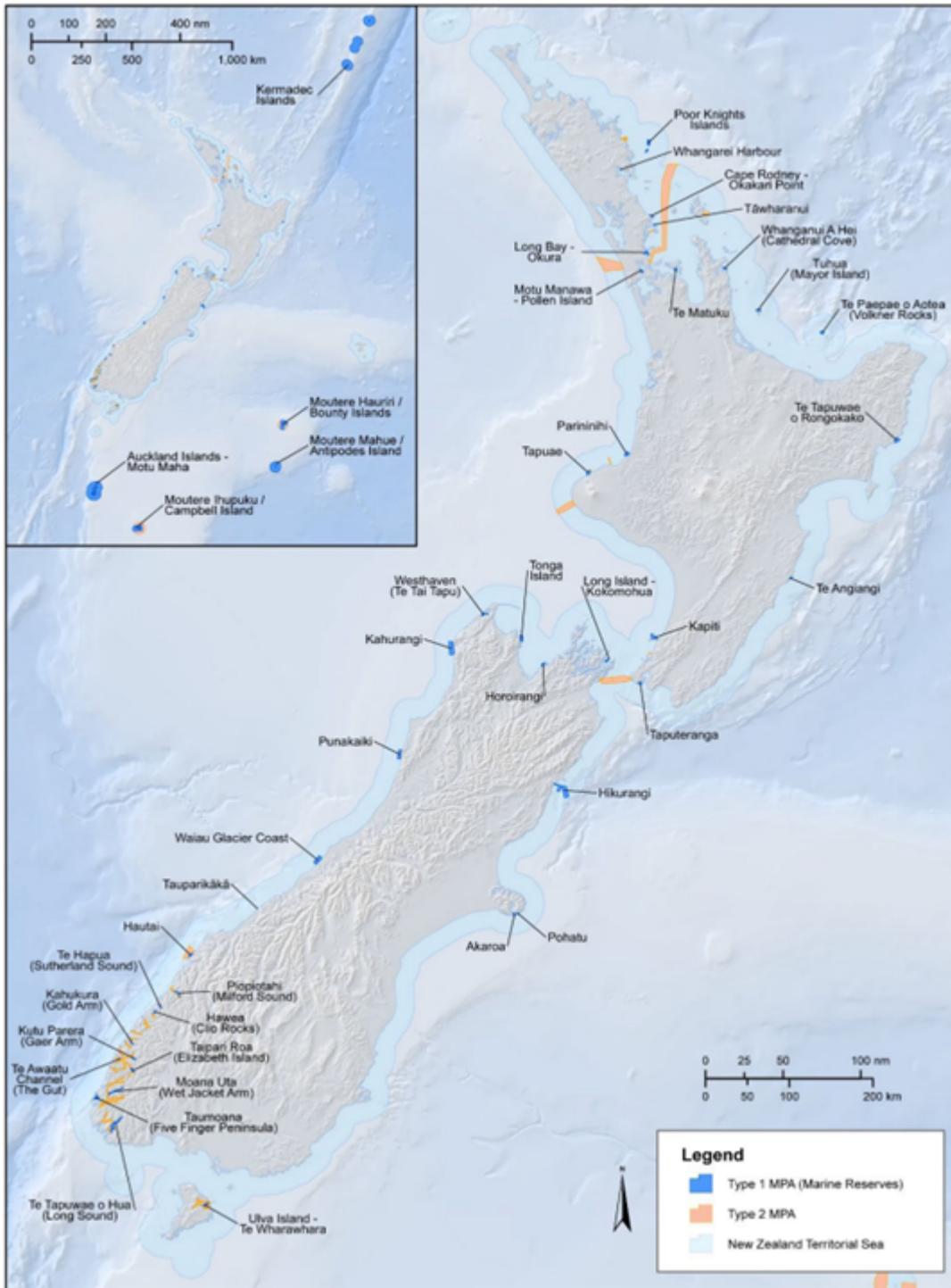


Figure 24: Marine Protected Areas in New Zealand - Type 1 (Marine reserves) and Type 2 MPAs, at December 2020
 Source: DOC Information Services²⁷⁸

²⁷⁸ Department of Conservation, 2021

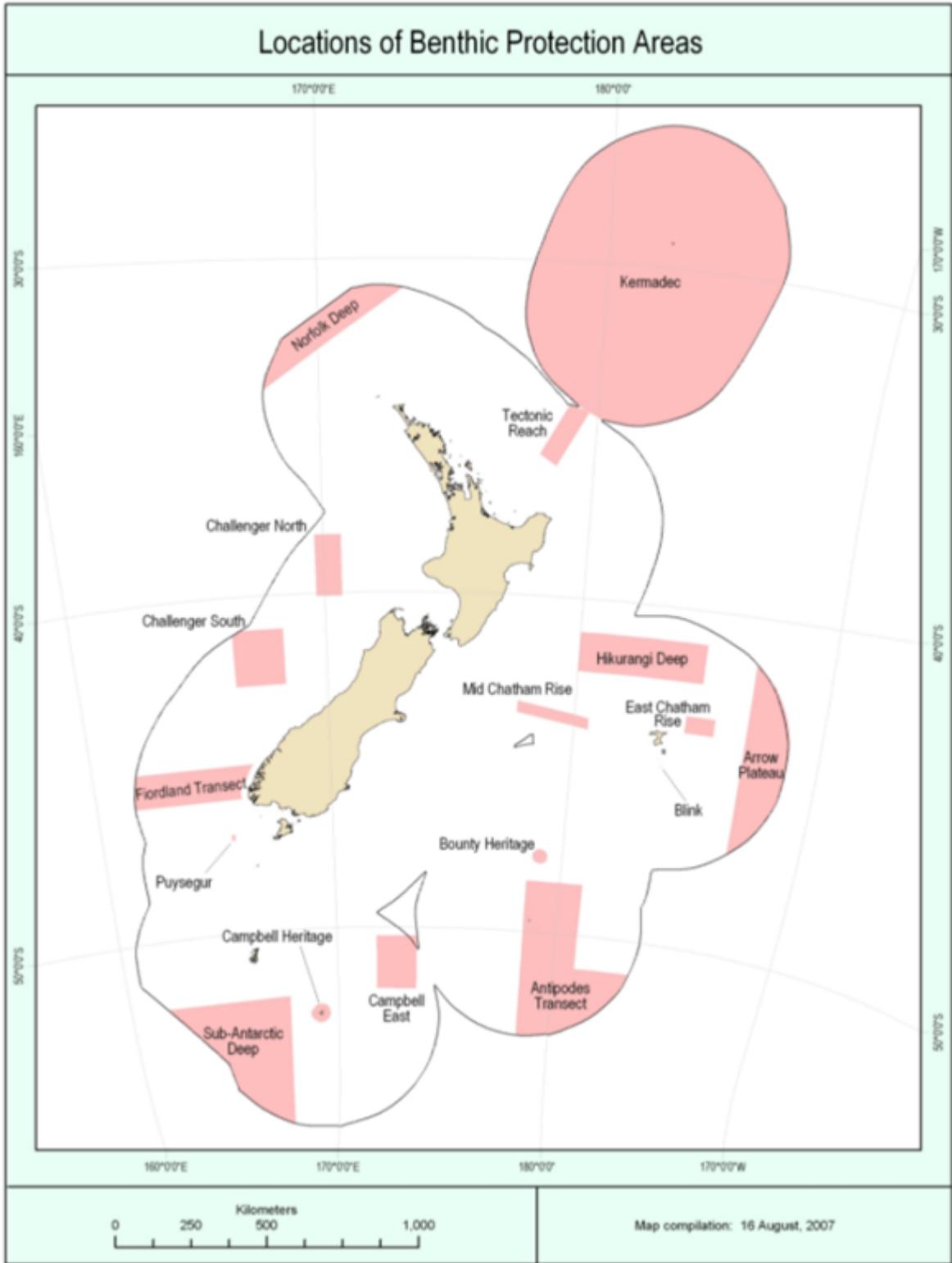


Figure 25: Map of Benthic Protected Areas within Aotearoa. Source: Fisheries New Zealand, 2009

Based on the depth distribution plots (Figures 5, 9, 11, 15 and 14) from Tracey & Hjørvarsdóttir (2019), and what is known about the depths different protected and non-protected corals live, it can be generalised that most Aotearoa deep-sea coral taxa can be found between 200m - 1,000m²⁷⁹, we refer to this as the ‘core depth distribution’ in this report.

Aotearoa deep-sea coral core depth distribution overlaps with ‘fishable depths’. Deep water fisheries in Aotearoa generally operate between 200m - 1,200m (maximum trawable depth is 1,600m)²⁸⁰, for example the main bottom trawl fishing depths for orange roughy is between 800m – 1,200m²⁸¹.

Nine protected coral groups bycatch distributions were reviewed based on observers reports between 2007 - 2010. Protected coral was reported from a wide range of depths, the most common was from between 800m - 1200m, and came from target fisheries for orange roughy, black oreo, smooth oreo and black cardinalfish²⁸².

These deep-sea coral core depth distributions highlight that it’s not the elevation of the seamount above the seafloor (>100m vs > 1,000m) that is a biological driving factor for these protected corals and other associated invertebrates, instead it’s the depth from the surface. Specifically, if seamounts occur within the core depth distribution range for deep-sea corals, then the biodiversity associated with these seamounts and similar features are likely to increase. Given much of the “*seabed within BPAs is below trawlable depth (maximum trawlable depth is about 1600m)*”²⁸³ it’s likely not all these deep seamounts would support these protected deep-sea corals, as they are simply too deep.

FNZ estimates approximately 70% of the EEZ is deeper than 1,250m²⁸⁴. The ‘fishable’ area, defined as waters shallower than 1,600m within the territorial sea and EEZ excluding all BPAs, seamount closures and marine reserves is 34% of Aotearoa’s waters²⁸⁵. This means 66% of the EEZ is deeper than 1,600m (excluding protected areas). Given this, and the depth restrictions of some corals, this means many species will also only be found within this ‘fishable’ area, the 34% or less of Aotearoa’s waters.

The BPAs have been widely criticised for lacking in scientific basis related to their biodiversity value or species representativeness. Genetic research in Aotearoa, looking at two stony and two black coral species, which are both VME indicator taxa, found that the current unrepresentative deep-sea spatial seamount closures (BPAs) need to be improved²⁸⁶. Not all seamounts are currently protected in Aotearoa, which means not all genetic diversity of the invertebrates like corals are protected. Some deep-sea corals have limited connectivity²⁸⁷ making them more vulnerable to benthic disturbances such as bottom trawling. Holland et al. (2020) found that the genetic diversity of corals on the Chatham Rise region (central region) is not well protected in BPAs and is one of the most heavily trawled areas.

²⁷⁹ Tracey & Hjørvarsdóttir, 2019

²⁸⁰ Fisheries New Zealand, 2020

²⁸¹ Deep Water Group Limited, 2021

²⁸² Baird et al, 2013

²⁸³ Fisheries New Zealand, 2020

²⁸⁴ Fisheries New Zealand, 2019

²⁸⁵ Black & Tilney, 2017

²⁸⁶ Holland et al. (2020)

²⁸⁷ Holland et al. 2020

Existing BPA protection is not preventing the potential loss of deep-sea biodiversity that hasn't even been discovered. A recent government workshop stated that new coral species are still being identified because global taxonomic experts are still reviewing historic samples and commercial fishing vessels are still going to new areas which means encountering new species that scientists didn't even know existed²⁸⁸. Some of these new species come from seamounts.

WWF-NZ (2021) found that in the ten years from 2009 to 2019 3,280 km² of the seabed was bottom trawled for the first time. This is an area more than three times the size of Auckland (1,086 km²)²⁸⁹. Over the past decade, most of the newly trawled areas (around 75%) were from inshore fisheries, however in 2019 85.7 km² of deep-sea was newly trawled and 62.3km² of inshore waters.

FNZ concluded that the overall trawl footprint when comparing 2007/08 to 2017/18 against 1989/90 to 2006/07 is decreasing “*after a peak period from 1998 - 2003 (Baird & Wood 2018), and has stabilised, at a lower level in recent years*”²⁹⁰. This statement does not reflect that while the annual footprint may be smaller, it has increasingly consisted of new areas that have never been trawled before and therefore the cumulative footprint of the seabed within ‘fishable’ depths that have been trawled one or more times within Aotearoa is actually still expanding.

There is limited data available on deep-sea coral biology, productivity and even endemism. What is known, is that Aotearoa is home to a diverse range of deep-sea corals that are widely distributed, some are highly endemic and rare, and in general deep-sea corals tend to occupy a broad depth range from shallow waters to well over 2,000m deep, but many taxa usually have a core depth range of between 200 – 1,000m depth²⁹¹ and are associated with seamounts within ‘fishable depths’. The VME locations (Figure 4) compared to the BPAs, highlights the vulnerability of those VMEs outside of existing protected areas.

The Environment Select Committee (ESC) is currently reviewing a request presented by the Deep Sea Conservation Coalition on behalf of Aotearoa eNGOs and fishing groups²⁹² to ban bottom trawling on all seamounts within Aotearoa's EEZ and a request for the Government to stop issuing high seas permits to New Zealand companies bottom trawling beyond Aotearoa's EEZ in areas like SPRFMO. At the date of this publication the ESC had not made their recommendation to the government and were still reviewing all the evidence received²⁹³.

Understanding the biological value of the 100m seamount elevation classification versus the 1,000m is important given the submission to the Environment Select Committee (ESC) by the Deepwater Group (DWG), an alliance of deepwater quota owners in response to the DSCC. The DWG submission states that the definition of a seamount is “*an underwater feature with an elevation of more than 1,000 meters*” and in terms of New Zealand, refers to the use of this elevation by New

²⁸⁸ Tracey & Hjørvarsdóttir, 2019 - Appendix 4:

²⁸⁹ WWF-NZ, 2021

²⁹⁰ Baird & Mules (2020a, in review) described in Fisheries New Zealand, 2020

²⁹¹ Tracey & Hjørvarsdóttir, 2019

²⁹² Coalition includes: Deep Sea Conservation Coalition, Environment and Conservation Organisations (ECO), Greenpeace Aotearoa, Forest & Bird, LegaSea, New Zealand Sports Fishing Council, Our Seas Our Future and WWF-New Zealand

²⁹³ All evidence of the Petition of Karli Thomas, on behalf of the Deep Sea Conservation Coalition and its member groups: Save deep sea corals - ban bottom trawling on seamounts can be found here: [Submissions and Advice - New Zealand Parliament \(www.parliament.nz\)](http://www.parliament.nz)

Zealand Geographic Board, New Zealand’s naming authority. The DWG did not refer or quote DOC and FNZ’s joint evidence to the ESC which was discussed in Section 1.

DOC and FNZ evidence highlights that the “IUCN *Atlas of Mediterranean Seamounts defines seamounts and seamount-like structures as all elevations that rise more than 100 meters from the surrounding seafloor*” and “*that the physical characteristic of small seamounts-like features less than 1,000 meters above the seafloor affect biology in very similar ways to large seamounts greater than 1,000 meters above the seafloor*”²⁹⁴. In their joint evidence, FNZ and DOC use the elevation of 100m above the seafloor to describe seamounts and seamount-like features²⁹⁵.

Can protection work?

Research following fisheries closures on Aotearoa and Australian seamounts after 5 and 10 years respectively found no change, suggesting that recovery in benthic community structure to pre-disturbed state on seamounts can take decades to centuries²⁹⁶ because of the slow growth rates and high longevity of a number of invertebrates such as corals²⁹⁷.

The most recent time-series study from Aotearoa looking at the impacts of bottom trawling and recovery of benthic communities surveyed seamounts from the Graveyard Knolls seamount group (Figure 26). The Graveyard Knolls seamounts comprises of 28 small volcanic cone features typically between 100 - 400 m high, rising from depths of 1,050 - 1,200m to summit depths of 750 - 1,000m²⁹⁸. A trawl fishery for spawning orange roughy began on the Graveyard Knolls in the early 1990s, with most effort on two features, “Morgue” and “Graveyard”²⁹⁹. In 2001 “Morgue” was closed. The study surveyed six of these seamounts representing a range of trawl histories, one with high and persistent trawling levels, two with intermediate trawling, two with low trawling or ‘untrawled’ during the survey and one closed³⁰⁰. Clark et al. (2019) found the communities of organisms that lived on seamounts prior to fishing activities showed no sign of recovery, even though the seamounts had not been fished for at least 15 years³⁰¹.

These studies confirm the vulnerability of slow growing, high longevity invertebrates, like corals and the low resilience benthic communities on seamounts³⁰² and similar underwater features have to the effects of benthic disturbance like bottom trawling.

An international study of shallower seamounts than these, in the northwest Hawaiian Ridge and southern Emperor Seamount Chain, found some indications of coral recovery 30 years after they were protected from trawling in the 1970s and 1980s³⁰³.

These results are significant because they confirm that regeneration of deep-sea coral communities on seamounts may be possible but require longer 30-40 year time scales³⁰⁴.

²⁹⁴ Department of Conservation & Fisheries New Zealand, 2021

²⁹⁵ Department of Conservation & Fisheries New Zealand, 2021

²⁹⁶ Williams et al. 2010

²⁹⁷ Clark et al. 2019

²⁹⁸ Clark et al. 2019

²⁹⁹ Clark et al. 2019

³⁰⁰ Clark et al. 2019

³⁰¹ Clark et al. 2019

³⁰² Williams et al. 2020

³⁰³ Baco et al. 2019

³⁰⁴ Baco et al. 2019

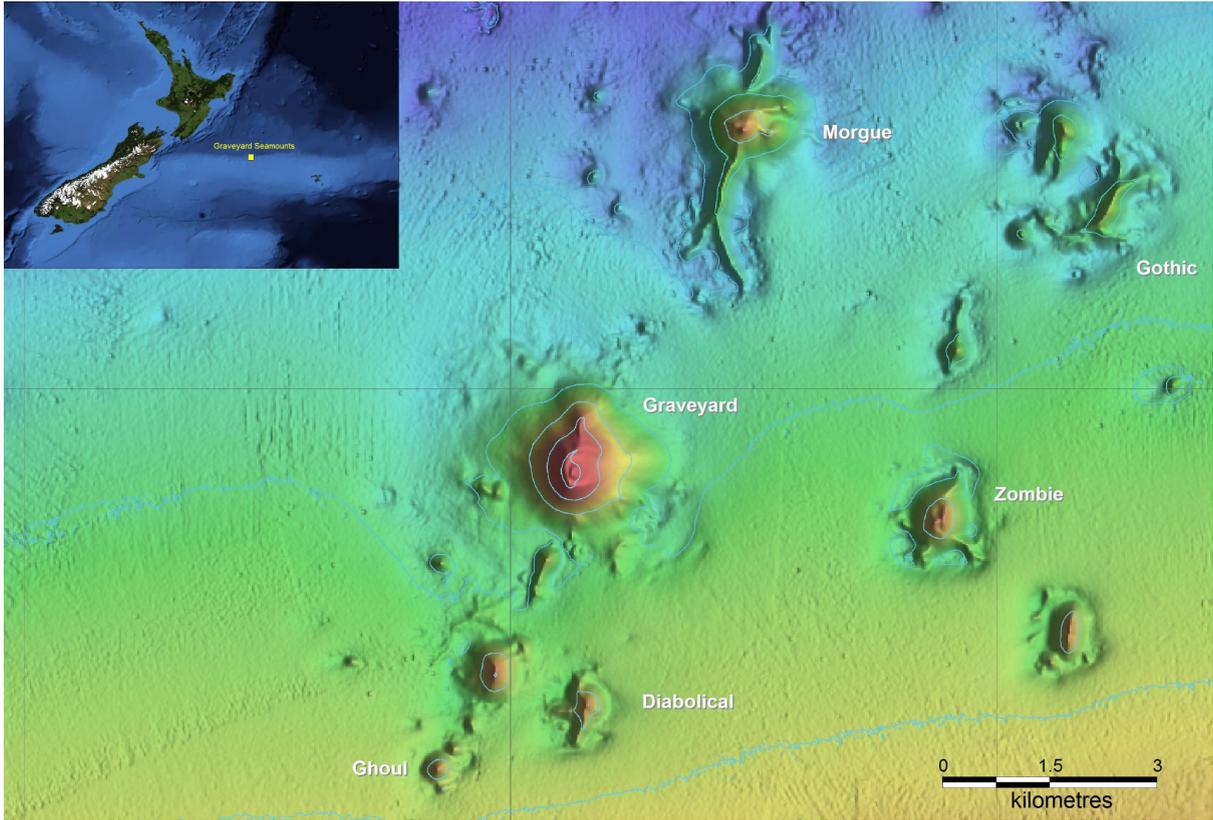
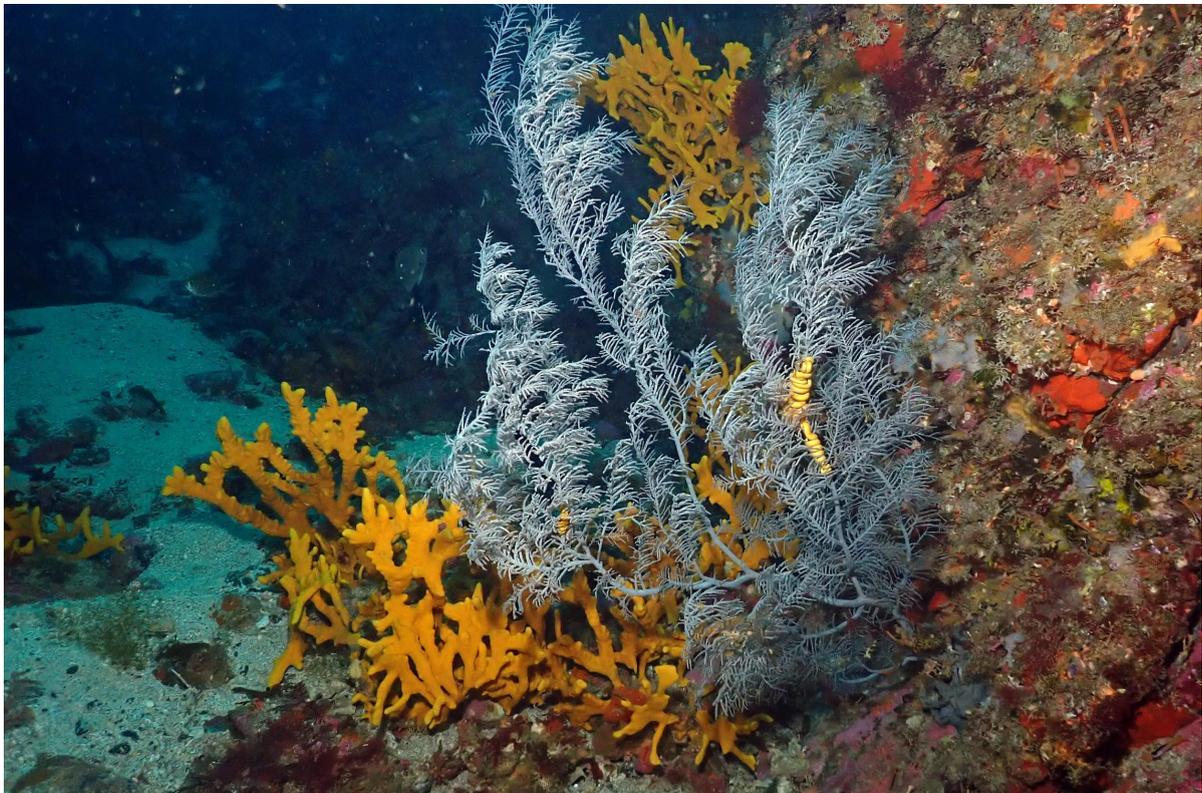


Figure 26: Shows the Graveyard Knolls, a group of small seamounts and features located east of Aotearoa. Source: Clark et al. 2019



© Andy Stewart. Black coral, Hen & Chickens Islands, Northland New Zealand, 2021

Acronyms and Abbreviations:

ANZBS	Te Mana o te Taiao Aotearoa New Zealand Biodiversity Strategy 2020
BPAs	Benthic Protected Areas
DOC	Department of Conservation
DWG	Deepwater Group Limited
EEZ	Exclusive Economic Zone (offshore to 200 nautical miles)
ESC	Environment Select Committee
FAO	Food and Agriculture Organisation of the United Nations
FMA	fisheries management area
FNZ	Fisheries New Zealand
RFMO	Regional Fisheries Management Organisations
SAI	significant adverse impact
SPFRMO	South Pacific Regional Fisheries Management Organisations
TS	Territorial Sea (0-12 nautical miles)
UNCLOS	United Nations Convention on the Law of the Sea
UNGA	United Nations General Assembly
VME	Vulnerable Marine Ecosystems

Key Definitions:

Seamount: Seamounts are underwater mountains that rise more than 100m above the surrounding seafloor, where the seafloor is more than 200 metres below the surface³⁰⁵.

The FAO definition is “*A large isolated elevation characteristically of conical form. Seamounts are under-sea mountains whose summits lie beneath the ocean surface. They are usually volcanic in origin and are generally defined as having an elevation of greater than 1 000 metres from the seabed*”³⁰⁶.

Knolls Relatively small (500 to 1000 m tall) isolated elevation of a rounded shape

Guyots A seamount with a truncated, flat top surface³⁰⁷.

VME Vulnerable marine ecosystems (VMEs) are those ecosystems highly vulnerable to one or more kinds of human activity such as fishing. VMEs are identified by the vulnerability of their species, communities, and or habitats to disturbance. A VME is described in the Deep-sea Fisheries Guidelines by its characteristics and by its vulnerability³⁰⁸.

SAI “negative effects on a VME resulting from damage caused during the operation of bottom-contact fishing gear. The scale and significance of the impact determines whether the impact can be considered an SAI. This occurs when the ecosystem function is impaired and the long-term natural productivity degraded on more than a temporary basis. Ecosystem recovery following impacts is case-by-case dependent, but is considered more than temporary if recovery takes more than 5 – 20 years.”³⁰⁹

³⁰⁵ Rogers, 2018

³⁰⁶ Cochrane, 2002

³⁰⁷ Pitcher et al., 2007

³⁰⁸ FAO, 2009; [Definitions | Vulnerable Marine Ecosystems | Food and Agriculture Organization of the United Nations \(fao.org\)](#)

³⁰⁹ FAO, 2009

References:

- Allain, V., Kerandel, J.A. & M. Clark.. (2008). Potential seamount location in the South Pacific RFMO area: prerequisite for fisheries management and conservation in the high seas. Paper SPRFMO-V-SWG-05 https://www.researchgate.net/publication/237754645_Potential_seamount_location_in_the_South_Pacific_RFMO_area_prerequisite_for_fisheries_management_and_conservation_in_the_high_seas
- Anderson, O., Clark, M.R. (2003) Analysis of bycatch in the fishery for orange roughy, *Hoplostethus atlanticus*, on the South Tasman Rise. *Marine and Freshwater Research*, 54: 643–652
- Annasawmy, P., Ternon, J-F., Cotel, P., Cherel, Y., Romanov, E.V., Roudaut, G., Lebourges-Dhaussy, A., Ménard, F. and F. Marsac. (2020) Micronekton distributions and assemblages at two shallow seamounts of the south-western Indian Ocean: Insights from acoustics and mesopelagic trawl data. *Progress in Oceanography* 178, 102161. <https://doi.org/10.1016/j.poccean.2019.10216>
- Baco, A. R., Roark, E. B. and N. B. Morgan. (2019) Amid fields of rubble, scars, and lost gear, signs of recovery observed on seamounts on 30- to 40-year time scales. *Science Advances*. 7 Aug 2019: Vol. 5, no. 8. <https://advances.sciencemag.org/content/5/8/eaaw4513>
- Baird, S., Tracey, D., Mormede, S., & M. Clark. (2013). The distribution of protected corals in New Zealand waters. NIWA report prepared for the Department of Conservation. WLG2012-43. NIWA, Wellington [FINAL FEB2013-DOC12303-WLG2012-43](https://www.doc.govt.nz/our-work/conservation-services-programme/csp-reports/2012/43/)
- Bijma, J., Portner, H., Yesson, C., & A. Rogers. (2013). Climate change and the oceans – What does the future hold? *Marine Pollution Bulletin* 74(2) 495 – 505 <https://doi.org/10.1016/j.marpolbul.2013.07.022>
- Bilewicz, J. and Tracey, D. (2020a) Coral biodiversity in deepwater fisheries bycatch Final Report. NIWA report INT2019-05 prepared for the Department of Conservation, July 2020. <https://www.doc.govt.nz/our-work/conservation-services-programme/csp-reports/201920/coral-biodiversity-in-deepwater-fisheries-bycatch/>
- Bilewicz, J. and Tracey, D. (2020b) Protected coral connectivity in New Zealand. Final Report for project POP2018-06 prepared by NIWA for the Department of Conservation. DOC19306- POP201806. <https://www.doc.govt.nz/our-work/conservation-services-programme/csp-reports/201920/protected-coral-connectivity-in-new-zealand>
- Boschen, R.E. Rowden, A.A., Clark, M.R., Barton, S.J., Pallentin, A., & J.P.A. Gardner. (2015). Megabenthic assemblage structure on three New Zealand seamounts: implications for seafloor massive sulfide mining. *Marine Ecology Progress Series* 523: 1-14. [Marine Ecology Progress Series 523:1 \(int-res.com\)](https://www.researchgate.net/publication/275111111_Marine_Ecology_Progress_Series_523:1)
- Bowden, D., & D. Luduc. (2017). Ocean survey 20/20, Chatham Rise Benthos: effects of seabed trawling on benthic communities. *New Zealand Aquatic Environment and Biodiversity Report No.183*. Ministry for Primary Industries. [*AEBR 183 - Ocean Survey 20/20, Chatham Rise Benthos: effects of seabed trawling on benthic communities \(mpi.govt.nz\)](https://www.mpi.govt.nz/our-work/conservation-services-programme/csp-reports/2017/183/)
- Bull, B., Doonan, I., Tracey, D., & A. Hart. (2001). Diel variation in spawning orange roughy (*Hoplostethus atlanticus*, Trachichthyidae) abundance over a seamount feature on the north-west Chatham Rise, New Zealand *Journal of Marine and Freshwater Research*, 35:3, 435-444 [Diel variation in spawning orange roughy \(Hoplostethus atlanticus, Trachichthyidae\) abundance over a seamount feature on the north-west Chatham Rise: New Zealand Journal of Marine and Freshwater Research: Vol 35, No 3 \(tandfonline.com\)](https://www.tandfonline.com/doi/abs/10.1080/10236198.2001.10799100)
- Clark, M. (2008). Seamount fisheries: understanding the impacts of trawling. *NIWA Water & Atmosphere* 16(2) [seamount fisheries - understand the impacts of trawling.pdf \(niwa.co.nz\)](https://www.niwa.co.nz/publications/2008/08/seamount-fisheries-understanding-the-impacts-of-trawling.pdf)
- Clark, M. (2021). Update on Seamount Database for Deepwater Group Ltd. NIWA client report number 2021034WN. Wellington. [NIWA Client report \(deepwatergroup.org\)](https://www.niwa.co.nz/publications/2021/03/4WN/)
- Clark, M., Bowden, D., Baird, S., and R. Stewart. (2010) Effects of fishing on the benthic biodiversity of seamounts of the “Graveyard” complex, northern Chatham Rise. *New Zealand Aquatic Environment and Biodiversity Report*

No. 46. 40 pp. Ministry for Primary Industries, New Zealand
https://www.researchgate.net/publication/290487233_Effects_of_fishing_on_the_benthic_biodiversity_of_seamounts_of_the_%27Graveyard%27_complex_northern_Chatham_Rise

Clark, M. R., Anderson, O. F., Bowden, D. A., Chin, C., George, S. G., Glasgow, D. A. et al. (2015). Vulnerable marine ecosystems of the Louisville Seamount chain: voyage report of a survey to evaluate the efficacy of preliminary habitat suitability models. N.Z. Aquat. Environ. Biodivers. Rep. 149, 1–88. (PDF) [Vulnerable Marine Ecosystems of the Louisville Seamount Chain: voyage report of a survey to evaluate the efficacy of preliminary habitat suitability models \(researchgate.net\)](#)

Clark, M., Bowden, D., Rowden, A.A., & R. Stewart. (2019). Little evidence of benthic community resilience to bottom trawling on seamounts after 15 years. *Frontiers in Marine Science* 6:63
<https://doi.org/10.3389/fmars.2019.00063>

Clarke, M., Mills, S., Leduc, D., Anderson, O.F., & A.A. Rowden. (2019). Biodiversity of Benthic Protected Areas and Seamount Closed Areas: a description of available benthic invertebrate data, and a preliminary evaluation of the effectiveness of BPAs for biodiversity protection. New Zealand Aquatic Environment and Biodiversity Report No 277. Fisheries New Zealand [Biodiversity of Benthic Protection Areas and Seamount Closure Areas: a description of available benthic invertebrate data, and a preliminary evaluation of the effectiveness of BPAs for biodiversity protection. \(mpi.govt.nz\)](#)

Cochrane, K.L. ed., (2002). A fishery manager's guidebook. Management measures and their application. (No. 424). Food and Agriculture Organisation of the United Nations [A fishery manager's guidebook - Second edition \(fao.org\)](#)

Consalvey, M., MacKay, K., and D. Tracey (2006) Information review for protected deep-sea coral species in the New Zealand region. NIWA Project: DOC06307. Department of Conservation, New Zealand Information review for protected deep-sea coral species in the New Zealand region (doc.govt.nz) [Information review for protected deep-sea coral species in the new zealand region \(doc.govt.nz\)](#)

Cryer, M., & S. Geange. (2018). Review of benthic sampling and bycatch data, including VME taxa, in SPRFMO bottom fisheries .Paper SC6-WD14 for 6th meeting of the SPRFMO Scientific Committee, Puerto Varas, Chile, 9-14 September, 2018 [SC6-DW14-Benthic-bycatch-summary.pdf \(sprfmo.int\)](#)

Cryer, M. Geange, S. and Nicol, S. (2018). Methods for deriving thresholds for VME encounter protocols for SPRFMO bottom fisheries. SC6-DW09.
<https://www.sprfmo.int/assets/2018-SC6/Meeting-Documents/SC6-DW09-Methods-deriving-VME-thresholds.pdf>

Day, J., Dudley, N., Hockings, M., Holmes, G., Laffoley, D., Stolton, S., Wells, S. and Wenzel, L. (eds.) (2019). Guidelines for applying the IUCN protected area management categories to marine protected areas. Second edition. Gland, Switzerland: IUCN. <https://portals.iucn.org/library/node/48887>

Deepwater Group Limited. (2021). Submission to the Environment Select Committee Deepwater Group Report. Received 25th September
[DWG-Submission-on-Seamounts-and-Bottom-Trawling-to-Environment-Select-Committee-300821-WEB.pdf \(deepwatergroup.org\)](#)

Demarcq, H., Noyon, M. and M.J. Roberts. (2020) Satellite observations of phytoplankton enrichments around seamounts in the South West Indian Ocean with a special focus on the Walters Shoal. *Deep-Sea Research Part II: Topical Studies in Oceanography* 176, 104800.
<https://doi.org/10.1016/j.dsr2.2020.104800>

Department of Conservation. (2021). Marine Protected Areas: Tier 1 Statistics, 2020. Retrieved from [Clear statement on degree of compliance with agreed definitions, methods and practices \(where identified\) including any reasons for deviation \(doc.govt.nz\)](#)

Department of Conservation and Fisheries New Zealand. (2021). Joint evidence for the Environment Select Committee on the Petition of Karli Thomas, on behalf of the Deep Sea Conservation Coalition and its member groups: Save deep sea corals – ban bottom trawling on seamounts. DOCCM 6704230 and FNZ SCR21-0018. [Joint evidence from DOC & FNZ - petition 2020/01 in the name of Karli Thomas - DOCCM-6704230 - FNZ-SCR21-0018 \(www.parliament.nz\)](https://www.parliament.nz/joint-evidence-from-doc-&fnz-petition-2020/01-in-the-name-of-karli-thomas-docco-6704230-fnz-scr21-0018)

Dijkstra, J.A., Mello, K., Sowers, D., Malik, M., Watling, L., & L. Mayer. (2021). Fine-scale mapping of deep-sea habitat-forming species densities reveals taxonomic specific environmental drivers. *Global Ecology and Biogeography*, 2021; DOI: [10.1111/geb.13285](https://doi.org/10.1111/geb.13285)

DSCC. (2021). Protecting Global Seamounts. Deep Sea Conservation Coalition [Protecting-Global-Ocean-Seamounts-final-web.pdf \(savethehighseas.org\)](https://www.savethehighseas.org/protecting-global-ocean-seamounts-final-web.pdf)

Edmonds, J.S., Caputi, N., & M. Morita. (1991). Stock discrimination by trace-element analysis of otoliths of orange roughy (*Hoplostethus atlanticus*), a deep-water marine teleost. *Australian Journal of Marine and Freshwater Research* 42: 383 - 389. [Sci-Hub | Stock discrimination by trace-element analysis of otoliths of orange roughy \(Hoplostethus atlanticus\), a deep-water marine teleost. Marine and Freshwater Research, 42\(4\), 383 | 10.1071/MF9910383](https://doi.org/10.1071/MF9910383)

FAO. 2009. *International Guidelines for the Management of Deep-sea Fisheries in the High Seas*. Rome. 73pp [International guidelines for the management of deep-sea fisheries in the high seas/Directives internationales sur la gestion de la pêche profonde en haute mer/Directrices internacionales para la ordenación de las pesquerías de aguas profundas en alta mar \(fao.org\)](https://www.fao.org/publications/default.aspx?info/9789251042396)

Fisheries New Zealand. (2009). Benthic Protected Areas Map. Retrieved 6th September 2021 from [Benthic Protection Area map \(fish.govt.nz\)](https://www.fish.govt.nz/benthic-protection-area-map)

Fisheries New Zealand. (2019). National Fisheries Plan for Deepwater and Middle-depth Fisheries 2019. Fisheries New Zealand Technical Paper No: 2019/03 [Fisheries New Zealand Discussion/Technical/Information Paper template \(mpi.govt.nz\)](https://www.mpi.govt.nz/technical-information-paper-template)

Fisheries New Zealand (2019). Sustainable New Zealand seafood - Protecting New Zealand's seabed from the impacts of bottom trawling. Retrieved from [Protecting New Zealand's Seabed from the impact of bottom trawling \(mpi.govt.nz\)](https://www.mpi.govt.nz/protecting-new-zealand-s-seabed-from-the-impact-of-bottom-trawling)

Fisheries New Zealand (2020). Aquatic Environment and Biodiversity Annual Review 2019– 20. Compiled by the Fisheries Science Team, Ministry for Primary Industries, Wellington New Zealand. <https://www.mpi.govt.nz/dmsdocument/40980-Aquatic-Environment-and-Biodiversity-Annual-Review-201920>

Fisheries New Zealand, (2021). New Zealand Annual Report on Fishing, Research Activities, and Observer Implementation in the SPRFMO Convention Area during 2020. Paper SC9-Doc15 for 9th meeting of the SPRFMO Scientific Committee, held virtually 27th September – 2nd October, 2021.

Fisheries New Zealand. (2021). New Zealand Annual Report on Fishing, Research Activities and Observer Implementation in the SPRFMO Convention Area during 2020. Paper SC9-DOC 15 for 9th meeting of the SPRFMO Scientific Committee, held virtually 27th September – 2nd October, 2021. [*SC9-Doc15 \(sprfmo.int\)](https://www.sprfmo.int/sc9-doc15)

Fisheries New Zealand, (2020). Aquatic Environment and Biodiversity Annual Review 2019– 20. Compiled by the Aquatic Environment Team, Fisheries Science and Information, Fisheries New Zealand, Wellington New Zealand. 765 p. <https://www.mpi.govt.nz/dmsdocument/40980-Aquatic-Environment-and-Biodiversity-Annual-Review-201920>

Food and Agriculture Organization. (2009) *International Guidelines for the Management of Deepsea Fisheries in the High Seas*. Rome, Italy. 90 pp. ISBN: 9789250062587 www.fao.org/3/i0816t/I0816T.pdf

Fuller, S., Currie, D., Gianni, M., Goldsworthy, L., Rigby, S., Schleit, K., Simpfendorfer, C., Watling, L & B. Weeber. (2020). Halting and reversing biodiversity loss in the deep sea – A critique of compliance by high seas fishing nations

and RFMOs with global environmental commitments – October 2020. Deepsea Conservation Coalition <http://www.savethehighseas.org/resources/publications/preventing-biodiversity-loss-in-the-deep-sea-a-critique-of-compliance-by-high-seas-fishing-nations-and-rfmos-with-global-environmental-commitments/>

Garrigue, C./, Clapham, P.J., Geyer, Y., Kennedy, A.S., & A.N. Zerbini. (2015). Satellite tracking reveals novel migratory patterns and the importance of seamounts for endangered South Pacific humpback whales. *Royal Society Open Science*. 2:150489 <https://doi.org/10.1098/rsos.150489>

Geange, S., Rowden, A., Cryer, M., & T. Bock. (2020). Developing a multi-taxonomic level list of VME indicator taxa for the SPRFMO Convention Area- VME taxa for SPRFMO Area. Paper SC8-DW11 for 8th meeting of the SPRFMO Scientific Committee, held virtually 3-8 October, 2020 [*SC8-DW11 \(sprfmo.int\)](https://www.sprfmo.int/SC8-DW11)

Hitt, N. T., Sinclair, D. J., Fallon, S. J., Neil, H. L., Tracey, D. M., and A. Komugabe-Dixson. (2020) Growth and longevity of New Zealand black corals. *Deep Sea Research Part 1: Oceanographic Research Papers* 162 (103298) <https://doi.org/10.1016/j.dsr.2020.103298>

Holland, L.P., Rowden, A. A., Hamilton, J.Z., Clark, M.R., Chiswell, S.M., & J. P. A. Gardner. (2020). Genetic connectivity of deep-sea corals in the New Zealand region. *New Zealand Aquatic Environment and Biodiversity Report No. 245*. Fisheries New Zealand [Genetic connectivity of deep-sea corals in the New Zealand region. \(mpi.govt.nz\)](https://www.mpi.govt.nz/genetic-connectivity-of-deep-sea-corals-in-the-new-zealand-region/)

Iyer, S.D., Mehta, C.M., Das, P. & N.G. Kalangutkar. (2012). Seamounts—characteristics, formation, mineral deposits and biodiversity. *Geologica Acta*, 10(3), pp.295-308. [Seamounts – characteristics, formation, mineral deposits and biodiversity | . \(ub.edu\)](https://www.geologica-acta.com/seamounts-characteristics-formation-mineral-deposits-and-biodiversity/)

Kim, S.S. (2014) Seamount. In: *Encyclopedia of Planetary Landforms*. Springer, New York, NY. https://doi.org/10.1007/978-1-4614-9213-9_550-1

Kitchingman, Adrian & Lai, Sherman & Morato, Telmo & Pauly, Daniel. (2008). How Many Seamounts are There and Where are They Located?. [10.1002/9780470691953.ch2](https://doi.org/10.1002/9780470691953.ch2).

Le, J.T., & K.N., Sato. (2017). Ecosystem services of the deep ocean. *Ocean-climate report*. Retrieved from [ecosystem-services-deep-ocean ScientificNotes Oct2016 BD ppp-9.pdf \(ocean-climate.org\)](https://www.ocean-climate.org/ecosystem-services-deep-ocean-ScientificNotes-Oct2016-BD-ppp-9.pdf)

Koslow, A., Bax, N.J., Bulman, C.M., Kloser, R.J., Smith, A.D.M., and A. Williams. (1997). Managing the fishdown of the Australian orange roughy resource: Developing and sustaining world fisheries resources. The state of science and management., CSIRO, Collingwood (Australia), pp. 558-562

Koslow, J.A., Gowlett-Holmes, K., Lowry, J.K., O'Harr, T., Poore, G.C., & A. Williams. (2001). Seamount benthic macrofauna off southern Tasmania: community structure and impacts of trawling. *Marine Ecology Progress Series* 213: 111- 125. [Marine Ecology Progress Series 213:111 \(int-res.com\)](https://www.int-res.com/abstracts/meps/v213/p111-125)

Leathwick, J.R., Moilanen, A., Francis, M., Elith, J., Taylor, P., Julian, K., Hastie, T., and C. Duffy. (2008) Novel methods for the design and evaluation of marine protected areas in offshore waters. *Conservation Letters* 1: 91–102. <https://onlinelibrary.wiley.com/doi/full/10.1111/j.1755-263X.2008.00012.x>

Lundquist, C., Davies, K., and L. McCartain. (2015) Best practice guidelines for marine protected area network design and evaluation. NIWA Client Report No: HAM2015-051, June 2015. <https://www.researchgate.net/publication/319847702>

Marriott, P., Tracey, D. M., Bostock, H., Hitt, N., and S. J. Fallon. (2020) Ageing Deep-Sea Black Coral Bathypathes patula. *Frontiers in Marine Science* 7:479 [Frontiers | Ageing Deep-Sea Black Coral Bathypathes patula | Marine Science \(frontiersin.org\) https://doi.org/10.3389/fmars.2020.00479](https://doi.org/10.3389/fmars.2020.00479)

Milardi, M., Bock, T., & S. Geange. (2021). New Zealand 2020 VME encounter review. Paper SC9-DW09 for 9th meeting of the SPRFMO Scientific Committee, held virtually 27th September – 2nd October, 2021. [SC9-DW09 \(sprfmo.int\)](https://www.sprfmo.int/SC9-DW09)

Milardi, M., Bock, T., & S. Geange. (2021b). New Zealand 2020 VME encounter review. Paper SC9-DW09_rev1 for 9th meeting of the SPRFMO Scientific Committee, held virtually 27th September – 2nd October, 2021 <https://www.sprfmo.int/assets/2021-SC9/SC9-DW09-rev1-New-Zealand-2020-VME-encounter-review.pdf>
Retrieved 25th September, 2021

Ministry for the Environment & Stats NZ (2019). New Zealand's Environmental Reporting Series: Environment Aotearoa 2019. <https://environment.govt.nz/assets/Publications/Files/environment-aotearoa-2019.pdf>

Ministry for Primary Industries. (2021) Benthic protection areas. Web page, accessed 6th September 2021. www.mpi.govt.nz/fishing-aquaculture/sustainable-fisheries/protected-areas/benthicprotection-areas

Ministry of Fisheries. (2011). A guide to common deep sea invertebrates in New Zealand waters – Third edition. The New Zealand Aquatic Environment and Biodiversity Report No 86. ISSN 1176-9440 [AEBR_86.pdf](http://www.fish.govt.nz/AEBR_86.pdf) (fish.govt.nz)

Morato, T., Hoyle, S.D., Allain, V., S. J. Nicol. (2010). Seamounts are hotspots of pelagic biodiversity in the open ocean. *PNAS* 107:21 9707-9711 [pnas200910290.9707..9711](https://doi.org/10.1073/pnas.200910290)

New Zealand Government, 2021a. Closed Seamount Areas. Data accessed from [Closed Seamount Areas - Datasets - data.govt.nz - discover and use data](https://data.govt.nz/discover-and-use-data)

New Zealand Government, 2021b. Department of Conservation Marine Reserves. Data accessed from [Department of Conservation Marine Reserves - Datasets - data.govt.nz - discover and use data](https://data.govt.nz/discover-and-use-data)

New Zealand Government, 2021c. Benthic Protection Areas. Data accessed from [Benthic Protection Areas - Datasets - data.govt.nz - discover and use data](https://data.govt.nz/discover-and-use-data)

NIWA (2016). Vulnerable marine ecosystems in the South Pacific Ocean region. National Institute of Water and Atmospheric Research (NIWA), Wellington, New Zealand, 202873 records, Online http://nzbisipt.niwa.co.nz/resource.do?r=vme_inverts released on February 3, 2016. Retrieved from OBIS

NIWA (2018): NIWA Invertebrate Collection. v1.1. The National Institute of Water and Atmospheric Research (NIWA). Dataset/Occurrence. <https://nzobisipt.niwa.co.nz/resource?r=obisspecify&cv=1.1>

Department of Conservation. (2020). Te Mana o te Taiao Aotearoa New Zealand Biodiversity Strategy 2020. New Zealand Government. [Te Mana o te Taiao - Aotearoa New Zealand Biodiversity Strategy 2020 \(doc.govt.nz\)](https://www.doc.govt.nz/te-mana-o-te-taiao-aotearoa-new-zealand-biodiversity-strategy-2020/)

Owen, A., Guinotte, J., Rowden, A., Tracey, D., Mackay, K., & M. Clark. (2016). Habitat suitability models for predicting the occurrence of vulnerable marine ecosystems in the seas around New Zealand. *Deep-sea Research Part 1* [http://dx.doi.org/10.1016/j.dsr.2016.07.006](https://doi.org/10.1016/j.dsr.2016.07.006)

Owen, A., Stephenson, F., & E. Behrens. (2020). Updated habitat suitability modelling for protected corals in New Zealand waters, final report for the Department of Conservation, NIWA [NIWA Client report \(doc.govt.nz\)](https://www.doc.govt.nz/niwa-client-report/)

Pitcher, T. J., Morato, T., Hart, P.J.B., Clark, M.R., Haggan, N. & R.S. Santos (eds.). (2007). *Seamounts: Ecology, Fisheries & Conservation*. Fish and Aquatic Resources: Series 12. Blackwell Publishing, Oxford, United Kingdom [Frontmatter - Seamounts: Ecology, Fisheries & Conservation - Wiley Online Library](https://www.wiley.com/doi/10.1002/9781118152222)

Pitcher, R. Williams, A. and Georgeson, L. (2019) Progress with investigating uncertainty in the habitat suitability model predictions and VME indicator taxa thresholds underpinning CMM 03- 2019. Paper for SPRFMO SC7, SC7-DW21_rev1. <https://www.sprfmo.int/assets/2019-SC7/MeetingDocs/SC7-DW21-rev1-Uncertainty-in-model-predictions-and-VME-thresholds-for-CMM-03-2019.pdf>

Pitcher, R. Williams, A. and Georgeson, L. (2019) Progress with investigating uncertainty in the habitat suitability model predictions and VME indicator taxa thresholds underpinning CMM 03- 2019. Paper for SPRFMO SC7,

SC7-DW21_rev1.

<https://www.sprfmo.int/assets/2019-SC7/MeetingDocs/SC7-DW21-rev1-Uncertainty-in-model-predictions-and-VE-thresholds-for-CMM-03-2019.pdf>

Richer de Forges, B., Koslow, J.A., Poore, G.C.B. (2000). Diversity and endemism of the benthic seamount fauna in the southwest Pacific. *Nature* 405: 944-947 [Diversity and endemism of the benthic seamount fauna in the southwest Pacific | Nature](#)

Rieser, A., Watling, L., & J. Guinotte (2013) Trawl fisheries, catch shares and the protection of benthic marine ecosystems: Has ownership generated incentives for seafloor stewardship? *Marine Policy* 40. <http://dx.doi.org/10.1016/j.marpol.2012.12.028>

Rogers, A. (2018). The biology of seamounts: 25 years on. *Advances in Marine Biology* 79. 137 – 223. <https://doi.org/10.1016/bs.amb.2018.06.001>

Rogers, A. D. (2019) Chapter 23 - Threats to Seamount Ecosystems and Their Management, Editor(s): Charles Sheppard, *World Seas: an Environmental Evaluation (Second Edition)*, Academic Press, 2019, Pages 427-451, ISBN 9780128050521, <https://doi.org/10.1016/B978-0-12-805052-1.00018-8>

Rogers, A.D. (1994) The biology of seamounts. *Advances in Marine Biology*, 30, 304–360. www.researchgate.net/publication/279523256

Rowden, A., and M. Clark. (2004) Marine Biodiversity uncovering secrets of our seamounts. *Water and Atmosphere* 12(3). NIWA [wa12-3-rev27sept.indd \(niwa.co.nz\)](http://www.niwa.co.nz/wa12-3-rev27sept.indd)

Rowden, A.A.; Oliver, M.; Clark, M.R.; Mackay, K. (2008). New Zealand's "SEAMOUNT" database: recent updates and its potential use for ecological risk assessment. *New Zealand Aquatic Environment and Biodiversity Report No. 27*. 49 p. Seamount spreadsheet provided by email.

Sabine, C., Feely, R., Gruber, N., Key, R., Lee, K., Bullister, J., Wannankhof, R., Wong, C., & D. Wallace. (2004). The Oceanic sink for anthropogenic CO₂. *Science* 305: 5682. 376-371 <https://doi.org/10.1126/science.1097403>

Sala, E., Mayorga, J., Bradley, D., Cabral, R. B., Atwood, T. B., Auber, A., Cheung, W., Costello, C., Ferretti, F., Friedlander, A.M., Gaines, S.D., Garilao, C., Goodell, W., Halpern, B.S., Hinson, A., Kaschner, K., Kesner-Reyes, K., Leprieur, F., McGowan, J., Morgan, L. E., Mouillot, D., PalaciosAbrantes, J., Possingham, H. P., Rechberger, K. D., Worm B., and J. Lubchenco (2021) Protecting the global ocean for biodiversity, food and climate. *Nature* 592 <https://www.nature.com/articles/s41586-021-03371-z>

Schlacher, T. A., Baco, A. R., Rowden, A. A., O'Hara, T. D., Clark, M. R., Kelley, C., et al. (2014). Seamount benthos in a cobalt-rich crust region of the central Pacific: conservation challenges for future seabed mining. *Diver Distribution*. 20, 491–502. <https://doi.org/10.1111/ddi.12142>

Schnabel, K. E., Mills, V.S., Tracey, D.M., Macpherson, D., Kelly, M., Peart, R.A., Maggs, J.Q., Yeoman, J., & C.R. Wood. (2021). Identification of benthic invertebrate samples from research trawls and observer trips, 2020-21. *New Zealand Aquatic Environment and Biodiversity Report No. 269* [Identification of benthic invertebrate samples from research trawls and observer trips 2020–21. \(fish.govt.nz\)](http://www.fish.govt.nz/identification-of-benthic-invertebrate-samples-from-research-trawls-and-observer-trips-2020-21)

Staudigel, H.; A. Koppers, J. Lavelle, T. Pitcher and T. Shank (2010) Defining the word “seamount”. *Oceanography* 23: 20-21. www.researchgate.net/publication/274135460

Sweetman, A.K., Thurber, A.R., Smith, C.R., Levin, L.A., Mora, C., Wei, C., Gooday, A.J., Jones, D.O.B., Rex, M., Yasuhara, M., Ingels, J., Ruhl, H.A., Frieder, C.A., Danovaro, R., Wurzberg, L., Baco, A., Grupe, A. M., Pasulka, A., Meyer, K.S., Dunlop, K.M., Henry, L., & J.M. Roberts. (2017). Major impacts of climate change on deep-sea benthic ecosystems. *Elementa Science of the Anthropocene*. 5(4). DOI: <https://doi.org/10.1525/elementa.203>

Thiagarajan, N., Gerlach, D., Roberts, M., Burke, A., McNichol, A., Jenkins, W., Subhas, A., Thresher, R., & J. Adkins. (2013). Movement of deep-seas coral populations on climatic timescales. *Paleoceanography and Paleoclimatology* 28 (2) 227 – 236. <http://dx.doi.org/10.1002/palo.20023>

Tracey & Hjørvarsdóttir, 2019 - Appendix 4: Workshop on research needs for protected corals in New Zealand waters. Workshop minutes and Gaps identification. <https://www.doc.govt.nz/contentassets/1b230eee4e214f0da8ed6298d0c95add/doc-coral-workshop-minutes-and-gaps-nov-2017-final.pdf> and <https://www.doc.govt.nz/contentassets/1b230eee4e214f0da8ed6298d0c95add/doc-coral-workshop-gaps-final.pdf>.

Tracey, D., Braid, S., Sanders, B., & M. Smith. (2011). Distribution of protected corals in relation to fishing effort and assessment of accuracy of observer identification. NIWA report prepared for Marine Conservation Services, Department of Conservation [MCSINT201003-DOC11302X \(deepwatergroup.org\)](https://www.doc.govt.nz/contentassets/1b230eee4e214f0da8ed6298d0c95add/doc-coral-workshop-gaps-final.pdf)

Tracey, D., Stewart, R., Bilewitch, J., Macpherson, D., Chin, C., Mills, S., Neill, K., Kessel, G., Aguirre, D., & L. Liggins. (2019). Key Coral Groups, Chapter 12 In: Tracey, D.M. & Hjørvarsdóttir, F. (2019). The State of Knowledge of Deep-Sea Corals in the New Zealand Region. NIWA Science and Technology Series Number 84.62) <https://niwa.co.nz/sites/niwa.co.nz/files/Deepsea-corals-NZ-2019-NIWA-SciTechSeries-84.pdf>

Tracey, D.M. & Hjørvarsdóttir, F. (eds, comps) (2019). The State of Knowledge of Deep-Sea Corals in the New Zealand Region. NIWA Science and Technology Series Number 84. 140 p. <https://niwa.co.nz/sites/niwa.co.nz/files/Deepsea-corals-NZ-2019-NIWA-SciTechSeries-84.pdf>

Tremley-Boyer, L., & E. Abraham. (2020). Increased fisher-reporting of seabird captures during an electronic trial. New Zealand Aquatic Environment and Biodiversity Report No 238. Fisheries New Zealand. [AEBR 238 Increased fisher-reporting of seabird captures during an electronic-monitoring trial \(mpi.govt.nz\)](https://www.mpi.govt.nz/~/media/1/2/3/8/238-Increased-fisher-reporting-of-seabird-captures-during-an-electronic-monitoring-trial.pdf)

UNESCO. 2015. First Global Integrated Marine Assessment. Retrieved from: <https://www.unenvironment.org/resources/report/first-global-integrated-marine-assessment-worldocean-assessment-i>.

Watling L and Auster PJ (2021) Vulnerable Marine Ecosystems, Communities, and Indicator Species: Confusing Concepts for Conservation of Seamounts. *Front. Mar. Sci.* 8:622586. <https://doi.org/10.3389/fmars.2021.622586>

Watling, L. and Auster, P. J. (2017) Seamounts on the High Seas Should Be Managed as Vulnerable Marine Ecosystems. *Frontiers in Marine Science*. Vol 4 www.frontiersin.org/articles/10.3389/fmars.2017.00014

Watson, A.J., Schuster, U., Shutler, J.D., Holding, T., Ashton, I.G., Landschutzer, P., Woolf, D.K., and L. Goddihn-Murphy (2020) Revised estimates of ocean-atmosphere CO₂ flux are consistent with ocean carbon inventory. *Nature Communications* 11. [Revised estimates of ocean-atmosphere CO₂ flux are consistent with ocean carbon inventory | Nature Communications](https://www.nature.com/articles/s41467-020-18888-4)

Weaver, S. (2020) Conservation Services Programme Final Annual Research Summary 2018 - 2019. Department of Conservation, Wellington, New Zealand. February 2020. <https://www.doc.govt.nz/globalassets/documents/conservation/marine-and-coastal/marineconservation-services/reports/final-reports/final-csp-annual-research-summary-2018-19.pdf>

Williams, A., Osterhage, D., Althaus, F., Tyan, T., Green, M., & J. Pogonoski. (2021). A very large spawning aggregation of a deep-sea eel: magnitude and status. *Journal of Marine Science and Engineering*. 2021; 9(7):723. <https://doi.org/10.3390/jmse9070723>

WWF-NZ. (2021). WWF Short Briefing: the ever increasing destruction of the sea floor in New Zealand / Aotearoa. [wwf short briefing the ever increasing destruction of the sea floor.pdf \(panda.org\)](https://www.wwf.org.nz/~/media/1/2/3/8/238-Increased-fisher-reporting-of-seabird-captures-during-an-electronic-monitoring-trial.pdf)